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aPL IMMUNOREACTIVE PEPTIDES,
CONJUGATES THEREOF AND METHODS OF TREATMENT
FOR aPL ANTIBODY-MEDIATED PATHOLOGIES

COPY

Cross-Reference Applications

This application is a continuation-in-part application of U.S. Serial No. 08/660,092, filed June 6, 1996, which is a continuation-in-part of U.S. Serial No. 08/482,651, filed June 7, 1995.

Technical Field

This invention is in the field of immunology and relates to compositions and methods for treating and diagnosing antiphospholipid (aPL) antibody-mediated pathologies. More specifically, the invention relates to conjugates of chemically-defined nonimmunogenic valency platform molecules and immunospecific analogs of aPL-binding epitopes as well as methods and compositions for producing these conjugates. Optimized analogs lack T cell epitopes. In addition, the invention relates to diagnostic assays for detecting the presence of and quantitating the amount of antiphospholipid antibodies in a biological sample. The invention also relates to a method of utilizing random peptide libraries to identify immunospecific analogs of aPL-binding epitopes.

Background of the Invention

Antiphospholipid antibodies occur in autoimmune diseases such as systemic lupus erythematosus (SLE) and antiphospholipid antibody syndrome (APS) as well as in association with infections and drug therapy. APS is characterized by one or more clinical features such as arterial or venous thrombosis, thrombocytopenia and fetal loss. APS may be primary or it may be associated with other conditions, primarily

SLE. (PHOSPHOLIPID-BINDING ANTIBODIES (Harris *et al.*, eds., CRC Press, Boca Raton, FL, 1991); McNeil *et al.* ADVANCES IN IMMUNOLOGY, Vol. 49, pp. 193-281 (Austen *et al.*, eds., Academic Press, San Diego, CA, 1991)). Approximately 30-40% of patients with SLE have aPL, however, 50% of patients with aPL antibodies do not have SLE. This 50% may have other autoimmune rheumatic diseases, miscellaneous conditions or they may have been subjected to drug therapy, particularly chlorpromazine. In one study of 70 patients, 26 males and 44 females, with primary APS (PAPS) but no evidence of SLE, the following features were observed: deep venous thrombosis (DVT) in 31; arterial occlusion in 31, particularly stroke or transient ischemia; myocardial infarctions in 15; recurrent fetal loss in 24; thrombocytopenia (TCP) in 32; 10 had a positive Coombs' test; Evans' syndrome in 7; anti-nuclear antibody (ANA) in 32, but less than 1:160 in 29; and antimitochondrial antibody (AMA) in approximately 24. (McNeil *et al.*, *supra*). Estimates vary but in about 5% of all stroke patients, aPL antibodies are thought to be an important contributing factor.

Transient aPL antibodies, such as those detected in a VDRL test, occur during many infections. Approximately 30% of patients possessing persistent aPL antibodies have suffered a thrombotic event. The presence of aPL antibodies defines a group of patients within SLE who display a syndrome of clinical features consisting of one or more of thrombosis, TCP, and fetal loss. The risk of this syndrome in SLE overall is around 25%; this risk increases to 40% in the presence of aPL antibodies and decreases to 15% in their absence. Because aPL antibodies were thought to be directed at phospholipids in plasma membranes, it has been postulated that they may exert direct pathogenic effects *in vivo* by interfering with hemostatic processes that take place on the phospholipid membranes of cells such as platelets or endothelium. In patients with PAPS, the fact that aPL antibodies appear to be the only risk factor present is further evidence that these antibodies have a direct pathogenic role. Induction of PAPS by immunizing mice with human anticardiolipin antibodies is the best evidence yet that aPL antibodies are directly pathogenic (Bakimer *et al.* 1992 *J. Clin. Invest.* 89:1558-1563; Blank *et al.* 1991 *Proc. Natl. Acad. Sci.* 88:3069-3073).

Measurement of aPL antibodies in the clinical environment is still an imperfect art. A commercially available set of standard antisera (APL Diagnostics,

Inc., Louisville, KY) allow generation of a standard curve for comparison of assays performed in various laboratories. A great deal of inconsistency exists, however, between the results obtained at these laboratories regarding the exact GPL and MPL, the unit of measurement for IgG and IgM antiphospholipid antibodies, respectively, ratings for given sera and the levels of GPL and MPL that are categorized as high, medium or low titer. The available commercial kits vary greatly in the values assigned to the commercially available standards (Reber *et al.* (1995) *Thrombosis and Haemostat.* 73:444-452). In spite of these limitations, there is general agreement that the epitopes recognized by antibodies in APS, PAPS and other aPL antibody-mediated diseases including recurrent stroke and recurrent fetal loss are located in the 5th domain of β_2 -GPI and are exposed to the antibody following binding of β_2 -GPI to cardiolipin.

It is now generally accepted that aPL antibodies recognize an antigenic complex comprised of β_2 -glycoprotein I (β_2 -GPI) and negatively-charged phospholipid, e.g., cardiolipin (McNeil *et al.* (1990) *Proc. Natl. Acad. Sci.* 87:4120-4124; Galli *et al.* (1990) *Lancet* I:1544-1547) (hereinafter "aPL immunogen"). β_2 -GPI is a minor plasma glycoprotein found free and in association with lipoprotein lipids where it is also known as apolipoprotein H (apo H). It consists of five independently folding domains referred to as Sushi or short consensus repeat domains that resemble similar domains in other proteins. β_2 -GPI has been reported to undergo antigenic and conformational changes upon binding phospholipid (Wagenknecht *et al.* (1993) *Thromb. Haemostas.* 69:361-365; Jones *et al.* (1992) *Proc. 5th Intl. Symp. Antiphospholipid Antibodies* (Abstract S5)). The fifth domain contains the putative sites of lipid binding and aPL antibody binding (Hunt J. and S. Krilis, (1994) *J. Immunol.* 152:653-659; Lauer *et al.* (1993) *Immunol.* 80:22-28). The pathological mechanism for aPL is unknown (McNeil *et al.*, *supra*). Most explanations invoke endothelial cell function or platelet involvement (Haselaar *et al.* (1990) *Thromb. Haemostas.* 63:169-173). These explanations suggest that following blood vessel endothelial cell injury or platelet activation, the exposure or transbilayer migration of anionic phospholipid to the plasma-exposed surface may lead to β_2 -GPI-binding and trigger aPL antibody formation.

aPL antibodies may be directly prothrombotic by reducing prostacyclin formation (Vermylen, J. and J. Arnout (1992) *J. Clin. Lab. Med.* **120**:10-12); by direct interference with the action of coagulation proteins; or by blocking the ability of β_2 -GPI to inhibit the intrinsic blood coagulation pathway, platelet prothrombinase activity, and ADP-mediated platelet aggregation (Arvieux *et al.* (1993) *Thromb. Haemostas.* **60**:336-341).

A major new tool in medicinal chemistry in the search for lead compounds has been the advent of combinatorial libraries providing vast molecular diversity. Molecular diversity may arise from chemical synthesis or from biological systems (Scott., J.K. RATIONAL DRUG DESIGN (CRC Press, Weiner, D. B. and W.V. Williams, eds., Boca, Raton, FL., 1994); Moos *et al.* (1993) *Ann. Reports Med. Chem.* **28**:315-324). By displaying random peptides on the surface of filamentous phage, epitope libraries containing hundreds of millions of clones for probing by clinically significant antibodies have been created (Scott, J.K. and G.P. Smith (1990) *Science* **249**:286-390; Cesareni, G. (1992) *FEBS Lett.* **307**:66-70). Such phage libraries are prepared by incorporating randomized oligonucleotide sequences into the phage genome, usually the pIII gene, which encode unique peptide sequences on the surface of each phage. Following sequential rounds of affinity purification and amplification, those phage that bind antibody are propagated in *E. coli* and the binding peptides identified by sequencing the corresponding coding region of viral DNA. In most cases, subsequent study will involve corresponding synthetic peptides after establishing their ability to bind antibody. Phage-based libraries have been used to mimic discontinuous epitopes (Luzzago *et al.* (1993) *Gene* **128**:51-57; BaLass *et al.* (1993) *Proc. Natl. Acad. Sci.* **90**:10638-10642). The potential plasma instability of peptide-based drugs has been successfully overcome by *N*-terminal blocking or by the judicious use of amino acid analogs (Powell, M.F. (1993) *Ann. Reports Med. Chem.* **28**:285-293).

At present there is no selective, immunospecific therapy for patients showing high titers of aPL antibodies. In many cases use of drugs such as aspirin, steroids, and warfarin has proven to be largely inadequate (PHOSPHOLIPID-BINDING ANTIBODIES (Harris *et al.*, eds., CRC Press, Boca Raton, FL, 1991); McNeil *et al.*, *supra*). Synthetic mimetic peptides, characterized by (i) the inability to activate T cells while (ii) retaining the ability to bind immune B cells, are used to tolerize B cells in an

antigen-specific manner. This technology is disclosed in co-owned, co-pending U.S. Patent Application, Serial No. 08/118,055, filed September 8, 1993, and U.S. Patent No. 5,268,454, which are incorporated by reference herein in their entirety. As disclosed in the application and patent cited above, B cell tolerance entails administering such peptides conjugated to multivalent, stable, non-immunogenic valency platforms in order to abrogate antibody production via B cell anergy or clonal deletion after cross-linking surface immunoglobulin.

Although the exact molecular nature of the target epitopes recognized by aPL antibodies is unknown, the use of peptides derived from epitope libraries will allow for the construction of successful tolerogens. B cell tolerogens for the treatment of human systemic lupus erythematosus-related nephritis have also been disclosed in co-owned U.S. Patent Nos. 5,276,013 and 5,162,515 which are incorporated by reference herein in their entirety.

Disclosure of the Invention

This invention resides in the discovery of a method for identifying analogs of key epitopes recognized by aPL antibodies in patients suffering from PAPS, APS and other aPL antibody-mediated diseases, such as recurrent stroke and recurrent fetal loss, using random peptide phage libraries.

Accordingly, one aspect of the invention is an improved method for screening random peptide phage libraries in order to identify the peptide sequences which best mimic the epitopes recognized by aPL antibodies. This method comprises the steps of: (a) biopanning the library using methods modified from those known in the art; (b) eliminating very weakly-binding phage by micropanning the phage screened from step (a) by (i) incubating the phage in microplate wells coated with aPL antibody bound to Protein G, (ii) washing the microplate wells to remove unbound phage, (iii) eluting the bound phage, and (iv) infecting a microorganism such as *E. coli* with the eluted phage and counting the number of infected microorganisms by plating on agar; (c) determining the strongest-binding clones recovered in (b) by evaluation via phage-capture ELISA by (i) coating the wells of a microplate with aPL antibody, (ii) incubating the strongest-binding clones identified by micropanning in (b) in the coated wells and washing away unbound phage, (iii) quantitating the number of phage

bound to the antibody using an enzyme-conjugated goat anti-phage antibody in a colorimetric ELISA assay and, if several equivalent strongly-binding clones are identified, an additional round of (d) phage-ELISA on the strongest-binding phage-capture-ELISA clone.

In this regard, the invention encompasses a method for identifying analogs of epitopes which specifically bind aPL antibodies isolated from humans suffering from an aPL antibody-mediated disease comprising: (a) preparing phage random peptide libraries; (b) screening said libraries with aPL antibodies to identify aPL mimetic epitopes, wherein said screening comprises (i) screening said libraries by biopanning; (ii) further screening phage isolated by biopanning in (i) by micropanning; and (iii) identifying phage containing aPL antibody high-affinity binding peptides recovered in (ii) by immunoassay.

The invention also encompasses a method of biopanning phage random peptide libraries to identify and isolate peptides which bind to aPL antibody comprising: (a) reacting affinity-purified aPL antibody with phage bearing random peptide inserts; (b) recovering phage bearing random peptide inserts which bind to the aPL antibody; (c) infecting a microorganism with phage recovered in (b); and (d) culturing the infected microorganism in an antibiotic-containing medium in order to isolate the phage.

The invention further encompasses a method of micropanning phage random peptide libraries to identify and isolate peptides having a high binding affinity to aPL antibodies comprising: (a) isolating phage bearing random peptide inserts by biopanning; (b) incubating the phage recovered in step (a) in microplate wells coated with aPL antibody bound to Protein G; (c) washing the microplate wells to remove unbound phage; (d) eluting bound phage; and (e) infecting a microorganism with phage recovered in (d); and (f) culturing the infected microorganism in an antibiotic-containing medium in order to isolate the phage.

The invention also encompasses the above method described wherein the immunoassay is a phage-capture ELISA comprising: (a) incubating phage bearing random peptide inserts isolated by micropanning in the microplate wells coated with aPL antibody; (b) washing away unbound phage; (c) incubating an enzyme-labeled anti-phage antibody to the wells; (d) washing away unbound enzyme-labeled anti-phage antibody; (e) adding a colorimetric substrate; and (f) measuring the absorbance of the substrate to identify high affinity-binding phage.

Also encompassed by the invention is the method described above and further comprising performing an additional phage-capture ELISA assay of the high affinity-binding phage comprising: (a) coating a uniform amount of the phage on microplate wells; (b) incubating aPL antibody in the wells; (c) washing away unbound antibody; (e) incubating an enzyme-labeled anti-aPL antibody with the bound aPL antibody; (f) washing away unbound enzyme-labeled anti-aPL antibody; (g) adding a colorimetric substrate to the wells; and (h) measuring the absorbance of the substrate to measure the relative binding affinity of the phage.

The invention also encompasses the method described above wherein the immunoassay is a colony-blot immunoassay comprising: (a) culturing a microorganism infected with phage bearing random peptide inserts on a nitrocellulose membrane atop an agar-containing culture medium; (b) replicate transferring the microorganism cultured in (a) by blotting the microorganism on a second nitrocellulose membrane atop an agar-containing culture medium; (c) incubating the transferred microorganism; (d) lysing the microorganism; (e) digesting the microorganism with lysozyme; (f) blocking the membrane with a gelatin solution; (g) incubating the membrane with aPL antibody; (h) washing away unbound aPL antibody; (i) incubating a enzyme-labeled anti-aPL antibody with the nitrocellulose membrane; (j) washing away unbound enzyme-labeled anti-aPL antibody; (k) adding a colorimetric substrate; and (l) measuring the absorbance of the substrate to identify high affinity-binding phage.

A method for assaying and ranking, for affinity-binding characteristics, epitopes which specifically bind aPL antibodies isolated from humans suffering from an aPL antibody-mediated disease is also encompassed, the method comprising: (a) coating wells of a microtitration plate with cardiolipin; (b) adding adult bovine or human serum as a source of β_2 -GPI to bind to the cardiolipin and to prevent non-specific binding to the wells of the plate; (c) incubating a solution of monomeric analog and a high-titered aPL antibody for a pre-determined time; (d) adding the aPL antibody/analog mixture to wells of the microtitration plate and incubating for a pre-determined time; (e) washing the wells to wash away unbound aPL antibody; (f) adding anti-human IgG conjugated with a label (e.g., an enzyme) to the wells of the plate and incubating for a pre-determined time; (g) washing the wells to wash away unbound anti-human IgG conjugate; (h) adding a substrate for the labeled conjugate and developing the substrate/label reaction for a pre-

determined time;(i) measuring the end-product of the substrate/label reaction to quantitate the amount of aPL antibody bound to the well; (j) calculating the percentage inhibition, if any, of binding of the aPL antibody to determine the affinity of the analog to the aPL antibody.

Another aspect of the invention encompasses a fluorescence polarization peptide binding assay for determining the dissociation constants for peptides that bind to aPL antibodies. This assay detects direct binding of peptides to aPL antibodies.

The invention also encompasses a diagnostic immunoassay for determining the presence of aPL antibody in body fluids taken from subjects suspected of suffering from an aPL antibody-mediated disease comprising contacting a sample of a body fluid with an analog of an epitope which specifically binds aPL antibodies and determining by methods well known in the art whether aPL antibodies are present in the body fluid and, if present, quantitating the amount of aPL antibodies present in the fluid. One such immunoassay comprises: (a) coating wells of a microtitration plate with an analog of an epitope which specifically binds aPL antibodies; (b) washing the wells to wash away unbound analog; (c) adding a test sample of a body fluid to the wells and incubating for a pre-determined time; (d) washing the wells to remove unbound test sample; (e) adding anti-human IgG conjugated with a label to the wells of the plate and incubating for a pre-determined time;(f) washing the wells to wash away unbound anti-human IgG conjugate; (g) adding a substrate for the labeled conjugate and developing the substrate/label reaction for a pre-determined time; (h) measuring the end-product of the substrate/label reaction to determine the presence of anti-aPL antibody in the test sample. A diagnostic immunoassay as described above wherein the immunoassay is quantitative is also encompassed.

The phage-ELISA assay consists of (i) coating a uniform amount of different clones on wells of a microtitration plate followed by (ii) identifying the peptide inserts which most strongly bind aPL antibody by adding antibody to the wells and developing the reaction with an enzyme-labeled anti-human IgG conjugate. The random peptides displayed by the phage which have a high binding affinity to aPL antibody as measured by phage-ELISA, colony blot or phage-capture-ELISA represent the analogs of the aPL-specific epitope. These peptides are then synthesized and ranked for strength of binding using competition assays.

Another aspect of the invention is aPL antibody-binding analogs that bind specifically to B cells to which an aPL epitope binds. Optimized analogs lack T cell epitope(s).

Yet another aspect of the invention is a composition for inducing specific B cell tolerance to an aPL immunogen comprising a conjugate of a nonimmunogenic valency platform molecule and an aPL antibody-binding analog that (a) binds specifically to B cells to which an aPL immunogen binds and (b) lacks T cell epitope(s).

Brief Description of the Drawings

Figure 1 shows that the substitution of fish gelatin for adult bovine serum abolished all anticardiolipin (ACA) activity in an ELISA assay of a commercial aPL antibody standard. This result supported the findings of McNeil *et al.*, *supra*, and Galli *et al.*, *supra*, concerning the importance of β_2 -glycoprotein I (β_2 -GPI) in defining the target epitope(s) of ACA.

Figure 2 shows that resin-bound analog 5A12 immunospecifically binds to affinity-purified IgG designated ACA-6501.

Figure 3 illustrates that the aPL antibody-binding analogs derived from screening with ACA-6626 bound aPL antisera but did not bind normal sera.

Figure 4 also illustrates that the ACA-6501/5A12 analog immunospecifically binds ACA-6501 antiserum and is crossreactive with ACA-6626.

Figures 4 and 5 illustrate that while the ACA-6501/5A12 and ACA-6626/4D3 aPL antibody-binding analogs derived from screening with methods within the instant invention bind preferentially with the screening antibody, a significant degree of crossreactivity was detected.

Figure 6 illustrates method for calculating the GPL value for ACA-6501 aPL antibody.

Figure 7 shows the activity of affinity-isolated ACA-6501 compared to GPL standard sera.

Figure 8 illustrates the dramatic drop in sequence diversity of the isolated clones by the fourth round of biopanning.

Figure 9 illustrates that three clones (3A12, 3B3 and 3A5) exhibited a very strong immunospecific signal in the phage-capture ELISA using ACA-6635 whereas all clones tested were unreactive with normal IgG.

Figure 10 shows the strong signal exhibited by seven clones in a phage-ELISA using ACA-6501.

Figure 11 shows the results of a competitive-binding ELISA obtained with peptides 5A12, CB2 and 3B10 using ACA-6501. 0.16 µg of Peptide 5A12 produced 50% inhibition of binding of ACA-6501 aPL antibody to tetravalent peptide ACA-6501/3B10 bound to polystyrene microplate wells, whereas 0.08 µg of CB2 and 0.5 µg of 3B10 were required to produce 50% inhibition.

Figure 12 illustrates the comparative activity of modified ACA-6641/3G3 analogs.

Figure 13 illustrates that the 50% inhibition values for peptides 139, 142 and 143 in a competitive-binding ELISA using ACA-6501 aPL antibody were 6.9, 0.7 and 0.9 µg, respectively.

Figure 14 illustrates the effects of substituting α-Me-Pro at positions 3, 9 and both 3 and 9 in peptide 3B10. Substitution of α-Me-Pro at position 9 increased activity of the peptide four-fold compared to the "native" peptide.

Figure 15 shows that peptide 6641/3G3 (LJP 688) is highly cross-reactive with nine affinity-purified ACA antibodies.

Figures 16 and 17 show the dose-dependent reduction in anti-685 antibody ABC at 10^8 M using spleen cells from mice immunized with LJP 685-KLH after the spleen cells were incubated with 100, 20 and 4 µM of LJP 685-MTU-DABA-PEG conjugate (compound 36 and LJP 685-ATU-MTU-AHAB-TEG conjugate (compound 35), respectively, for 2 hours.

Figure 18 displays the NMR structure closest to the centroid of the nine structures elucidated for peptide 925 and is a reasonable representation of the shape of the peptide 925 molecule.

Figure 19 compares the structure of peptide 925 (labeled at the bottom of the figure as 3G3) with the structure of peptide 5A12. Both peptides have turns at approximately the same positions in the peptide sequence.

Figures 20A and 20 B illustrate that the pharmacophore of aPL analogs has been tentatively identified as a small hydrophobic group and a positively charged group. The gem-dimethyl and amino groups of peptide 925 are tentatively identified as the pharmacophore of this peptide as shown in Figure 20A. The lengths of the hydrocarbon linkers that tether the pharmacophore groups to some scaffold are specified in Figure 20A as well as the distances separating the points at which these linkers are attached to the scaffold.

Figure 21 illustrates inhibition of β_2 GPI by 6501-derived peptides with diluted 6501 serum.

Figure 22 shows the ACA-6701 titration of CB2*-F; FITC-GPCILLARDRCG.

Figure 23 shows the ACA-6501 titration of CB2*-F: FITC-GPCILLARDRCG.

Figure 24 shows complete ACA-6501 titration of CB2*-F: FITC-GPCILLARDRCG.

Figure 25 shows the displacement of CB2*-F from ACA-6701 using 1.04 equivalents of CB2*.

Figure 26 shows cFP titration using CB2* to displace CB2*-F from ACA-6701.

Figure 27 shows cFP titration using 3B10 to displace CB2*-F from ACA-6701.

Figure 28 shows the dose response of the $(LJP685)_4/MTU-AHAB-TEG$ conjugate for tolerance activity.

Figure 29 shows the dose response of the $(LJP685)_4/MTU-DABA-TEG$ conjugate for tolerance activity.

Figure 30 shows the dose response of tolerance activity of the $(LJP685)_4/MTU-AHAB-TEG$ conjugate tested in the *in vitro* model.

Figure 31 shows the dose response of tolerance activity of the $(LJP685)_4/MTU-DABA-TEG$ conjugate tested in the *in vitro* model.

Figure 32 shows the tolerizing effect of the $(LJP685)_4/MTU-AHAB-TEG$ conjugate comparing various administrative routes and dosage ranges.

Modes for Carrying Out the Invention

A. Definitions

As used herein the term "aPL antibody" means any antibody which specifically binds β 2-GPI that mediates disease.

As used herein the term "B cell anergy" intends unresponsiveness of those B cells requiring T cell help to produce and secrete antibody and includes, without limitation, clonal deletion of immature and/or mature B cells and/or the inability of B cells to produce antibody.

"Unresponsiveness" means a therapeutically effective reduction in the humoral response to an immunogen. Quantitatively the reduction (as measured by reduction in antibody production) is at least 50%, preferably at least 75%, and most preferably 100%.

"Antibody" means those antibodies which are T cell dependent.

As used herein the term "immunogen" means an entity that elicits a humoral immune response comprising aPL antibodies. Immunogens have both B cell epitopes and T cell epitopes. aPL immunogens that are involved in aPL antibody-mediated pathologies may be external (foreign to the individual) immunogens such as drugs, including native biological substances foreign to the individual such as therapeutic proteins, peptides and antibodies, and the like or self-immunogens (autoimmunogens) such as those associated with antibody-mediated hypercoagulability (stroke).

The term "analog" of an immunogen intends a molecule that (a) binds specifically to an antibody to which the immunogen binds specifically and (b) lacks T cell epitopes. Although the analog will normally be a fragment or derivative of the immunogen and thus be of the same chemical class as the immunogen (*e.g.*, the immunogen is a polypeptide and the analog is a polypeptide), chemical similarity is not essential. Accordingly, the analog may be of a different chemical class than the immunogen (*e.g.*, the immunogen is a carbohydrate and the analog is a polypeptide) as long as it has the functional characteristics (a) and (b) above. The analog may be a peptide, carbohydrate, lipid, lipopolysaccharide, nucleic acid or other biochemical entity. Further, the chemical structure of neither the immunogen nor the analog need be defined for the purposes of this invention.

The term "analog" of an immunogen also encompasses the term "mimotope." The term "mimotope" intends a molecule which competitively inhibits the antibody from binding the immunogen. Because it specifically binds the antibody, the mimotope is considered to mimic the antigenic determinants of the immunogen.

As used herein "valency platform molecule" means a nonimmunogenic molecule containing sites which facilitate the attachment of a discreet number of analogs of immunogens.

"Nonimmunogenic" is used to describe the valency platform molecule and means that the valency platform molecule elicits substantially no immune response when it is administered by itself to an individual.

As used herein "individual" denotes a member of the mammalian species and includes humans, primates, mice and domestic animals such as cattle and sheep, sports animals such as horses, and pets such as dogs and cats.

As used herein "pharmacophore" means the three dimensional orientation and chemical properties of key groups involved in binding of an aPL analog to the antibody target.

B. Identification of aPL Antibody-binding Analogs

aPL antibody-binding analogs may be identified by screening candidate molecules to determine whether or not they (a) bind specifically to aPL antibodies and (b) lack T cell epitopes. Specific binding to aPL antibodies may be determined using conventional immunoassays such as the ELISA assays described in the examples below and the presence or absence of T cell epitopes may be determined by conventional T cell activation assays also described in the examples. In this regard, an analog which "binds specifically" to serum antibodies to the immunogen exhibits a reasonable affinity thereto, e.g., $10^7 M^{-1}$. The presence or absence of T cell epitopes may be determined using a tritiated thymidine incorporation assay disclosed in Serial No. 08/118,055. The presence of T cell epitopes can also be determined by measuring secretion of T cell-derived lymphokines by methods well known in the art. Analogs that fail to induce statistically significant incorporation of thymidine above background are deemed to lack T cell epitopes. It will be appreciated that the quantitative amount of thymidine incorporation may vary with the immunogen.

Typically a stimulation index below about 2-3, more usually about 1-2, is indicative of a lack of T cell epitopes.

C. Preparation of Conjugates

The aPL antibody-binding analogs are coupled to a nonimmunogenic valency platform molecule to prepare the conjugates of the invention. Preferred valency platform molecules are biologically stabilized, *i.e.*, they exhibit an *in vivo* excretion half-life often of hours to days to months to confer therapeutic efficacy, and are preferably composed of a synthetic single chain of defined composition. They will normally have a molecular weight in the range of about 200 to about 200,000, usually about 200 to about 20,000. Examples of valency platform molecules within the present invention are polymers such as polyethylene glycol (PEG), poly-D-lysine, polyvinyl alcohol and polyvinylpyrrolidone. Preferred polymers are based on polyethylene glycols (PEGs) having a molecular weight of about 200 to about 8,000.

Other valency platform molecules suitable for use within the present invention are the chemically-defined, non-polymeric valency platform molecules disclosed in co-owned, co-pending U.S. patent application Serial No. 08/152,506, filed November 15, 1993, which is incorporated by reference herein in its entirety. Particularly preferred homogeneous chemically-defined valency platform molecules suitable for use within the present invention are derivatized 2,2'-ethylenedioxydiethylamine (EDDA) and triethylene glycol (TEG).

Additional suitable valency platform molecules include tetraaminobenzene, heptaaminobetacyclodextrin, tetraaminopentaerythritol, 1,4,8,11-tetraazacyclotetradecane (Cyclam) and 1,4,7,10-tetraazacyclododecane (Cyclen).

Conjugation of the aPL antibody-binding analog to the valency platform molecule may be effected in any number of ways, typically involving one or more crosslinking agents and functional groups on the analog and valency platform molecule.

Polypeptide analogs will contain amino acid sidechain moieties containing functional groups such as amino, carboxyl, or sulfhydryl groups that will serve as sites for coupling the analog to the carrier. Residues that have such functional groups may be added to the analog if the analog does not already contain these groups. Such

residues may be incorporated by solid phase synthesis techniques or recombinant techniques, both of which are well known in the peptide synthesis arts. In the case of carbohydrate or lipid analogs, functional amino and sulphydryl groups may be incorporated therein by conventional chemistry. For instance, primary amino groups may be incorporated by reaction with ethylenediamine in the presence of sodium cyanoborohydride and sulphydryls may be introduced by reaction of cysteamine dihydrochloride followed by reduction with a standard disulfide reducing agent. In a similar fashion, the valency platform molecule may also be derivatized to contain functional groups if it does not already possess appropriate functional groups.

Hydrophilic linkers of variable lengths are useful for connecting peptides or other bioactive molecules to valency platform molecules. Suitable linkers include linear oligomers or polymers of ethylene glycol. Such linkers include linkers with the formula $R^1S(CH_2CH_2O)_nCH_2CH_2O(CH_2)_mCO_2R^2$ wherein n = 0-200, m = 1 or 2, R¹ = H or a protecting group such as trityl, R² = H or alkyl or aryl, e.g., 4-nitrophenyl ester. These linkers are useful in connecting a molecule containing a thiol reactive group such as haloacyl, maleimide, etc., via a thioether to a second molecule which contains an amino group via an amide bond. These linkers are flexible with regard to the order of attachment, i.e., the thioether can be formed first or last.

The conjugates will normally be formulated for administration by injection (e.g., intraperitoneally, intravenously, subcutaneously, intramuscularly, etc.). Accordingly, they will typically be combined with pharmaceutically acceptable vehicles such as saline, Ringer's solution, dextrose solution, and the like. The conjugate will normally constitute about 0.01% to 10% by weight of the formulation. The conjugate is administered to an individual in a "therapeutically effective amount", i.e., an amount sufficient to produce B cell anergy to the involved immunogen and effect prophylaxis, improvement or elimination of the antibody-mediated condition being addressed. The particular dosage regimen, i.e., dose, timing and repetition, will depend on the particular individual and that individual's medical history. Normally, a dose of about 1 μ g to about 100 mg conjugate/kg body weight, preferably about 100 μ g to about 10 mg/kg body weight, will be given weekly. Other appropriate dosing schedules would be as frequent as daily or 3 doses per week, or one dose per week, or one dose every two to four weeks, or one dose on a monthly or less frequent schedule.

depending on the individual or the disease state. Repetitive administrations, normally timed according to B cell turnover rates, may be required to achieve and/or maintain a state of humoral anergy. Such repetitive administrations will typically involve treatments of about 1 µg to about 10 mg/kg body weight or higher every 30 to 60 days, or sooner, if an increase in antibody titer is detected. Alternatively, sustained continuous release formulations of the conjugates may be indicated for some pathologies. Various formulations and devices for achieving sustained release are known in the art.

Anti-helper T cell treatments may be administered together with the conjugates. Such treatments usually employ agents that suppress T cells such as steroids or cyclosporin.

D. Nature of the antigen for aPL antibodies

Initial studies on aPL antibodies by the inventors coincided with the publication of reports questioning the nature of the antigenic site recognized by these antibodies. Initially these antibodies were thought to recognize the cardiolipin molecule much like those antibodies detected in a VDRL test. As shown in Figure 1, substitution of fish gelatin for adult bovine serum in the anticardiolipin antibody (ACA) solid phase ELISA essentially abolished all ACA activity of an antibody preparation obtained commercially as an ACA standard. This finding indicated the ACA antibodies recognized a determinant on a serum protein as proposed by McNeil *et al.*, *supra* and Galli *et al.*, *supra*, rather than cardiolipin itself. This protein was shown by these authors to be β_2 -GPI and, thus, the terms "ACA" or "anti-cardiolipin antibodies" are really misnomers but are still used today to refer to these antibodies.

The majority of autoimmune, IgG aPL antibodies recognize determinants on the β_2 -GPI molecule. These epitopes may be formed or exposed on β_2 -GPI only upon its binding to cardiolipin (a neo epitope). Alternatively, the epitope on β_2 -GPI may exist in a single copy per β_2 -GPI molecule and may have low affinity for aPL antibodies. Sufficient avidity to maintain an antibody-antigen interaction might be reached only with the alignment of two of these sites on adjacent β_2 -GPI molecules by their binding to cardiolipin.

Additional insight into the β_2 -GPI epitope structure has been gained by nuclear magnetic resonance (NMR) analysis of aPL analogs within the present invention which mimic the native epitope(s) of β_2 -GPI. For, example, a comparison of the NMR solution structure of two peptide aPL analogs that are highly cross-reactive with aPL antibodies showed that both peptides have turns at approximately the same positions in the peptide sequence (see Figures 18 and 19).

E. ACA ELISA

The necessity of large-scale testing in connection with GPL scoring of clinical samples and chromatographic purifications of research antibodies and β_2 -GPI led to the development of an ELISA assay for aPL antibodies with performance in close agreement with a commercial kit and similar to the design found to have the best reproducibility (Reber *et al.*, *supra*). A modified ACA ELISA has also been performed wherein β_2 -GPI bound directly to certain microplates is used to bind aPL IgG directly in the absence of any cardiolipin first added to the microplate wells. (See, Roubey *et al.* (1996) *Arthritis & Rheumatism* 39:1606-1607; R.A.S. Roubey (1996) *Arthritis & Rheumatism* 39:1444-1454).

F. Immunoaffinity Purification of aPL Antibodies

In order to isolate the aPL antibodies, multilamellar, cardiolipin-containing dispersions (liposomes; also containing cholesterol and dicetylphosphate) are incubated with aPL plasma (or serum). These liposomes are pelleted from the serum by centrifugation. After washing, the liposome mixture is disrupted by 2% octylglucoside detergent and applied to a protein A-agarose column. Following extensive washings to first remove lipids and then to remove non-IgG components, IgG aPL antibody is eluted from protein A with mild acid, neutralized, buffer-exchanged, and tested in the ACA ELISA. This procedure yields aPL antibody enriched up to 10,000-fold that is devoid of any contaminating β_2 -GPI as shown by western blotting with rabbit IgG anti-human β_2 -GPI antisera. An additional affinity-purification step is performed by chromatography of the affinity-purified antibody on solid phase β_2 -GPI. This second affinity-purification step is recommended as a result

of the new awareness regarding the greater clinical relevance of aPL antibodies that directly bind to β_2 -GPI. It also serves to further ensure a final preparation devoid of contaminants, in particular β_2 -GPI.

G. Construction of Filamentous Phage Random Peptide Libraries

Eleven different fUSE 5 filamentous phage random peptide libraries on the p-III protein (five copies of p-III with peptide per phage) are constructed. These libraries provide a vast array of shapes and structures for the discovery of mimetic epitopes. Four libraries, designated "x" libraries, have peptide inserts that are 8, 10, 12, and 15 residues in length, respectively, and are flanked by proline residues on both the amino and carboxyl ends. The purpose of these proline residues is to disrupt any contribution to secondary structure that might arise from the native p-III protein and to project the insert into the solvent. The "y'" libraries contain cysteine-bounded inserts that are 6, 7, 8, 9, 11, and 13 amino acids long. The "y" library is the same as the "y'" library except that it lacks the 6 and 8 amino acid inserts. These peptide inserts for both "y" and "y'" libraries are flanked by cysteine residues at both the amino and carboxyl ends to form cyclic, more rigid structures. Proline residues are incorporated outside these cysteine residues for reasons similar to those for the "x" libraries above. The "x," "y'", and "y" libraries are located five residues from the amino terminus of the native p-III protein. The "z" library consists of random eight amino acid inserts located at the amino terminus of the p-III protein and do not contain any flanking proline or cysteine residues. A combination of the "x," "y'" and "z" libraries represents eleven different libraries each with approximately one hundred million different peptide inserts.

These libraries are constructed by incorporation of random oligonucleotide sequences of the length appropriate to give the desired length insert into the p-III gene of fUSE 5 using standard molecular biology techniques. Following restriction endonuclease digestion of the fUSE 5 DNA, an excess of kinased oligonucleotides provided as gapped duplexes is added and ligated. The DNA is then electroporated into *E. coli* and inserts are selected by culturing in tetracycline-containing media. The phage from this culture (which contain the peptide insert) are isolated from the

supernatant, washed and resuspended in buffer. Typically libraries are shown to have 7×10^8 independent clones at 8×10^{12} transducing units per mL.

H. Phage-screening methodology

The essence of screening phage display peptide libraries lies in the ability to collapse billions of potential candidate phage to a relative few with outstanding properties. The original screening protocols recommended by Scott, J.K. and G.P. Smith, (1990) *Science* 249:315-324 are significantly modified to facilitate the selection of the best epitopes for various aPL antibodies. These procedures are designed to apply greater stringency of selection as the screen progressed until a point is reached where a useful number of clones representing the best sequences can be thoroughly investigated. With some antibodies, the library does not appear to have sequences which bind very tightly and if a method with a high degree of stringency is applied to the screen, no clones survive that are specific. On the other hand, the library frequently yields many clones that represent good analogs of the antigen and it is necessary to employ a method with a high degree of stringency to identify the best epitopes. For that reason, assays were developed with varying degrees of stringency in order to identify the best epitopes from an epitope library screen. The assays are listed here in order of increasing stringency: Biopanning < Micropanning < Phage-Capture ELISA < Phage ELISA = Colony Blot = Peptide ELISA.

(i) Biopanning

"Biopanning" describes the technique wherein affinity-purified aPL antibody and phage bearing random peptide inserts are allowed to mix, following which antibody-specific recovery captures the bound phage. The phage confer tetracycline resistance to *E. coli* that are propagated in a tetracycline-containing medium and then isolated. Multiple rounds of biopanning enrich the number of immunospecific phage in a sample. Phage are always recovered at the end of three to five rounds of selection but may represent only sequences that are nonspecifically bound at low affinities for the selecting antibodies. A method for further evaluating these phage (micropanning) is required.

(ii) Micropanning

An estimation of the relative strength of binding of the phage to the aPL antibody can be determined by "micropanning." Micropanning is carried out following three or more rounds of biopanning and uses the same antibody as employed in the biopanning method. The method consists of dilution of the phage from the last round of biopanning and analyzing fifty or more of these clones by micropanning. Micropanning is accomplished by growing each clone to a similar density and then incubating dilute phage at an optimal single concentration in microtitration wells previously coated with a constant amount of antibody. The optimal single concentration of phage is that concentration most likely to reveal the widest range of micropanning scores (from 0 to 4+) and, thus, permit the greatest discrimination among the clones being tested. It is based on the micropanning behavior of six randomly selected clones where the score is determined at each of several concentrations of phage obtained by serial dilution. Following the incubation with antibody, the unbound phage are washed away and the amount of bound phage is used as an indication of the affinity of the phage insert for the antibody. The amount of bound phage is determined by elution with mild acid followed by neutralization and infection of *E. coli*. The number of infected *E. coli* are then quantitated by plating the microorganisms on agar plates containing tetracycline and then determining colony densities achieved by each clone.

(iii) Phage-Capture ELISA

The phage-capture ELISA test was developed to provide an intermediate level assay to bridge the gap between the relatively low stringency of the micropanning assay and the high stringency of the phage- or peptide-ELISA assays. Preliminary studies show that some antibody preparations give too many positive clones by micropanning but none by phage-ELISA or peptide-ELISA. The limitation of the phage-ELISA described below is that only five copies of p-III are located on each phage and even with a large number of phage coated on a well, few copies of the insert are represented and detection requires that the antibody have a very high affinity for the insert. With the phage-capture ELISA, the signal is amplified many times which facilitates the detection of lower affinity, stable interactions between the antibody and the insert.

The phage-capture ELISA consists of the following steps. Microtitration wells are coated with aPL antibody and phage clones are added as in the micropanning assay. Unbound phage are washed away and the amount of bound phage is quantitated using an enzyme-conjugated goat antiserum which binds the major coat protein of the phage. Phage screened using phage-capture ELISA react with many aPL antibodies and provide a strong signal in subsequent ELISA assays. This intermediate level of sensitivity allows for greater efficiency in the peptide synthesis effort since few micropanning-positive phage are phage-capture ELISA positive. As a result, peptides synthesized from positive phage-capture ELISA phage are generally immunoreactive.

(iv) Phage-ELISA

This method of selecting phage requires very tight binding of the insert to the screening antibody. Phage are directly coated onto wells of a microtitration plate and incubated with the screening antibody. Following washes to remove unbound antibody, an anti-human IgG alkaline phosphatase conjugate is added to bind any aPL antibodies bound to the phage. APL antibodies are then detected by adding a colorimetric substrate to the well which will react with alkaline phosphatase according to methods well known in the art.

(v) Colony Blot

This assay allows large-scale colony screening of *E. coli* infected by biopanned phage. This procedure is an alternative to phage-ELISA for identifying immunoreactive clones and exhibits a comparable level of sensitivity without requiring culturing of individual phage clones prior to testing. In this assay, *E. coli* infected with phage from a round of biopanning are spread on a large diameter nitrocellulose (NC) membrane and cultured overnight on the surface of an agar plate containing tetracycline (Barbas *et al.* (1991) *Proc. Natl. Acad. Sci. USA* 88:7978-7982). Each colony results from infection by phage containing identical sequences. Several replicate transfer blots on NC are made using this NC "master" and are allowed to grow on the surface of an agar plate. Following the chemical and enzymatic disruption of phage-infected colonies on the blots, the phage may be probed by the techniques commonly used in Western blotting, *i.e.*, staining or immunoblotting. Blots that have been blocked may be incubated with the screening

aPL antibody. Following washes to remove unbound antibody, an anti-human IgG horseradish peroxidase conjugate is added to bind to any aPL-antibody that is bound to phage. The addition of a colorimetric substrate allows one to localize the discrete colonies in the master plate which represent immunospecific phage that may be cloned for further study.

(vi) Peptide-ELISA

Following DNA sequencing to determine the peptide insert sequences of the best-reacting phage in the assays described above, the corresponding peptides are made using standard Fmoc peptide chemistry as is well known in the art. For the peptide-ELISA assay, the peptides can be made, for example, as branched tetravalent molecules, *i.e.*, each molecule has four copies of the insert. Such a molecule can coat the well of a microtitration plate and still have epitopes exposed to the solution to allow binding by an antibody. The tetravalent peptides are synthesized by incorporating lysines as branch points at the first two couplings analogous to the methods used for Multiple Antigenic Peptides (MAPS) (Posnett *et al.* (1988) *J. Biol. Chem.* 263:1719-1725). A spacer consisting of glycine-serine-glycine-serine is added on each arm after the lysines and then the insert, including the framework amino acids found in the phage, proline-glycine at the carboxyl terminus and alanine-glycine-proline at the amino terminus. All amino acids in this synthesis are added one at a time using standard Fmoc methods.

These peptides are then assayed by ELISA which is carried out by coating the peptides on microtitration wells and then assaying their reactivity with aPL antibody in a standard ELISA format. In practice, the peptides usually bind very strongly to the original screening antibody and show some cross-reactivity with other aPL antibodies. Controls of non-aPL antibodies are included to eliminate nonspecific binding peptides.

(vii) Competitive Binding Peptide-ELISA

Once ELISA-positive peptides are identified, it is necessary to quantitate their relative binding affinity to the aPL antibodies and to determine whether or not two peptides bind the same population of antibodies in a given patient serum via a peptide-competition ELISA assay. In this assay, various monomeric peptides compete with tetravalent peptides coated on a microtitration plate well. To perform

the assay, the peptides to be evaluated are synthesized as monomers, *i.e.*, without the lysine branches employed in the synthesis of the tetravalent peptides, using standard Fmoc chemistry. The monomeric peptides are then purified and dissolved at known concentrations. Wells of a microtitration plate are coated with a tetravalent peptide known to bind to the aPL antibody. Serial dilutions of the monomeric peptides are incubated with a constant dilution of the aPL antibody. The dilution of the aPL antibody was previously determined by titrating the antibody against the tetravalent peptide and selecting a dilution on the downslope of the titration curve. After incubating the antibody and monomeric peptides for one hour, the antibody/peptide solutions are added to the microtitration wells and a standard colorimetric ELISA is performed. The concentration of each monomeric peptide that decreases binding of the aPL antibody with the tetravalent peptide is determined by plotting the colorimetric readings obtained for each well. The 50% inhibition point is used as the measure of the relative strength of binding for the monomeric peptides.

A variation of this assay uses microtitration plates coated with human β_2 -glycoprotein I/cardiolipin (β_2 -GPI/CL) instead of tetravalent peptide and tests the ability of monomeric peptides to block the binding of aPL antibody to the epitope(s) on β_2 -GPI/CL. In this assay, IgG-depleted human serum at an optimized concentration is used as a source of β_2 -GPI. The monomeric peptides at several concentrations are incubated with an optimized concentration of aPL antibody in a manner analogous to the assay which employs tetravalent peptide as a plate substrate. Following the incubation of aPL/peptide in (β_2 -GPI/CL) plates, antibody binding and the peptide concentration required for 50% inhibition is determined at half-maximal absorbance as in the tetravalent assay.

An additional variation of this assay tests the ability of monomeric peptides to block the binding of aPL antibody to β_2 -GPI coated directly on the wells of Nunc Maxisorp microtitration plates. In this variation, the use of cardiolipin is omitted and instead of fish gelatin, the reagent diluent and blocker used is nonfat milk/Tween.

(viii) Fluorescence Polarization Peptide Binding Assay

This assay detects direct binding of the peptide to aPL antibody. Since aPL antibodies bind to β_2 -GPI (the antigen), the ELISA competitive inhibition assay can show inhibition due to binding to β_2 -GPI as well inhibition due to binding of the

peptide to the aPL antibody. Because binding to antibodies is required in order for the peptide to function as a Tolerogen, it is essential to establish that a peptide can directly bind to an aPL antibody. This assay is used to determine the dissociation constants for peptides that bind to two aPL antibodies, ACA-6501 and ACA-6701. While the ELISA assay is useful for high throughput screening because it requires less antibody than the Fluorescence Polarization assay, however, ELISA-positive peptides should be further evaluated by the Fluorescence Polarization assay to determine whether they are capable of directly binding with aPL antibody.

(ix) Evaluation of amino acid contributions to binding by substitution and deletion synthesis

The desired epitope for tolerance induction should have as strong an interaction with as many of the aPL antibodies as possible but not contain any unnecessary residues. In order to deduce the minimum constitution of an epitope, analogs of each peptide are made (i) that lack given residues, for example, the framework residues at the carboxyl and/or amino termini are deleted, or (ii) in which amino acid substitutions have been made which differ from sequences found in the epitope library screen. These amino acid substitutions may be either natural, *e.g.*, isoleucine for leucine, or unnatural, *e.g.*, alpha methyl proline for proline. The effect of these deletions and/or substitutions are then measured via peptide-competition ELISA.

(x) Grouping of aPL Sera Specificities by Mutagenesis of the 5th domain of β_2 -GPI

For a Tolerogen to be generally effective, it must bind a major portion of the aPL antibodies in the majority of patients. It is important to determine if several antibodies from different patients bind identical residues within the eighty-four amino acid 5th domain of β_2 -GPI which has been suggested by others to contain the target epitope. If several antibodies bind identical residues, a single mimotope derived from the structural data of the peptides can be constructed which will react with all the antibodies. On the other hand, if the antibodies bind to different residues, a unique tolerogen would be required for each antibody. Site-directed mutagenesis was performed to identify if key residues involved in aPL antibody binding reside in the 5th domain of β_2 -GPI. The resulting mutant β_2 -GPIs were assayed for reactivity with

several aPL antibodies. The results were inconclusive. A fusion protein comprising the 5th domain of β_2 -GPI and glutathione S transferase (GST) was obtained from A. Steinkasserer and expressed in *E. coli* (Steinkasserer *et al.* (1992) *FEBS Lett.* 313:193-197). This fusion protein was successfully substituted for native β_2 -GPI in the ACA ELISA. Amino acid substitutions are engineered using standard site-directed mutagenesis.

I. Isolation of Synthetic aPL Epitopes

Antibody ACA-6501, from a patient with a GPL score of 151 (high titer) and a history of recurrent stroke, fetal loss, lupus and three aortic valve replacements was immunoaffinity purified on cardiolipin liposomes. The antibody was used in four separate phage library screens, using the xy, the xyz, the xy'z, and a special pro/cys-bounded 7-mer library where, based on the previous screens, Arg was fixed at the seventh position. As shown in Table 1, 36 sequences were obtained in phage that micropanned (out of 140 tested). These appear quite homologous and the conserved DR residues at positions 6 and 7 are notably striking. The consensus sequence is CLLLAPDRC. Despite this homology, only seven phage (see Table 2) were positive after phage-ELISA colorimetric testing. Screening the xy'z phage with affinity purified ACA-6626 (from a patient with a high, but lower than ACA-6501, titer) yielded five unique sequences that were phage-ELISA tested. None were color positive but two were positive when the ELISA immunoconjugate was developed with a chemiluminescent substrate. The sequence motif associated with ACA-6626 appears related but is different from that seen with ACA-6501 (see Table 3). Both antibodies preferred the cys-bounded (probably cyclized and constrained) epitopes over the open pro-bounded sequences.

TABLE 1
ACA-6501 Phage Library Sequences

<u>Clone</u>	<u>Sequence</u>	<u>Clone</u>	<u>Sequence</u>
xy Library			
2101	CNILVLDRC		
xyz Library			
5A12	CLILAPDRC	3C10	CILLAKNRC
2D7	CLLLAPDRC	3C5	CIVLVPDRC
3B6	CLVLALDRC	2F4	CLVIALDRC
3E4	CLFVALDRC	5B1	CWFRSQSSC
3E7	CILLAHDRC	3E11	CSPILRGNC
2H1	CIILAPGRC	3E8	CHKFFWLTC
xy'z Library			
2A10	CTILAPDRC	2D12	CLVLAADRC
2G12	CLLITPDRC	3B10	CLLLAPDRC
2G11	CLLITHDRC	3F2	CFFHFHDHSC
2F10	CNILVLDRC	2D3	CPLHTHHTC
2E3	CPLITHDRC		
Custom (X)₆R Library			
G11	CTILTPDRC	1A4	CNLLALDRC
2H5	CTILTPDRC	2H6	CNLLAIDRC
2H2	CTILTLDRC	1C3	CLLLAIDRC
2H10	CTLLTPDRC	1D10	CTIITQDRC
2E10	CIQLTPDRC	2H4	CNIITRDRC
1B7	CHLLTPDRC	2G12	CILHAAHRC
2H1	CLILTPDRC	1A9	CSSKSYWRC
2H12	CSILAPDRC		
xy'z Library, Colony Blot Screening Assay			
CB2	CILLARDRC		

TABLE 2
ACA-6501 Colorimetric ELISA-positive Phage

<u>Clone</u>	<u>Sequence</u>	<u>Clone</u>	<u>Sequence</u>
5A12	CLILAPDRC	3B6	CLVLALDRC
2H1	CLILTPDRC	2G11	CTILTPDRC
3B10	CLLLAPDRC*	3E7	CILLAHDRC
2H2	CTILTLDRC		

* Corresponds to consensus or average sequence

TABLE 3
ACA-6626 xy'z Phage Library Sequences

<u>Clone</u>	<u>Sequence</u>	<u>Clone</u>	<u>Sequence</u>
4B11	CGNAADARC	4G7	CTNLTDSRC
4D3	CTNWADPRC	4A2	CGNPTDVRC
4C7	CGNIADPRC		

ACA-6644 is another high-titered aPL antibody that was used to screen the pooled p-III phage libraries according to methods described herein. The following sequences were discovered:

ACA-6644/CBc	GILLNEFA
ACA-6644/CBd	GILTIDNL
ACA-6644/CBf	GILALDYV

These sequences all were derived from the component "z" epitope library that lacks phage framework residues at the N-terminus. When synthesized as peptides the sequences were immunoreactive with several ACA sera including ACA-6644 and ACA-6501. Analysis revealed unsuspected homologies with the sequences previously obtained with ACA-6501 as illustrated in Table 4.

TABLE 4

ACA-6644/CBd	<u>G I L T I D N L</u>
ACA-6501/2H2	C T <u>I L T L D R C</u>
ACA-6644/CBf	<u>G I L A L D Y V</u>
ACA-6501/2F10	C N <u>I L V L D R C</u>
ACA-6644/CBc	<u>G I L L N E F A</u>
ACA-6501/1D10	C T <u>I I T O D R C</u>

The convergent sequence homology from two very dissimilar source libraries screened by these two aPL antibodies suggests that the sequences may mimic a major, perhaps immunodominant, region in the native target antigen.

Screening of the p-III libraries with ACA-6701 yielded two unique sequences with a high degree of internal homology but unlike others previously obtained with other aPL antibody. The sequences are as shown:

ACA-6701/3B1 L S D P G Y V R N I F H

ACA-6701/3E1 L T D P R Y T R D I S N F T D

As resin-bound peptides, the sequences were strongly immunoreactive with the parent serum (ACA-6701) but were minimally cross-reactive with other aPL antibodies.

Continued screening of random pIII phage libraries with affinity purified ACA antibody resulted in the discovery of a peptide that displayed significant crossreactivity with all nine affinity purified ACA antibodies against which it was initially tested. The performance of this peptide as well as four others is shown in Table 5. Peptide sequences for all five peptides immediately follow the table. All were tested using the competitive binding peptide ACA ELISA using CL/β2-GPI plates as described herein.

TABLE 5
Percent Inhibition of AffACA by Peptide Monomers

Peptide	6626	6638	6641	6644	6701	7004	7005	6903	6501
	<u>Ave.</u>								
6641/3G3	100	100	93	100	100	100	95	100	86
	97								
1 mg/mL									
6501/3B10	76	87	52	55	51	87	47	69	60
	65								
1 mg/mL									
6626/4C7	88	96	45	45	69	99	54	77	39
	68								
1 mg/mL									
6701/3B1	43	65	43	37	96	72	23	51	41
	52								
1 mg/mL									
6644/CBf	37	60	28	18	22	54	57	43	37
	39								
200 μ g/mL									

Peptide	Sequence
ACA-6641/3G3:	AGP CLGVLGKLC PG
ACA-6501/3B10:	AGP CLLLAPDRC PG
ACA-6626/4C7:	AGP DGNIADPRC PG
ACA-6701/3B1:	AGP LSDPGYVRNIFH PG
ACA-6644/CBf:	GILALDYV GG

Subsequent testing determined that this peptide, ACA-6641/3G3 having the sequence AGP-CLGVLGKLC-PG (LJP 688) was cross-reactive with a number of ACA antisera. Chemical optimization of this peptide was pursued by truncation, systematic amino acid substitution, and non-disulfide cyclization studies.

The motifs of several peptides selected from the random phage library screening is set forth below.

TABLE 6

<u>ACA Antibody</u>	<u>Phage Insert Sequence</u>
6501	C L L L A P D R C (3B10)
6626	C T N W A D P R C
6641	C L G V L G K L C (3G3)
6644	G I L A L D Y V
6701	L T D P R Y T R D I S N F T D
6707	C A H P D W D R C

J. Immunoreactivity of aPL-related Peptides with Affinity Purified IgG aPL Antibody

Phage sequences obtained with affinity-purified ACA-6501 and found to be phage-ELISA-positive were synthesized on a solid support recently developed for combinatorial synthetic peptide libraries. This support, a Rapp resin, has a high peptide density and uses a hydrophilic polyethylene glycol spacer before the first amino acid is coupled. The synthesis resulted in resin-bound peptide that was ideally suited for antibody binding studies. As shown in Figure 2, peptide 5A12 (sequence CLILAPDRC) dramatically outperformed an unrelated control peptide while not significantly binding normal IgG. Similar results were obtained with the other phage-ELISA-positive peptides tested. In the experiment shown, resin peptide-bound affinity-purified ACA-6501 aPL antibody was detected by an immunoconjugate color reaction.

K. aPL Serum Antibody Reactivity with Synthetic Peptides

The discovery that resin-bound peptides could bind aPL immunospecifically using serum significantly enhanced the ability to test aPL antibodies. As shown in Figure 3, the peptide derived from screening with ACA-6626 bound aPL antiserum but did not demonstrate significant binding of normal serum. Figure 3 also illustrates the immunospecific-binding behavior of ACA-6501-5A12 peptide towards aPL serum. Binding of normal serum to peptide 5A12 was nil in data not shown.

L. Crossreactivity of Synthetic Peptides Towards aPL Antiserum That Was Not the Source of the Library Screening Antibody

Two peptides were selected for aPL sera testing, one (5A12) representing the ACA-6501 screen and another one (4D3) representing the ACA-6626 screen. As shown in Figures 4 and 5, each of the peptides reacted preferentially with the parent serum of the screening antibody. However, a significant degree of crossreactivity was detectable especially between ACA-6501 and ACA-6626. A survey of 19 patient sera with low or no GPL score and of 13 pathologic sera with moderate to high GPL carried out with the 5A12 resin-bound peptide showed 8 of the 13 pathologic samples with detectable peptide binding while only 2 out of 19 control samples showed low peptide binding. These results evidence that synthetic peptides are useful for diagnostic or prognostic assays in stroke patient care.

As noted in Section I above, one synthetic peptide, 6641/3G3 was tested against several high titer ACA antisera. This peptide appeared to be crossreactive with the vast majority of ACA antisera tested as illustrated in Figure 14. 6641/3G3 demonstrated dose-dependent cross-reactivity with 10 of 10 ACA antibodies with complete inhibition (>80%) at 1.8 mg/mL and appeared to be specific for ACA antibodies as demonstrated in Table 7 below.

TABLE 7
 CROSSREACTIVITY DATA FOR
 AGP-CLGVLGKLC-PG (LJP 688)
 ACA SERA WITH CARDIOLIPIN/ β 2-GPI-COATED PLATES

ACA SERUM Nr.	IC ₅₀ , mg/mL, for peptide LJP
	688
[1] 6501 RS, RFL	0.89, 1.1
[2] 6626 RS	1.09
[3] 6635 RS	1
[4] 6638 RS	0.81
[5] 6641 RS	0.89, 0.51
[6] 664 RS	0.76
[7] 6701 RS	1.07
[8] 6903 RS	0.8
[9] 7004 RFL	0.67
[10] 7005 RFL	0.75
mean \pm sd	0.86 \pm 0.16

RS = recurrent stroke

RFL = recurrent fetal loss

M. New Synthetic Peptide Methodologies

The identification of new candidate sequences by aPL library screening required testing of the new synthetic peptides for antibody binding. With Rapp resin of known peptide substitution, it is possible to carry out quantitative binding studies such as saturation binding analysis and equilibrium measurements using radiolabeled aPL IgG.

Peptide synthesis allows the molecular dissection of the mimotope by selective synthesis. This includes the modification of each amino acid along the chain with the goal of enhancing antibody binding. Selective synthesis reveals the relative importance of each amino acid in the sequence. If necessary, selective substitution at particular residue locations can be designed to maintain B cell reactivity while abolishing any T cell proliferative reactivity discovered during T cell assays.

Peptide 6641/3G3 (LJP 688) was subjected to a number of analyses. The analyses included truncation at both the N-terminus and the C-terminus, disulfide substitution, substitution of alanine and glycine for amino acids in positions 2 through 8, substitution of branched aliphatic amino acids in positions 2, 4, 5 and 8, substitution of amino acids affecting conformation of the peptide, including substitution of α -methyl amino acids, substitution of basic amino acids in position 7, substitution of D-amino acids in positions 2 through 9, and the substitution of N- α -methyl amino acids in positions 1 through 9. The results are shown below in Table 9. The structure/activity relationship of these substitutions and truncations is shown in Table 8.

TABLE 8
Optimization of Peptide 3G3

Position Number	Total Peptides												144 with ACA 6501		
	-2	-1	0	1	2	3	4	5	6	7	8	9	A	B	
Truncation analysis														<u>150. ug/mL</u>	<u>std</u>
1	A	G	P	C	L	G	V	L	G	K	L	C	P	G	850
2		G	P	C	L	G	V	L	G	K	L	C	P	G	180
3			P	C	L	G	V	L	G	K	L	C	P	G	420
4				C	L	G	V	L	G	K	L	C	P	G	460
5	A	G	P	C	L	G	V	L	G	K	L	C	G		450
6					C	L	G	V	L	G	K	L	C		10 #906 (LJP 690)
Disulfide surrogates															
1				C	L	G	V	L	G	K	L	C		110 #952 = 100	
2				Hc	L	G	V	L	G	K	L	C		46	
3				C	L	G	V	L	G	K	L	Hc		92	
4				Hc	L	G	V	L	G	K	L	Hc		80	
Ala scan															
1				C	A	G	V	L	G	K	L	C		200 #952 = 80	
2				C	L	A	V	L	G	K	L	C		170	
3				C	L	G	A	L	G	K	L	C		260	
4				C	L	G	V	A	G	K	L	C		inf	
5				C	L	G	V	L	A	K	L	C		40	
6				C	L	G	V	L	G	A	L	C		inf	
7				C	L	G	V	L	G	K	A	C		360	
Gly scan															
1				C	G	G	V	L	G	K	L	C		310 #952 = 120	
2				C	L	G	G	L	G	K	L	C		inf.	
3				C	L	G	V	G	G	K	L	C		inf	
4				C	L	G	V	L	G	G	L	C		inf	
5				C	L	G	V	L	G	K	G	C		500	

Sequence optim.	Branched aliphatic amino acids		
1	C I G V L G K L C	60	#910 = 40
2	C V G V L G K L C	130	
3	C M G V L G K L C		
4	C Cy G V L G K L C		
5	C tL G V L G K L C		
6	C mL G V L G K L C		
7	C Iv G V L G K L C		
8	C L G L L G K L C		
9	C L G I L G K L C		
10	C L G M L G K L C		
11	C L G Cy L G K L C		
12	C L G tL L G K L C		
13	C L G mL L G K L C		
14	C L G Iv L G K L C		
15	C L G V I G K L C		
16	C L G V V G K L C		
17	C L G V M G K L C		
18	C L G V Cy G K L C		
19	C L G V tL G K L C		
20	C L G V mL G K L C		
21	C L G V Iv G K L C		
22	C L G V L G K I C		
23	C L G V L G K V C		
24	C L G V L G K M C		
25	C L G V L G K Cy C		
26	C L G V L G K tL C		
27	C L G V L G K mL C		
28	C L G V L G K Iv C		

Conformational scan amino acids

1	C L P V L G K L C	
2	C L mP V L G K L C	
3	C L mA V L G K L C	
4	C L cG V L G K L C	
5	C L G V L P K L C	
6	C L G V L mP K L C	
7	C L G V L mA K L C	
8	C L G V L cG K L C	
9	C L dA V L G K L C	
10	C L G V L dA K L C	
11	C P G V L G K L C	160 #952 = 100
12	C L P V L G K L C	340 (Pro Scan)
13	C L G P L G K L C	180
14	C L G V P G K L C	Inf.
15	C L G V L P K L C	350
16	C L G V L G P L C	160
17	C L G V L G K P C	550
18	C pG G V L G K L C	130 #910 = 40
19	C L pG V L G K L C	70
20	C L G pG L G K L C	70
21	C L G V pG G K L C	35
22	C L G V L pG K L C	30 #910 = 5
23	C L G V L G pG L C	230
24	C L G V L G K pG C	80
alpha Me AA		
25	C aiB G V L G K L C	370 #910 = 40
26	C L aiB V L G K L C	80
27	C L G aiB L G K L C	110
28	C L G V aiB G K L C	inf.
29	C L G V L aiB K L C	110
30	C L G V L G aiB L C	460
31	C L G V L G K aiB C	>470

Basic amino acid

1	C	L	G	V	L	G	R	L	C	160	#952 = 120
2	C	L	G	V	L	G	Or	L	C	40	910 = 60
3	C	L	G	V	L	G	mK	L	C	-	-

D amino acids (lower case d)

1	dP	C	L	G	V	L	G	K	L	C	120	#952 = 100
2	C	dL	G	V	L	G	K	L	C	180	#952 = 30	
3	C	L	G	dV	L	G	K	L	C	260	#952 = 30	
4	C	L	G	V	dL	G	K	L	C	230	#952 = 30	
5	C	L	G	V	L	G	dK	L	C	170	#952 = 50	
6	C	L	G	V	L	G	K	dL	C	140	#952 = 30	
7	C	dL	G	dV	L	G	K	L	C	-	-	
8	C	dV	G	dV	L	G	K	L	C	-	-	
9	C	dL	G	dL	L	G	K	L	C	-	-	
10	C	L	G	dV	dL	G	K	L	C	50	#952 = 30	
11	C	L	G	dL	dV	G	K	L	C	-	-	
12	C	L	G	dV	dV	G	K	L	C	-	-	
13	C	L	G	dL	dL	G	K	L	C	-	-	
14	C	L	G	V	dL	G	dK	L	C	-	-	
15	C	L	G	V	dK	G	dL	L	C	-	-	
16	C	L	G	V	cL	G	dL	L	C	-	-	
17	C	L	G	V	dK	G	dK	L	C	-	-	
18	C	L	G	V	L	G	dK	dL	C	150	#952 = 50	
19	C	L	G	V	L	G	dL	dK	C	-	-	
20	C	L	G	V	L	G	dK	dK	C	-	-	
21	C	L	G	V	L	G	dL	dL	C	140	#952 = 30	
22	C	L	G	V	L	dA	K	L	C	140	#952 = 50	
	C	L	G	V	L	G	K	L	dC	-	-	

N-Me amino acids (nAA)

1	nC	L	G	V	L	G	K	L	C		
2	C	L	G	V	L	G	K	L	C		
3	C	L	nG	V	L	G	K	L	C	70	#952 = 50
4	C	L	G	nV	L	G	K	L	C	220	
5	C	L	G	V	nL	G	K	L	C	180	
6	C	L	G	V	L	nG	K	L	C	550	
7	C	L	G	V	L	G	nK	L	C		
8	C	L	G	V	L	G	K	nL	C	240	#952 = 50
9	C	L	G	V	L	G	K	L	nC		

Others

1	C	L	G	V	L	G	K	L	C	ATU-Y	300	#952 = 50
2	Y	ATU	C	L	G	V	L	G	K	L	C	170
3		C	L	G	V	L	G	K	L	C	TGR resin	500
4		C	L	G	V	L	G	K	L	C	dC TGR resin	620
5		dC	L	G	V	L	G	K	L	C	C	
6		cC	L	G	V	L	G	K	L	C	dC TGR resin	

Thioethers:**single deletion**

1	Hc	L	G	V	L	A	K	L	C		
2	Hc		G	V	L	A	K	L	C		
3	Hc	L		V	L	A	K	L	C		
4	Hc	L	G		L	A	K	L	C		
5	Hc	L	G	V		A	K	L	C		
6	Hc	L	G	V	L		K	L	C		
7	Hc	L	G	V	L	A		L	C		
8	Hc	L	G	V	L	A	K		C		

shrink minimization

1	Hc		V	L	A	K	L	C		
2	Hc			L	A	K	L	C		
3	Hc			L	A	K		C		

branched

1	Hc	L	G	V	L	G	K	L	Pe
2	Hc	L	G	V	L	G	K	L	dPe
3	Pe	L	G	V	L	G	K	L	Hc
4	dPe	L	G	V	L	G	K	L	Hc
5	dPe	L	G	V	L	G	K	L	dPe

D amino acids

1	dHc	L	G	V	L	G	K	L	C
2	dHc	L	G	V	L	G	K	L	dC
3	Hc	L	G	V	L	G	K	L	dC

Des amino

1	By	L	G	V	L	G	K	L	Hc	desamino HHTE
2	By	L	G	V	L	G	K	L	C	desamino HCTE
3	Pp	L	G	V	L	G	K	L	Hc	desamino CHTE
4	Pp	L	G	V	L	G	K	L	C	desamino CHTE

Abbreviations		
	Hc	homocysteine
	Cy	cyclohexyl alanine
	tL	tertiary leucine
	mL	alpha methyl leucine
	Iv	alpha methyl, alpha amino butyric acid
	mP	alpha methyl proline
	mA	alpha methyl alanine
	aiB	aminoisobutyric acid
	pG	phenylglycine
	cG	cyclopropyl glycine
	dA	D alanine
	dP	D proline
	dL	D leucine
	dV	D valine
	dK	D lysine
	nC	N alpha-methyl cysteine

nL	N alpha-methyl leucine
nG	N methyl glycine
nV	N alpha-methyl valine
nK	N alpha-methyl lysine
mK	N epsilon-methyl lysine
Pe	penicillamine
By	butyroyl
Pp	propionyl
Or	ornithine

TABLE 9
Structure Activity Relationship of the Crossreactive ACA Epitope

<u>Structure</u>	<u>Relative Activity</u>
AGPCLGVLGKLC PG (3G3) (LJP 688)	100
CLGVLGKLC (LJP 690)	3.5
CLGVLA KLC	1.8
CLGVLP GKLC	1.4
CLG _d V _d LGKLC	5.9
HcLGVLGKLC (thioether)	1.6
SLGVLGKLS	∞
CLGVAGKLC	∞
CLGVLGALC	∞

N. Use of α -methyl Amino Acids Substitutions and Unnatural Amino Acids to Enhance Peptide Immunoreactivity and Confer Resistance to Protease Attack

Proline residues have a special significance due to their influence on the chain conformation of polypeptides. They often occur in reverse turns on the surface of globular proteins. In the phage epitope libraries of the present invention, all random peptide inserts are flanked by boundary prolines. In addition, most of the mimotopes discovered with ACA-6501 have a third proline which, based on computer-based predictions, likely exists as part of a β -turn. β -turn mimetics can be used to enhance the stability of reverse turn conformations in small peptides. Such a mimetic is (*S*)- α -methyl proline (α -MePro), a proline analog that, in addition to stabilizing turn conformations, confers resistance to protease degradation. Protease resistance is a desirable property for a potential drug designed to act in the plasma. Peptide ACA-6501/3B10 AGPCLLLAPDRCPG (*insert highlighted*) is a consensus peptide. It has a sequence featuring the most prevalent residue at each position based on a comparison with 35 other homologous sequences. Due to its representative character, the sequence was subjected to a number of systematic modifications and deletions and its activity subsequently evaluated by aPL antibody binding. Among the most important findings was the discovery that the prolines at the 3 and 9 positions are important for activity. Proline-3 is derived from the phage framework and is not part of the random insert. The most dramatic effect was obtained by the substitution of α -MePro for proline at the 9-position. This substitution led to a six-fold enhancement in immunoreactivity.

The results of α -methyl and N- α -methyl amino acid substitution studies carried out on peptide 6641/3G3 are shown above in Table 8.

In addition to the twenty naturally-occurring amino acids and their homoanalogues and noranalogues, several other classes of alpha amino acids can be employed in the present invention. Examples of these other classes include *d*-amino acids, N^{α} -alkyl amino acids, alpha-alkyl amino acids, cyclic amino acids, chimeric amino acids, and miscellaneous amino acids. These non-natural amino acids have been widely used to modify bioactive polypeptides to enhance resistance to proteolytic degradation and/or to impart conformational constraints to improve

biological activity (Hruby *et al.* (1990) *Biochem. J.* **268**:249-262; Hruby and Bonner (1995) *Methods in Molecular Biology* **35**:201-240).

The most common N^{α} -alkyl amino acids are the N^{α} -methyl amino acids, such as N^{α} -methyl cysteine (nC), N^{α} -methyl glycine (nG), N^{α} -methyl leucine (nL), N^{α} -methyl lysine (nK), and N^{α} -methyl valine (nV). Examples of alpha-alkyl amino acids include alpha-methyl alanine (mA), alpha-aminoisobutyric acid (aiB), alpha-methyl proline (mP), alpha-methyl leucine (mL), alpha-methyl valine (mV), alpha-methyl-alpha-aminobutyric acid (tv), diethylglycine (deG), diphenylglycine (dpG), and dicyclohexyl glycine (dcG) (Balaram (1992) *Pure & Appl. Chem.* **64**:1061-1066; Toniolo *et al.* (1993) *Biopolymers* **33**:1061-1072; Hinds *et al.* (1991) *Med. Chem.* **34**:1777-1789).

Examples of cyclic amino acids include 1-amino-1-cyclopropane carboxylic acid (cG), 1-amino-1-cyclopentane carboxylic acid (Ac5c), 1-amino-1-cyclohexane carboxylic acid (Ac6c), aminoindane carboxylic acid (ind), tetrahydroisoquinoline carboxylic acid (Tic), and pipecolinic acid (Pip) (C. Toniolo (1990) *Int'l. J. Peptide Protein Res.* **35**:287-300; Burgess *et al.* (1995) *J. Am. Chem. Soc.* **117**:3808-3819). Examples of chimeric amino acids include penicillamine (Pe), combinations of cysteine with valine, 4R- and 4S-mercaptoprolines (Mpt), combinations of homocysteine and proline and 4R- and 4S-hydroxyprolines (hyP) and a combination of homoserine and proline. Examples of miscellaneous alpha amino acids include basic amino acid analogs such as ornithine (Or), N^{ϵ} -methyl lysine (mK), 4-pyridyl alanine(pyA), 4-piperidino alanine (piA), and 4-aminophenylalanine; acidic amino acid analogs such as citrulline (Cit), and 3-hydroxyvaline; aromatic amino acid analogs such as 1-naphthylalanine (1-Nal), 2-naphthylalanine (2-Nal), phenylglycine (pG), 3,3-diphenylalanine (dpA), 3-(2-thienyl)alanine (Thi), and halophenylalanines (e.g., 2-fluorophenylalanine and 4-chlorophenylalanine); hydrophobic amino acid analogs such as *t*-butylglycine (*i.e.*, tertiary leucine (tL)), 2-aminobutyric acid (Abu), cyclohexylalanine (Cy), 4-tetrahydropyranyl alanine (tpA), 3,3-dicyclohexyl alanine (dcA), and 3,4-dehydroproline.

In addition to alpha-amino acids, others such as beta amino acids can also be used in the present invention. Examples of these other amino acids include 2-aminobenzoic acid (Abz), β -aminopropanoic acid (β -Apr), γ -aminobutyric acid (γ -

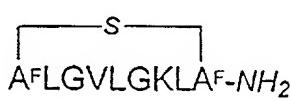
Abu), and 6-aminohexanoic acid (ϵ -Ahx). Carboxylic acids such as 4-chlorobutyric acid (By) and 3-chloropropionic acid (Pp) have also been used as the first residue on the N-terminal in the synthesis of cyclic thioether peptides.

O. Preparation of Cyclic Thioether Analogs

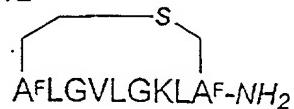
The mimetic peptides identified by the methods of the instant invention can be further modified to contain thioether substitutions. Modification of cyclic disulfide analogs to cyclic thioether analogs will extend the plasma half-life of the analog conjugates and, therefore, require a lower dosage. Cyclic thioether analogs also eliminate the problem of disulfide bond exchange which often occurs with cyclic disulfide polypeptides. In addition, the cyclic thioether analogs may also interfere with MHC Class II presentation to T cells and, thus, facilitate induction of anergy. Finally, the cyclic thioether analogs are useful in the thiol-dependent conjugation reactions used in the production of valency platform molecule conjugates.

Four cyclic thioether analogs were prepared according to the methodology described in co-owned, co-pending patent application, attorney docket number 252312006600, which is incorporated herein in its entirety. Using either a RinkTM amide 4-methyl benzhydryl amino resin or MBHA resin, full length peptide analogs of 6641/3G3 were prepared, converted into chloro-peptides, cleaved from a solid support and then cyclized. See Thioether Reaction Scheme below. Suitable cyclic thioether analogs include the analogs shown below.

CCTE



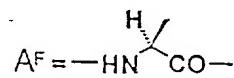
HCTE



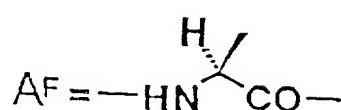
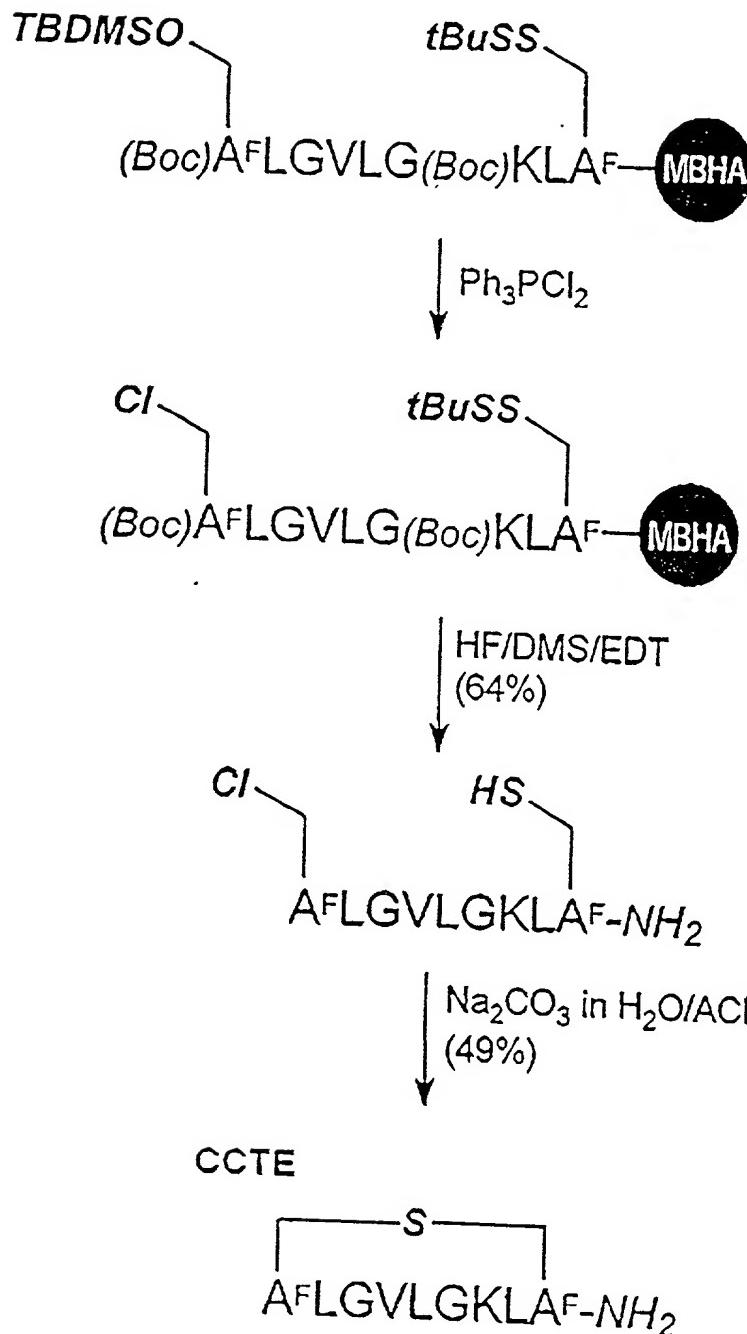
CHTE



HHTE

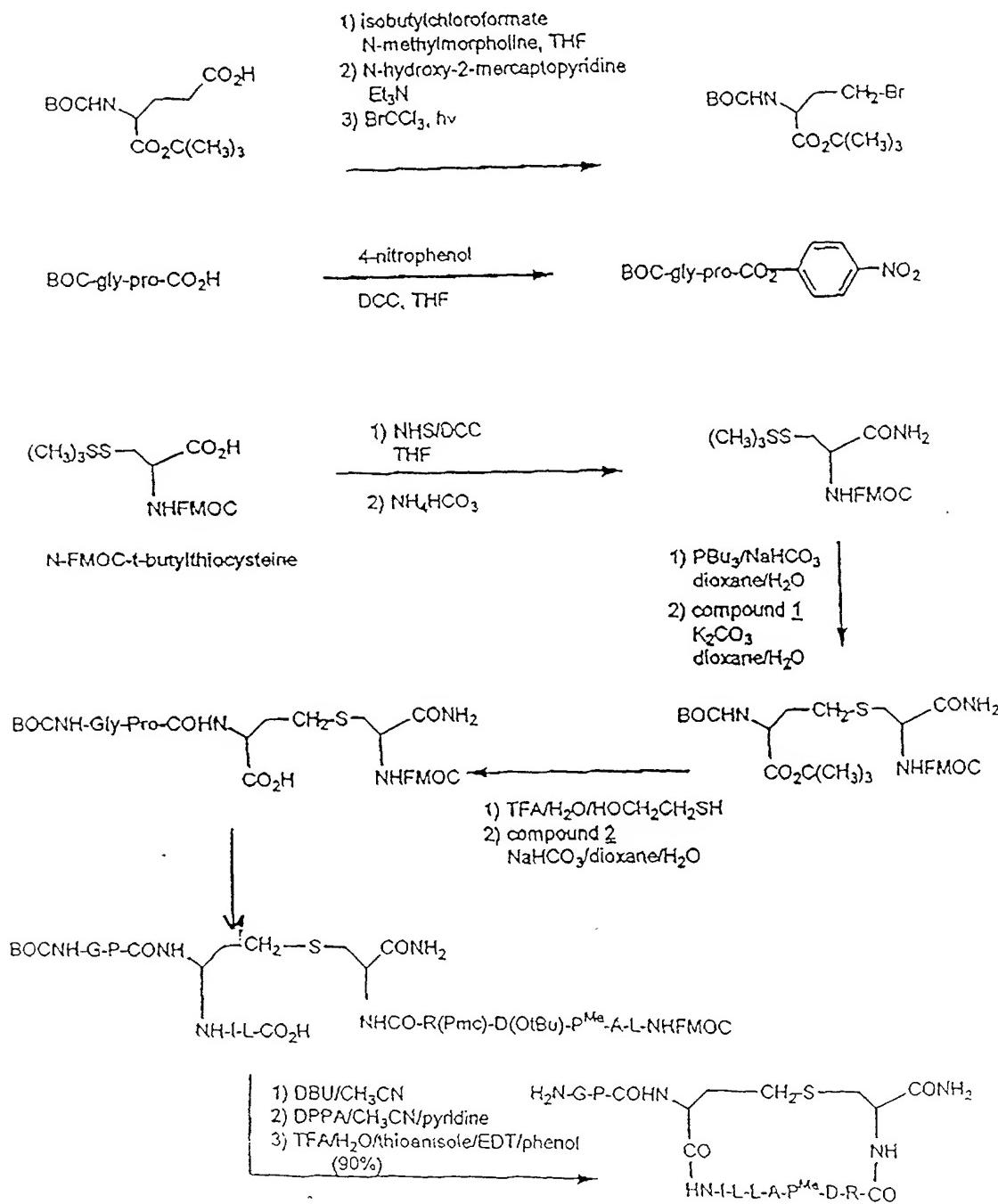


CYCLIC THIOETHER REACTION SCHEME 1

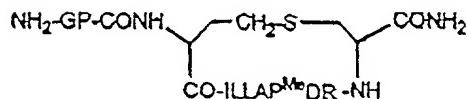


Alternatively, thioether analogs are prepared according to the reaction scheme below.

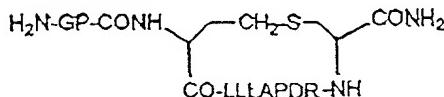
CYCLIC THIOETHER REACTION SCHEME 2



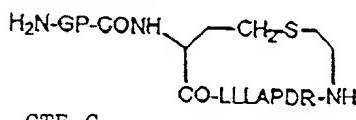
The following analogs are representative as analogs made according to cyclic thioether reaction scheme 2.



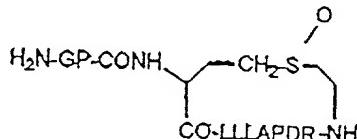
CTE A



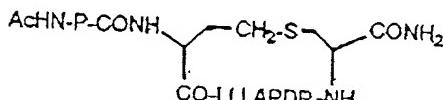
CTE B



CTE C



CTE D



CTE E

Exemplary cyclic thioether analogs were tested for activity against ACA antibodies ACA-6501 and ACA-6701. The results are shown in Table 10 below.

TABLE 10
IC₅₀ Values (μg/0.1 mL)

Thioether	ACA-6501	ACA-6701
CCTE, LJP 698	11.0	5.0
HCTE, LJP 699	4.6	3.0
CHTE, LJP 702	9.2	3.2
HHTE, LJP 703	8.0	3.6
LJP 690	10.0	4.0
CTE A	0.08	
CTE B	0.5	
CTE C	0.9	
CTE D	3.2	
CTE E	6.3	

The HCTE analog, LJP 699, outperformed the reference peptide, LJP 690, which is the truncated version, CLGVLGKLC, of LJP 688, AGPCLGVLGKLCPG.

All articles, documents, patents and patent applications cited herein are incorporated by reference herein in their entirety. The following examples are intended to further illustrate the invention and its uniqueness. These examples are not intended to limit the scope of the invention in any manner.

EXAMPLES

Example 1: Measurement of Anticardiolipin Antibodies (ACA) in serum 6501

Even numbered wells of an Immulon I microtitration plate (Dynatech Laboratories, Inc., Chantilly, VA) were coated with 50 µg cardiolipin (Sigma Chemical, St. Louis, MO) in 30 µL ethanol per well. The plate was dried overnight at 4°C and blocked with 200 µL of 10% adult bovine serum (ABS) (Irvine Scientific Co., Santa Ana, CA) in phosphate-buffered saline (ABS/PBS) for 2 hours at room temperature (RT). The plates were washed 5 times with Tris-buffered saline (TBS) prior to the addition of 10 µL of aPL standards and the test serum, ACA-6501. The aPL standards (APL Diagnostics, Inc., Louisville, KY) were reconstituted according to the manufacturer's instructions and diluted 1:50 with 10% ABS-PBS. The test serum, ACA-6501, diluted 1:50 to 1:2,000 in serial dilutions with 10% ABS-PBS, was added to selected duplicate wells and incubated for 2 hours at RT. The plate was washed five times with TBS and 100 µL of 1:1,000 goat-anti-human-IgG/alkaline phosphatase conjugate (Zymed, South San Francisco, CA, Cat. No. 62,8422), in 10% ABS-PBS was added and incubated for 1 hour at RT. Again, the plate was washed five times with TBS and the assay was developed by adding 100 µL phenolphthalein monophosphate (PPMP) substrate solution (Sigma, Cat. No. P-5758), prepared from a stock solution of 0.13 M PPMP and 7.8 M 2-amino-2-methyl-1-propanol, adjusted to pH 10.15 with HCl, after a dilution of 1:26 with deionized water. After approximately 30 minutes, the reaction was stopped with 50 µL of 0.2 M dibasic sodium phosphate (Mallinckrodt, Analytical Reagent) added per well. The optical density was read at 550 nm in a microplate autoreader (Bio-Tek Instruments, Winooski, VT, Model EL311). The optical density of the odd-numbered control wells (blank, without cardiolipin (CL)) was subtracted from the optical density of the even-numbered wells. The absorbance readings of the aPL standards were plotted using Graph Pad Prizm (Graph Pad Software, Inc., San Diego, CA) to generate the GPL (IgG phospholipid) standard curve. The diluted 6501 test serum absorbance readings were used to calculate GPL scores based on the GPL standard curve.

In the modified ACA ELISA, Nunc Maxisorp microtitration plates were coated with 100 µL /mL β₂-GPI made up in PBS following incubation at 1 hour at

room temperature. Following this step, microplates were blocked with PBS containing 2% nonfat milk and 0.4% (w/v) Tween 80 for 2 hours at room temperature. Except for the use of nonfat milk/Tween as reagent diluent and blocking agent, the Nunc microplates coated with β_2 -GPI directly were used for ACA IgG binding studies as previously described for the Dynatech microplates coated first with cardiolipin before the β_2 -GPI was added.

Example 2: Anticardiolipin Antibody (ACA) Purification from Serum 6501

In a 25 mL round-bottom flask (Kontes Scientific Co., Vineland, N.J.) a mixture of 1.2 mL cardiolipin (Sigma Chemical, St. Louis, MO, #C-1649), 0.464 mL cholesterol (Sigma Diag., St. Louis, MO., #965-25), 0.088 mL of 5 mg dicetylphosphate (Sigma Chemical, St. Louis, MO, D-2631) per mL chloroform was dried for approximately 5 minutes in a Rotavap (Buchi, Switzerland). Following the removal of solvent, 2 mL of 0.96% (wt./vol.) NaCl (J.T. Baker, Inc., Phillipsburg, NJ) Baker analyzed reagent was added and mixed in a Vortex Genie Mixer (Scientific Industries, Inc., Bohemia, NY) for 1 minute. The liposome suspension was incubated for 1 hour at 37°C. Meanwhile, serum 6501 was spun at 600 X g in a Sorvall RT 6000 centrifuge (Dupont Co. Wilmington, DE) for 10 minutes at 8°C. Four mL of the supernatant was placed in a 25 mL round-bottom flask with 1 mL of the prepared liposome suspension and the mixture was incubated with agitation at medium speed in an orbital shaker, Tektator V (Scientific Products, McGraw Park, IL) for 48 hours at 4°C, and an additional 2 hours at 37°C. Twenty mL of cold TBS was added and the mixture was transferred into a 50 mL polycarbonate centrifuge tube (Nalge Co., Rochester, NY) and centrifuged at 27,000 X g for 15 minutes at 4°C. in an RC3 centrifuge in a SS-34 rotor (Sorvall-Dupont, Wilmington, DE). The precipitate was washed 3 times with 25 mL of cold 0.96% NaCl using the RC3 centrifuge. The pellet was dissolved in 1 mL of 2% (wt/vol) solution of n-octyl- β -D-glucopyranoside (Calbiochem, La Jolla, CA) in TBS and applied to a 0.6 mL protein A/cross-linked agarose (Repligen Corporation, Cambridge, MA) column which had been pre-washed with 15 times bed volume of 1 M acetic acid and equilibrated with 15 times bed volumes of TBS. The antibody-protein A/agarose column was washed with 40 times bed volume of 2% octylglucopyranoside to remove lipids, followed by extensive

washings with TBS until the optical density of the eluate at 280 nm approached the baseline. The bound antibody was eluted with 1 M acetic acid. One mL fractions were collected, neutralized immediately with 0.34 mL 3 M Tris (Bio-Rad, electrophoresis grade reagent) per fraction and kept in an ice bath. The optical density of each fraction was determined at 280 nm in a spectrophotometer (Hewlett-Packard, 8452A Diode Array Spectrophotometer, Palo Alto, CA). Fractions containing antibody were pooled, concentrated and washed 4 times with TBS in Centricon-30 concentrators (Amicon Division, W.R. Grace & Co., Beverly, MA) per manufacturer's protocol. The final yield of purified antibody from 4 mL of serum 6501 was determined by reading the optical density at 280 nm of an aliquot from the concentration, where 1 mg = 1.34 OD₂₈₀. The average yield obtained was 750 µg antibody from 4 mL of serum 6501. The purified antibody was tested for ACA activity and checked for purity with Laemmli SDS-PAGE. An affinity adsorbent containing β₂-GPI was prepared using CNBr-activated agarose (Pharmacia, Inc., Piscataway, NJ) or Affi-Gel 10 (BioRad, Richmond, CA) in accordance with the manufacturer's instructions using purified β₂-GPI obtained commercially (PerImmune, Rockville, MD). To a 1 mL gel column containing the β₂-GPI affinity adsorbent, up to 100 µg of liposome-purified aPL IgG in 1 mL TBS was added. After 1 hour, the column was extensively washed with buffer to displace contaminants and any IgG that did not bind β₂-GPI. The β₂-GPI-binding IgG was displaced with 1 M HOAc and the fractions neutralized with Tris as described above. Fractions containing IgG were pooled, concentrated and subjected to buffer exchange as previously described.

Example 3: Construction of a p-III Library Vector Preparation

fUSE 5 (Scott, J.K. and G. Smith, *supra*) was used as the vector for the construction of p-III libraries, and a variation of the method of Holmes, D.S. and M. Quigley (1981), *Anal. Biochem.* 144:193) was employed to generate the double-stranded replicative form (RF). Briefly, an 800 mL culture of *E. coli* K802, harboring fUSE 5, was grown in 2YT medium (Disco Labs, Ann Arbor, MI) containing 20 micrograms/mL tetracycline for 18 hours at 37 degrees with vigorous shaking. Cells were collected by centrifugation and resuspended in 75 mL STET. STET

consists of 8% sucrose in 50 mM Tris/HCl pH 8.0, 50 mM EDTA and contains 0.5% Triton X-100. Lysozyme, 10 mg/mL in STET, was added to a final concentration of 1 mg/mL. After 5 minutes at RT, three equal aliquots were placed in a boiling water bath with occasional shaking for 3.5 minutes. The viscous slurry was centrifuged for 30 minutes at 18000 x G and an equal volume of isopropanol was added to the supernatant. The solution was cooled to -20°C and the nucleic acids were collected by centrifugation. The RF was isolated from a CsCl gradient as described by Sambrook *et al.* MOLECULAR CLONING: A LABORATORY MANUAL (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 2d ed., 1989).

Preparation of the random insert

The DNA for insertion was generated by the "gapped duplex" method described by Cwirla *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87:6378-6382. In this method, an oligonucleotide containing a degenerate region in the middle is surrounded by short constant regions on each end of the oligonucleotide. Two shorter, complementary oligonucleotides are annealed to form a "gapped duplex" possessing overhangs that are complementary to the sticky ends produced by the restriction endonuclease used to digest the vector. In this case, the longer degenerate oligonucleotide has the sequence:

5' GGGCTGGACCC(NNK)_xCCGGGGGCTGCTG 3' where N = A or C or G or T, K = G or T, and x is the number of codons in the random regions. EpiGS2 was designed to base pair to the 5' end of the degenerate oligonucleotide and has the sequence 5' GGGTCCAGCCCCGT 3'. Similarly EpiGS3 was designed to anneal to the 3' end of the degenerate oligonucleotide and has the sequence 5' CAGCCCCCGG 3'. When correctly annealed, the three oligonucleotides form a "gapped duplex" which, when inserted into fUSE 5 digested with Sfi1, restores the reading frame of p-III with the random insert near the 5' end.

Oligonucleotides described above were prepared by excision from polyacrylamide gels. One nanomole of EpiGS2, EpiGS3, and 50 picomoles of the degenerate oligonucleotide were kinased separately in 66 microliter volumes. The three oligonucleotides were then pooled, NaCl was added to 50 mM and the mixture was heated to 65°C for 5 minutes followed by slow cooling to RT. The annealed

oligonucleotides were then cooled on ice and used immediately in the ligation reaction with fUSE 5. The ligation reaction consisted of 10 micrograms of fUSE 5 DNA digested to completion with Sfi1, 30 μ L of the "gapped duplex" solution and 1000 U of T4 ligase in a total volume of 450 microliters. The ligation was incubated for 18 hours at 16°C. The mixture was then phenol-chloroform extracted, precipitated with ethanol and the precipitate dissolved in 20 μ L of water.

Generating and Amplifying the Library

The ligated DNA was introduced into *E. coli* by electroporation (Dower *et al.* (1988) *Nucleic Acid Res.* 16:6127-6145). Frozen electrocompetent MC1061 cells (0.1 mL) were mixed with 4 μ L of ligated DNA in a cold 2 mm cuvette and subjected to 2.5 kV, 5.2 mS pulse by a BTX electroporation device (BTX Corp., San Diego, CA). Immediately after the pulse, 1 mL of SOC, a cell growth medium (see Dower *et al.*, *supra*) was added. Five separate electroporations were carried out, pooled, and incubated at 37°C for 1 hour. At that time, samples were removed and diluted to determine the total number of clones generated. The balance of the mixture was diluted in 1L of 2YT (1.6% Peptone, 140, 1% Yeast Extract, and 0.5% NaCl, Difco Labs, Ann Arbor, MI) containing 20 micrograms per mL of tetracycline and grown for 18 hours at 37°C while shaken at 275 rpm. Phage were purified by 2 rounds of PEG/NaCl precipitation and resuspension in 1.2 mL TBS containing 0.02% sodium azide. Particle number was estimated by absorbance at 269 nm. The titer of the phage was determined by mixing 10 μ L of dilutions of phage with 10 microliters of starved *E. coli*.

Example 4: Screening a p-III Library with aPL Antibody

Affinity-purified ACA-6501 (affACA-6501, 10 μ g; 7 μ L of 1.44 mg/mL stock solution) was incubated for 2 hours at RT in siliconized 1.4 mL microfuge polypropylene tubes with pooled phage from the x, y and z libraries (epi^{xyz}) [11 μ L + 8.5 μ L + 2 μ L, respectively; total volume = 21.5 μ L, $\sim 10^{10}$ clones] in a final volume of 100 μ L TBS, pH 7.4, with 0.5% bovine serum albumin (BSA). During this incubation, the final steps in preparing freshly starved *E. coli*, strain K-91, were carried out. A suspension of *E. coli* freshly grown in 2YT medium for about 5 hours with 250 rpm

shaking at 37°C was spun at 1000 x g for 10 minutes at RT in 50 mL polypropylene tubes. Twenty mL of 80 mM NaCl was added to the packed *E. coli* pellet and then incubated for 45 minutes at 37°C at 100 rpm. Following centrifugation as above, the starved *E. coli* pellet was suspended in 1 mL of 50 mM ammonium phosphate/80 mM NaCl and used later for phage amplification. Protein G-agarose beads were washed 2x in TBS/BSA and 2 times in TBS/0.5% Tween-20 and stored at 4°C as a 50% suspension in TBS/Tween.

Two hundred μ L of the protein G-agarose bead suspension was then added to the ACA-6501/phage mixture and incubation continued for an additional 1 hour at RT. At this point, the mixture was chilled and washed 3 times with cold TBS/Tween and the precipitate was collected in a microfuge in a cold room. The washed beads were transferred to new microfuge tubes prewashed with TBS/BSA and TBS/Tween to prevent non-specific adherence. After three additional washes with TBS/Tween, the beads with bound ACA-6501-bearing phage were eluted with 300 μ L 0.2 N HCl/glycine, pH 2.1 by tumbling for 10 minutes at RT. Following centrifugation at 16,000 X g, the acidic eluate supernatant was collected and an additional 100 μ L of elution solution was added to the bead pellet and the procedure repeated. After 10 minutes, the phage-containing eluates (representing the unamplified first round phage) were pooled (~400 μ L) and placed in a sterile 17 x 100 mm polypropylene cell culture tube to which was added 50 μ L 0.5 M NaCl followed by pH neutralization with 2.5 M Tris base (usually ~25-35 μ L). An equal volume of the starved *E. coli* suspension was added immediately and then incubated for 10 minutes at 37°C at 100 rpm. The mixture was then transferred to a 250 mL sterile culture flask containing 25 mL 2YT with 20 μ g/mL tetracycline (Tet) and incubated overnight at 37°C at 250 rpm.

To isolate amplified phage from overnight cultures, the suspension was centrifuged at 12,000 x g for 10 minutes in polycarbonate tubes and the pellet discarded. After heating the supernatant at 70°C for 30 minutes in polypropylene tubes, the material was spun again in polycarbonate tubes and the supernatant saved. To the supernatant, 1/4 volume of 20% (w/v) polyethylene glycol, molecular weight 8000 (PEG 8000) was added to precipitate phage. The solution was mixed by inversion 100 times and then incubated at 4° C for 2 hours. After centrifugation at

35,000 x g for 30 minutes at 4° C, the phage-containing pellet was resuspended in ~0.5 mL of TBS/BSA and transferred to a 1.4 mL microfuge tube. After a 1 minute spin in a microfuge at 16,000 x g, the supernatant was transferred to a clean tube and labeled first round amplified phage.

During second, third and fourth rounds of biopanning, 75 µL of amplified phage from the preceding round was incubated with 7 µL affACA-6501 in a final volume of 100 µL. For fifth round phage, affACA-6501 was diluted first at 1:1000, then treated as described for the other rounds. All subsequent steps were carried out as described for the first round. Phage from five rounds of biopanning were spot-titered on 2YT/Tet plates to determine phage concentration. The spot titer of amplified phage requires an initial phage dilution of 1 X 10⁶ in TBS/BSA or 2YT media. For each round, 10 µL of the dilute phage was incubated with 40 µL of starved *E. coli* for 10 minutes at 37°C with no shaking. Following the addition of 950 µL of 2YT/dilute Tet (0.2 µg/mL), the mixture was incubated for 30-45 minutes at 37°C with 250 rpm shaking. Ten µL aliquots of neat and diluted phage solutions, 1:10, 1:100, and 1:1000, were spotted in 2YT/dilute Tet using agar plates containing 20 µg/mL Tet.

Micropanning

Immulon type 2 plates were coated with protein G. Protein G was prepared at 10 µg/mL in 0.1 M NaHCO₃ and 100 µL per well was added to the wells of microtitration plates and incubated overnight at 4°C. After discarding excess protein G solution from plates, each well was blocked with 250-300 µL 2YT for 1 hour at RT with agitation on an oscillating platform. Tris-buffered saline, pH 7.4/0.5% Tween 20 (TBS/Tween), was used with an automatic plate washer to wash the wells 4 times with 200 µL. One hundred µL affACA-6501 (or control normal IgG), diluted to 2.5 µg/mL with 2YT, was added to washed wells. The plate was transferred to a cold room rotator near the end of a 1 hour incubation at RT on a rotating platform.

Phage to be tested by micropanning were obtained from the agar plates generated by biopanning. Each clone to be tested was transferred using sterile toothpicks to a separate well of a round-bottom 96-well microtitration plate (Corning,

Corning, NY) containing 250 µL 2YT/Tet per well and cultured overnight at 37°C. Clone designations are based on the screening antibody, the biopanning round of origin, and the location of the clone in the overnight culture plate, *e.g.*, ACA-6501/3B10 refers to the clone isolated by ACA-6501 in the third round located in the well designated B10 on the microtitration plate. Following overnight incubation, phage cultures were centrifuged using a microtitration plate holder at 1300 x g for 10 minutes at RT. Supernatants constituted the source of "neat" phage.

Initial micropanning was restricted to six clones which were tested at dilutions expanded by factors from 1:10 to 1:10⁶. The results from these pilot clones were used to suggest the appropriate single dilution that would yield gradable results for clones in the source plate for each round. Plates representing cultured, randomly chosen clones from the 3rd, 4th, and 5th rounds of biopanning were diluted to 1:100,000 from the "neat" solution using 2YT in microtitration plates with a final volume per well of 250 µL. From the last plate representing the desired dilution, 100 µL was added to the plate containing protein G-bound ACA-6501 and normal IgG prepared as described above. The incubation of dilute phage with aPL antibody or control IgG was carried out for 2 hours at 4°C on a flat rotator. After 9 washes with TBS/Tween in an automated plate washer, the IgG-bound phage was eluted with 20 µL of 0.2 N HCl-glycine/0.1% BSA, pH 2.2. The elution incubation continued for 10 minutes at RT, during which time a new Corning microtitration plate was prepared containing 20 µL of freshly starved *E. coli* per well and kept chilled. One hundred forty µL of 29 mM Tris was added to the plate containing the phage eluates in order to neutralize the pH, following which 20 µL of phage suspension was transferred from each well to the corresponding well in the plate containing starved *E. coli*. After a 10 minute incubation at 37°C, 200 µL 2YT/dil Tet was added and incubation carried out an additional 30 minutes at 37°C. Using multichannel pipettors, 10 µL from each well was spotted on a large 2YT/Tet agar plate while retaining the original 8 x 12 well pattern and orientation from the last microtitration plate. After allowing the spots to dry for 30 minutes, the plate was incubated overnight at 37°C. The following day, colonies were semiquantitatively scored from 0 to 4+, with 0 symbolizing <10 colonies; +/-, 10-20; 1+, 20-50; 2+, 50-70% confluent; 3+, 70-90% confluent; and 4+ representing >90% confluent colonies. Of the 94 clones that were examined at a

dilution of 1:10⁵ [representing 81 third, 6 fourth, and 7 fifth round clones], six clones had micropanning scores of zero, three scored 1+, 14 scored 2+, 62 scored 3+, and 9 scored 4+. A survey of random clones from the plates representing the second through fifth rounds of biopanning was carried out by G-track DNA sequencing as described below. The results showed a dramatic drop in sequence diversity by the fourth round of biopanning (see Figure 8), therefore, a second plate was micropanned using phage at a dilution of 1:3 X 10⁵, this time including clones from an earlier (second) round. A total of 94 clones were tested, including 29 which had scored high at 1 X 10⁵ dilution [26 from the 3rd round, 1 from the 4th found, and 2 from the 5th round] plus 65 clones from the 2nd round that had not been previously tested. Of the 94 clones tested at a dilution of 1:3 X 10⁵, 26 scored zero, 11 scored +/-, 10 scored 1+, 13 scored 2+, 34 scored 3+, and zero scored 4+.

G-track DNA Sequencing

Single-stranded viral DNA was isolated from cultures incubated overnight at 37°C. To prepare cultures, 2YT/Tet, either as 2 mL in tubes or 250 µL in microtitration plate round-bottom wells, was inoculated with individual phage from spread plates of previously grown cultures in microtitration plates. The purification of phage by 20% PEG/2.5 M NaCl precipitation of culture supernatants as well as the isolation or release of virion DNA by phenol-chloroform extraction or by alkali denaturation was performed as described in Smith, G.P. and J.K. Scott, "Libraries of peptides and proteins displayed on filamentous phage" (1993) *Meth. Enzymol.* 217:228-257 for cultures in tubes and as described in Haas, S.J. and G.P. Smith G.P., "Rapid sequencing of viral DNA from filamentous phage" (1993) *BioTechniques* 15:422-431 for phage in microtitration plates. The dideoxy nucleotide chain termination DNA sequencing technique of Sanger *et al.* (Sanger *et al.*, "DNA sequencing with chain terminating inhibitors" (1970) *Proc. Natl. Acad. Sci.* 74:5463-5467) was carried out using a commercial Sequenase kit (U.S.

Biochemical/Amersham, Arlington Heights, IL) as described by Smith and Scott, *supra*, for tube culture phage DNA and as in Haas and Smith, *supra*, for phage DNA from microtitration plates. By using only the ddCTP termination mixture, G tracking or the sequence pattern suggested by a single base (G) was obtained following

electrophoresis on 7% polyacrylamide/urea sequencing gels and exposure by autoradiography. Standard gel electrophoresis and autoradiography procedures were followed (Sambrook *et al.*, *supra*).

G-tracking of 76 clones from the micropanning plate tested at a phage dilution of 1:1 X 10⁵ from the ACA-6501 library screen revealed 11 unique sequences, while 64 clones from the second micropanning plate tested at a phage dilution of 1:3 X 10⁵ showed 30 unique sequences. Conventional DNA sequencing using all four dideoxynucleotide triphosphates was applied to the phage clones with the highest micropanning scores and unique sequences and resulted in the 12 sequences shown in Table 1 for the xyz library.

Phage-Capture ELISA

Clones ACA-6635/3A12, 3B3, 3C8, 3A5, 3C9, and 3B7 were grown as 3 mL cultures. Affinity purified ACA-6635 was diluted to 2.5 µg/mL in phosphate-buffered saline, pH 7.2, and 100 µL added to Immulon-2 microtitration plate wells. After 2 hours, the plates were washed 3 times with TBS/Tween in an automated plate washer with no shaking. The plate was then blocked with 150 µL 0.1% BSA (globulin-free) in PBS per well. After 1 hour at 4°C, the plate was washed 3 times as previously described. After centrifuging each phage culture 3 minutes at 17,000 x g, each supernatant was diluted 1:10 in 0.1% BSA/PBS and 100 µL added to each of the wells coated with affinity purified ACA-6635 and then incubated for 2 hours at 4°C. Plates were then washed with TBS/Tween as before. Horseradish peroxidase-conjugated sheep IgG anti-M13 phage antibody (Pharmacia, Inc., Piscataway, NJ) was diluted 1:5,000 in 0.1% BSA/PBS and 100 µL applied to each well. Following incubation for 1 hour at 4°C, the plate was washed 4 times as before. One hundred µL of substrate prepared according to the conjugate manufacturer's instructions was added to each well. After 19 minutes, the absorbance at 405 nm of each well was read in the automated microplate absorbance reader (Biotek, Winooski, VT). The seven clones tested from the ACA-6635 phage library screen were selected because of their high micropanning scores and negative phage-ELISA scores (see below). As shown in Figure 9, three clones (3A12, 3B3, and 3A5) out of the seven tested exhibited a very strong immunospecific signal in the phage-capture ELISA.

Phage-ELISA

Three mL cultures were prepared from 35 clones previously isolated with affinity purified ACA-6501 in several phage library screens as well as one fUSE 2 phage clone lacking a peptide insert which was used as a control. After centrifugation for 3 minutes at 17,000 X g, 100 µL from each phage supernatant (adjusted to $\sim 2 \times 10^{11}$ particles/mL, based on absorbance) was added to microtitration plate wells (Falcon, Becton-Dickinson Labware, Lincoln Park, NY) and allowed to incubate overnight at 4°C. Following four washes with TBS 7.4, the plate was blocked with 125 µL of 0.5% BSA/TBS for 1 hour at RT. After another four TBS washes, 100 µL of affACA-6501 previously diluted to 2.5 µg/mL in TBS/BSA was added to each well and allowed to incubate 1 hour at 37°C. Following an additional four washes, 100 µL anti-human IgG diluted 1:1000 in TBS/0.5% Tween was added to each well. After 1 hour at RT, enzyme substrate was added and the incubation allowed to proceed for 2 hours. Following the addition of 50 µL 0.2 M Na₂HPO₄ to stop the reaction, absorbance was measured at 550nm in the automated plate absorbance reader. As shown in Figure 10, seven of the clones had significant signals in the phage-ELISA with ACA-6501: 5A12, 3B6, 3E7, 3B10 (same sequence as 2D7), 2G11, 2H1, and 2H2. The sequences for these clones are shown in Table 2.

Peptide-ELISA

In the standard protocol, stock solutions of tetrameric peptide in dimethylformamide were diluted 1000 times to 10 µg/mL in pH 9.5 carbonate buffer. Each microtitration plate well was coated overnight at 4°C with 100 µL of dilute peptide followed by blocking with buffered albumin. Peptide-coated microtitration plates were incubated 1-2 hours at room temperature with aPL sera at several dilutions starting at 1:50. Following washes, the presence of peptide-bound human IgG was determined with enzyme-conjugated anti-human IgG according to standard ELISA procedures.

Competitive Binding Peptide-ELISA

(A) Each well of an Immulon II plate (Dynatech Laboratories, Inc., Chantilly VA) was coated with 100 µL of a solution containing 10 µg tetravalent peptide ACA 6501/3B10 in 50 mM sodium carbonate, pH 9.5, containing 35 mM sodium bicarbonate (Fisher Scientific, Pittsburgh, PA, reagent grade) for at least 1 hour at RT, except for three wells used as blank controls. The liquid was then removed from the wells and 200 µL of 0.5% (wt/vol) BSA (Sigma Chemical, St. Louis, MO, #A7638) in TBS was added per well including the blank wells for blocking and incubated for at least 1 hour at RT. Four 1.5 mL microfuge tubes were numbered 1 to 4. The following reagents were mixed in the first microfuge tube (Brinkman Instruments, Westbury, NY): 30 µL of 5% BSA; 284 µL TBS; 8 µL of a stock solution of approximately 400-500 µg/mL of monomeric peptide (ACA-5A12 or -CB2 or -3B10 or scrambled -3B10 as negative control) in TBS; and 8.2 µL of 1:10 diluted serum 6501 in 0.5% BSA-TBS. The following reagents were mixed in the second microfuge tube: 30 µL of 5% BSA; 290 µL TBS; 2 µL of a stock solution of approximately 400-500 µg/mL of monomeric peptide (ACA-5A12 or -CB2 or -3B10 or scrambled -3B10 as negative control) in TBS; and 8.2 µL of 1:10 diluted serum 6501 in 0.5% BSA-TBS. The following reagents were mixed in the third microfuge tube: 30 µL of 5% BSA, 287 µL of a 1:10 dilution of approximately 400-500 µg/mL of monomeric peptides (5A12, CB2, 3B10, or scrambled sequence 3B10 control), and 8.2 µL of ACA-6501 serum previously diluted 1:10 in 0.5% BSA-TBS. The following reagents were mixed in the fourth Eppendorf microfuge tube: 60 µL 5% BSA; 584 µL TBS and 16.5 µL of 1:10 diluted serum 6501 in 0.5% BSA-TBS. The blocked plate was washed 5 times with TBS. The solution in the first microfuge tube was added to triplicate wells, 100 µL per well. Identical amounts of the solutions in the second, third and fourth microfuge tubes were also added to triplicate wells. An aliquot of 100 µL of the solution in the fourth microfuge tube was added to each of the three blocked blank wells of the microtitration plate. The plate was incubated for 1 hour at RT with agitation at 40 rpm in an orbital shaker (American Dade, Miami, FL, Rotator V) and then washed 5 times with TBS. An aliquot of 100 µL of 1:1000 diluted goat-anti-human-IgG/alkaline phosphatase conjugate (Zymed, South San Francisco, CA,

Cat. no. 62-8422) in 0.5% BSA-TBS was added and incubated for 1 hour at RT. The plate was then washed 5 times with TBS and the assay was developed by adding 100 μ L PPMP diluted substrate solution as described in Example 1. After 20 minutes, the reaction was stopped by adding 50 μ L of 0.2 M Na₂HPO₄ (Mallinckrodt, St. Louis, MO., reagent grade) per well. The optical density was read at 550 nm in a microplate reader (Bio-Tek Instruments, Winooski, VT, Model EL 311). The optical density at 550 nm versus the amount of the peptide per well was plotted in Graph Pad Prizm (Graph Pad Software, Inc., San Diego, CA). The amount of peptide required for 50% inhibition of binding of serum 6501 to tetravalent 3B10 was calculated from the graph.

(B) Immulon I® 96-well, flat-bottom, polystyrene microtitration plates (Dynatech Laboratories, Inc., Chantilly, VA) were coated with 30 μ L/well of cardiolipin (CL, 50 μ g; Sigma Chemical Co., St. Louis, MO) in ethanol. Two control wells received 30 μ L ethanol only. After overnight evaporation at 4°C, each well was blocked with 200 μ L of 5% (w/v) fish gelatin in PBS for 2 hours at RT. The CL-coated, blocked plate was washed 5 times with TBS and then to each well was added β 2-GPI as 100 μ L of 2.3% (v/v in PBS) IgG-depleted human serum (Sigma Chemical Co., St. Louis, MO) and incubated 2 hours at RT.

During this incubation, variable amounts of each of six peptides were mixed with 22 μ L ACA-6501 serum diluted with 3% fish gelatin in 1:1 TBS/PBS (final dilution of 1:400) in a final volume of 220 μ L using Eppendorf microcentrifuge tubes. Specifically, in tube #1 were mixed 181.3 μ L of 3% fish gelatin in TBS-PBS, 16.7 μ L of peptide stock solution plus 22 μ L of ACA-6501 serum diluted 40 times in 3% fish gelatin/TBS-PBS. Stock solutions ranged from 450 - 800 μ g/mL for peptides #951 (dserine non-cyclized negative control), #952 (a lot of LJP 690), and thioethers CCTE-3G3, CHTE-3G3, HCTE-3G3 and HHTE-3G3. To tube #2 the following were added: 148 μ L fish gelatin/TBS-PBS, 50 μ L of peptide stock solution, plus 22 μ L of 40 times diluted ACA-6501 serum. Tube #3 contained 48 μ L fish gelatin/TBS-PBS, 50 μ L of peptide stock solution, plus 22 μ L of 40 times diluted ACA-6501 serum. The control tube #4 received 396 μ L fish gelatin/TBS-PBS and 44 μ L of 40 times diluted ACA-6501 serum (no peptide). Each of the tubes incubated for approximately 1 hour at RT.

The CL/ β 2-GPI microtitration plate was washed 5 times with TBS and 100 μ L aliquots in duplicate from Eppendorf tubes #1, 2, 3, and 4 containing the antibody-peptide (or no peptide mixtures) were added to the wells. A volume of 100 μ L from tube #4 was added to the duplicate control wells containing no cardiolipin. The microtitration plate was incubated for 1 hour at RT with agitation at 40 rpm in an orbital shaker (American Scientific, Rotator V), washed 5 times with TBS and then 100 μ L of 1:1000 goat anti-human IgG alkaline phosphatase conjugate (Zymed, Cat No. 62-8422) in 0.5% (w/v) BSA-TBS was added. Following incubation for 1 hour at RT, the microtitration plate was again washed 5 times with TBS and the colorimetric enzyme detection developed by adding 100 μ L of PMPP solution (7.8 g phenolphthalein monophosphate plus 69.5 g of 2-amino-2-methyl-1-propanol in 100 mL water stock solution diluted 1:26 with water). After 21 minutes, the reaction was stopped by adding 50 μ L of 0.2 M Na₂HPO₄ (Mallinckrodt) to each well. Absorbance at 550 nm was read in a microplate reader (Bio-Tek Instruments, Model EL 311). Absorbance vs. peptide added was plotted on Graph Pad Prism (Graph Pad Software, Inc.) as shown in Figure 12. The amount of peptide that inhibited ACA-6501 binding by 50%, the IC₅₀, was calculated from the graph at the intersection of half-maximal absorbance with amount of peptide added.

(C) Nunc Maxisorp® 96-well, flat-bottom microtitration plates were coated with 100 μ L/well of purified human β_2 -glycoprotein I (PerImmune, Rockville, MD) at 10 μ g/well in PBS. Two control wells received 100 μ L PBS only. After two hours incubation at room temperature, the liquid was removed. The coated plate was blocked for 2 hours at room temperature with 200 μ L/well of PBS containing 2% (w/v) nonfat dry milk (Carnation, Glendale, CA) and 0.4% (w/v) Tween 80 (Calbiochem, San Diego, CA). The blocking solution was also used as a reagent diluent.

During the second hour of incubation, variable amounts of each of four test peptides were mixed with 22 μ L of ACA-6501 serum diluted with sample diluent in a final volume of 220 μ L using Eppendorf microcentrifuge tubes. Specifically, in tube #1, 188 μ L of sample diluent, 10 μ L of peptide stock solution (2 mg - 4 mg/mL diluent), and 22 μ L of AC-6501 serum diluted 1:35 in sample diluent were added. To tube #2, 158 μ L sample diluent, 40 μ L peptide stock solution and 22 μ L of 1:35 diluted ACA-6501 serum were added. Tube #3 contained 38 μ L sample diluent, 160 μ L peptide stock solution and 22 μ L of 1:35 diluted ACA-6501 serum. The control tube, tube #4, contained 396 μ L

sample diluent and 44 µL of 1:35 diluted ACA-6501 serum (no peptide). Each of the tubes was incubated for approximately 1 hours at room temperature.

The β_2 -GPI microplate was washed 5 times with TBS and 100 µL of the mixtures contained in Eppendorf tubes #1, 2, 3 and 4 was added to duplicate wells of the microplate. 100 µL of the contents of tube #4 was added to each of the duplicate control wells that were not coated with β_2 -GPI. The plate was incubated for 1 hour at room temperature, washed 5 times with TBS and 100 µL of 1:1000 goat anti-human IgG/AP conjugate (Zymed, Cat. No. 62-8422) diluted in sample diluent buffer was added. Following incubation for 1 hour at room temperature, the plate was again washed 5 times with TBS and the colorimetric enzyme detection developed by adding 100 µL PPMP solution (7.8 g phenolphthalein monophosphate and 69.5 g 2-amino-2-methyl-1-propanol in 100 mL water stock solution diluted 1:26 with water). After 22 minutes, the reaction was stopped with 50 µL of 0.2 M Na₂HPO₄ (Mallinckrodt) added per well. A_{550 nm} was read in a microplate reader (Bio-Tek Instruments, Model EL 311). Absorbance vs. peptide added was plotted on Graph Pad Prism (Graph Pad Software, Inc.) as shown in Figure 21. The amount of peptide that inhibited ACA-6501 binding by 50%, the IC-50, was calculated from the graph at the intersection of half-maximal absorbance with the amount of peptide added.

Example 5: Truncation Experiments of Peptide 3B10 and the Resulting Peptide Competition ELISA Results

Microtitration plates (96-well, flat bottom polystyrene, Immulon-2, Dynatech Laboratories, Inc., Chantilly, VA) were coated with 100 µL/well for 1 hour at RT with tetrameric ACA-6501/3B10 peptide at 10 µg/mL in carbonate buffer, pH 9.6 (15 mM Na₂CO₃/35 mM NaHCO₃). After the liquid from the wells was removed, each well was blocked for 1 hour at RT with 200 µL 0.5% (wt/vol) BSA (globulin-free, cat. no. A7638, Sigma Chemical Co., St. Louis, MO) in TBS. Three wells on the plate were left uncoated by tetravalent peptide to serve as blank control wells.

During the blocking step, each soluble, monomer peptide to be tested was set up in three test tubes (Eppendorf micro test tubes, Brinkmann Instruments, Westbury, NY) each containing 30 µL 5% BSA/TBS, 8.2 µL ACA-6501 serum (at 1:10 dilution with 0.5% BSA/TBS), plus a variable volume of the peptide/TBS stock and the

necessary volume of TBS buffer to yield a final volume of 330 µL. The 330 µL volume was sufficient to generate triplicate 100 µL samples for each peptide concentration that was tested for its ability to block ACA-6501 binding to the tetravalent peptide-coated plate. For peptide 139, which was truncated at the amino terminus and lacks the framework ala-gly contribution normally tested, the concentration of the stock solution was approximately 340-400 µg/mL TBS and aliquots of 19 µL, 75 µL, and 292 µL were removed to prepare the three peptide concentration tubes. For peptide 142 (lacking only the N-terminal ala) and peptide 143 (not truncated), stock solution concentrations were 400-500 µg/mL in TBS. Aliquots from each stock solution of 1 µL, 4 µL, and 16 µL were removed to set up the three concentration tubes for each peptide. For the peptide monomer control tube (lacking peptide), a tube with a final volume of 660 µL was prepared containing 60 µL 5% BSA, 16.5 µL of a 1:10 dilution of ACA-6501 and 583.5 µL TBS, *i.e.*, the same final concentrations (0.5% BSA and ACA-6501 serum at 1:400) as the 330 µL tubes but with twice the volume and without peptide.

Following the blocking incubation, the plate coated with tetravalent peptide was washed 5 times with TBS. From each 330 µL tube containing peptides 139, 142 and 143 at different concentrations, 100 µL was added to coated triplicate wells. Three 100 µL aliquots from the 660 µL control tube without peptide were each added to coated wells and three additional 100 µL aliquots were each added to uncoated blank wells. The plate was incubated at 40 rev/min on a rotary orbital shaker (Rotator V, American Dade, Miami, FL) for 1 hour at RT and then washed 5 times with TBS. One hundred µL/well of goat antihuman IgG/alkaline phosphatase conjugate (Cat. no. 62-8422, Zymed, South San Francisco, CA) diluted 1:1000 in BSA/TBS was added to the microplate. After incubation for 1 hour at RT, the plate again was washed 5 times with TBS. Color development followed the addition of 100 µL/well of freshly prepared dilute PPMP substrate solution. A dilute solution of PPMP (phenolphthalein monophosphate, cat. no. P-5758, Sigma Chemical Co., St. Louis, MO) was prepared by making a 1:26 dilution with water of the PPMP stock solution (0.13 M PPMP, 7.8 M amino-2-methyl-1-propanol adjusted to pH 10.15 with HCl). After 30 minutes, the reaction was stopped by adding 50 µL/well of 0.2M Na₂HPO₃ (reagent grade,

Mallinckrodt, St. Louis, MO). Absorbance measurements at 550 nm were carried out on a microplate reader (Bio-Tek Instruments, Winooski, VT) and the $A_{550\text{nm}}$ vs. peptide added per well results plotted using Graph Pad Prizm (Graph Pad Software, Inc., San Diego, CA). As shown on Figure 11, a horizontal line has been drawn corresponding to half-maximal the binding reaction obtained in the absence of monomer peptide competitive inhibition. The amount of peptide necessary for 50% inhibition can be read at the intersection of this line with the plot for each peptide tested. The 50% inhibition values are shown on Figure 12. The results indicate that while the loss of the N-terminal Ala had no negative consequences, the additional loss of Gly increased by about 8-fold the concentration necessary to achieve 50% inhibition.

Example 6: Substitution of alpha methyl proline into 3B10 and the resulting ELISAs

Testing of peptides ACA-6501/3B10 and analogs in which prolines at the 3 and 9 position were replaced by α -Me-Pro was carried out using the methodology described in Example 5. Peptides 3B10, 726 (α Me-Pro substituted at the 3 position), 727 (α Me-Pro substituted at the 9 position), and 728 (α Me-Pro substituted at both the 3 and 9 positions) were tested as soluble monomer peptides in a competitive- binding ELISA using tetramer peptide 3B10-coated microtitration plates and ACA-6501 serum.

Microtitration plates were coated with tetramer 3B10 peptide as described for Example 5. For each of the four peptides tested, three peptide concentrations were prepared in tubes. As in Example 5, these twelve tubes had final concentrations of 0.5% BSA/TBS and ACA-6501 serum at a final dilution of 1:400 in a final volume of 330 μL . All peptide stock solutions were at 400-500 $\mu\text{g}/\text{mL}$ TBS. To tubes containing 30 μL of 5% BSA/TBS and 8.2 μL of ACA-6501 serum (diluted 1:10 with 0.5% BSA/TBS), aliquots of 1 μL , 4 μL , or 16 μL of each of the four peptide stock solutions were added in addition to an appropriate volume of TBS to achieve a final volume of 330 μL . A control tube with no competitor peptide present was prepared with a final volume of 660 μL as described in Example 5.

Following the blocking incubation of the tetravalent peptide-coated plate, three 100 μL aliquots from each of the peptide concentration tubes for each of the four

peptides as well as the control tube containing no peptide and blank controls were tested as described in Example 5. The microtitration plate ELISA procedures as well as the data handling were performed as described in Example 5. As shown in Figure 13, peptide 727, where α -Me-Pro was substituted at the 9 position, was significantly more active than unmodified peptide 3B10 or the analog with both prolines changed (peptide 728). Peptide 726, which was substituted at position 3, lost activity as a result of the substitution.

Example 7: Abbreviated Description of Screen with 6626 Antibody and the Corresponding Sequences

Affinity purified ACA-6626 (AffACA-6626) was isolated by affinity purification from 8 mL of ACA-6626 plasma as previously described. AffACA-6626 (10 μ g) was incubated with the epitope $xy'z$ phage library consisting of a pool of all p-III component libraries in a final volume of 100 μ L as previously described for ACA-6501 biopanning. Following three rounds of biopanning, randomly selected phage from the second and third rounds were tested by micropanning. Only a few clones were weakly immunopositive at a 1:1000 dilution. An additional 4th round of biopanning was carried out. Micropanning of 94 fourth round clones revealed 43 immunopositives, some at phage dilutions as high as 1:100,000. G-Tracking DNA sequencing of the 43 immunopositive clones carried out as previously described for ACA-6501 revealed 5 unique sequences. After conventional four base DNA sequencing, the translated amino acid sequences of Table 3 were obtained.

Example 8: Identification of Sequences Specific for the ACA from Patient 6644

The epi $^{xy'z}$ phage display library was screened using methods similar to those in Example 4 with ACA affinity purified antibody from patient number 6644. A colony blot assay as described previously was employed as the final identification step prior to peptide synthesis. Approximately 150 colonies were plated on the original nitrocellulose membrane and assayed. Antibody from patient 6644 was used at a concentration of 1 μ g/mL. Of the 150 colonies plated on the nitrocellular membrane and assayed, only 4 were strongly positive and 2 weakly positive in this screen. Sequencing of the inserts of the six positive phage selected by this screen revealed

that the inserts were all derived from the 8-mer library with a free amino-terminus (epi^z):

Gly-Ile-Leu-Ala-Leu-Asp-Tyr-Val-Gly-Gly (3 inserts)

Gly-Ile Leu-Thr-Ile-Asp-Asn-Leu-Gly-Gly (1 insert)

Gly-Ile-Leu-Leu-Asn-Glu-Phe-Ala-Gly-Gly (2 inserts)

Example 9: Summary of Phage Library Screen with ACA-6641.

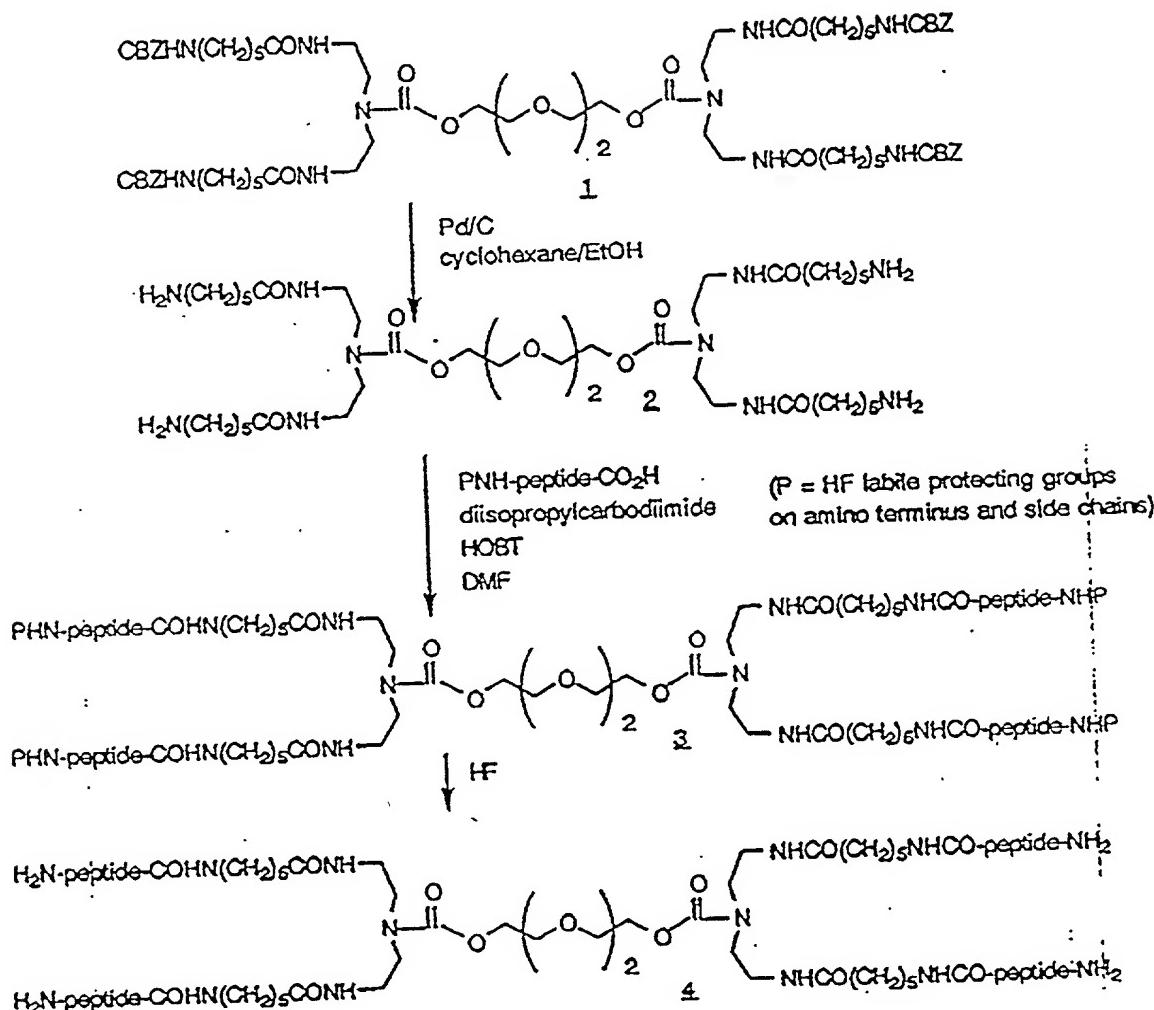
AffACA-6641 was isolated from 4 mL of plasma taken from patient number 6641. AffACA-6641 (10 µg) was incubated with the pooled p-III phage libraries in a final volume of 100 µL as described previously. Following four rounds of biopanning, 45 clones from the 3rd and 4th rounds were tested by micropanning. Of the 45, 23 scored negative. The 3rd round phage yielded two clones that scored 4+, two that scored 3+ and two that scored 2+. From the 4th round, one clone scored 4+, one scored 3+ and three scored 2+. G-tracking DNA sequencing revealed six unique sequences. Only one, clone 3G3, was strongly positive in the phage-capture ELISA. Four base DNA sequencing gave the following translated peptide sequence:

C L G V L G K L C.

Example 10: Peptide conjugation to non-immunogenic, multivalent carriers

Several tetravalent platforms for the development of B cell tolerogens have been developed as described in co-owned and co-pending U.S. Patent Applications, Serial Nos., 08/118, 055, filed September 8, 1993, 08/152,506, filed November 15, 1993, and U.S. Patents Nos. 5,268,454, 5,276,013 and 5,162,515 which are incorporated by reference herein in their entirety. Candidate peptides selected by aPL antibody screens of epitope libraries are conjugated and tested for modifications of immunochemical behavior such as antibody binding.

Non-immunogenic multivalent platforms with amine groups are synthesized as shown in the following scheme.

Amine on Platform - Carboxyl on Peptide

Compound 2: A solution of 8.0 g (5.7 mmol) of 1 in 50 mL of absolute EtOH and 35 mL of cyclohexene was placed under nitrogen, and 500 mg of 10% Pd on carbon was added. The mixture was refluxed with stirring for two hours. When cool, the mixture was filtered through Celite and concentrated to give 5.0 g of 2 as an oil. ^1H NMR (50/50 $\text{CDCl}_3/\text{CD}_3\text{OD}$) δ 1.21 (m, 8H), 1.49 (m, 8H), 1.62 (m, 8H), 2.19 (t, $J = 7.4$ Hz, 8H), 2.67 (t, $J = 7.4$ Hz, 8H), 3.36 (bd s, 16H), 3.67 (s, 4H), 3.71 (m, 4H), 4.21 (m, 4H).

Protected peptide with free carboxyl (PHN-peptide-CO₂H)

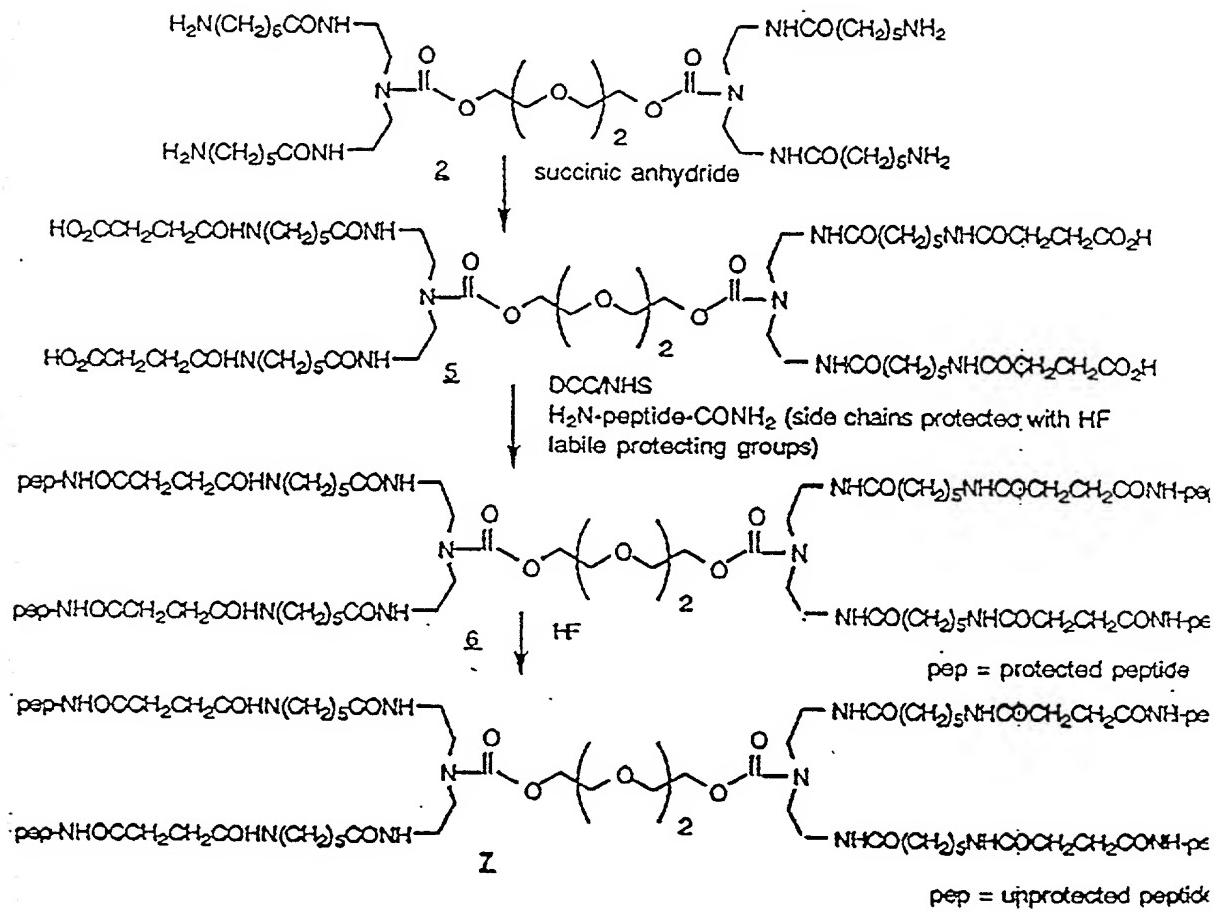
A peptide is synthesized with standard solid phase methods using FMOC chemistry on a Wang (p-alkoxybenzyl) resin, using trifluoroacetic acid (TFA) stable

protecting groups (benzyl ester or cyclohexyl ester on carboxyl groups and carbobenzyloxy (CBZ) on amino groups). Amino acid residues are added sequentially to the amino terminus. The peptide is removed from the resin with TFA to provide a peptide with one free carboxyl group at the carboxy terminus and all the other carboxyls and amines blocked. The protected peptide is purified by reverse phase HPLC.

Peptide - Platform Conjugate, 4

The protected peptide (0.3 mmol) is dissolved in 1 mL of dimethylformamide (DMF), and to the solution is added 0.3 mmol of diisopropylcarbodiimide and 0.3 mmol of 1-hydroxybenzotriazole hydrate (HOBT). The solution is added to a solution of 0.025 mmol tetraamino platform, 2 in 1 mL of DMF. When complete, the DMF is removed under vacuum to yield a crude fully protected conjugate 3. The conjugate, 3, is treated with hydrofluoric acid (HF) in the presence of anisole for 1 hour at 0° to give conjugate 4. Purification is accomplished by preparative reverse phase HPLC.

The following scheme shows the attachment of an amino group of a peptide to a carboxy group on a platform.

Carboxyl on Platform - Amine on Ligand

Compound 5 - Platform with Four Carboxylic Acid Groups

Succinic anhydride (1.0 g, 10 mmol) is added to a solution of 861 mg (1.0 mmol) of 2 and 252 mg (3.0 mmol) of NaHCO₃ in 20 mL of 1/1 dioxane/H₂O, and the mixture is stirred for 16 hours at RT. The mixture is acidified with 1N HCl and concentrated. The concentrate is purified by silica gel chromatography to provide 5.

Protected peptide with free amine (H₂N-peptide-CONH₂)

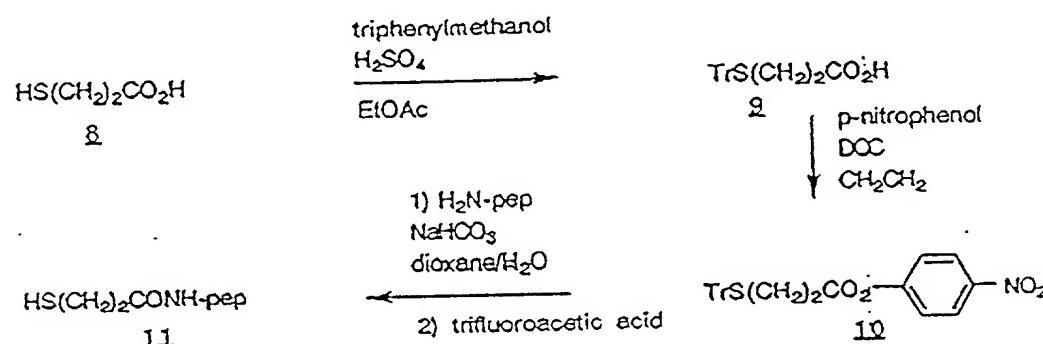
A peptide is synthesized with standard solid phase methods on an amide resin, which resulted in a carboxy terminal amide after cleavage from the resin, using TFA stable protecting groups (benzyl ester or cyclohexyl ester on carboxyl groups and CBZ on amino groups). Amino acid residues are added sequentially to the amino terminus using standard Fmoc chemistry. The peptide is removed from the resin with trifluoroacetic acid to provide a protected peptide with a free amine linker. The protected peptide is purified by reverse phase HPLC.

Peptide - Platform Conjugate, 7

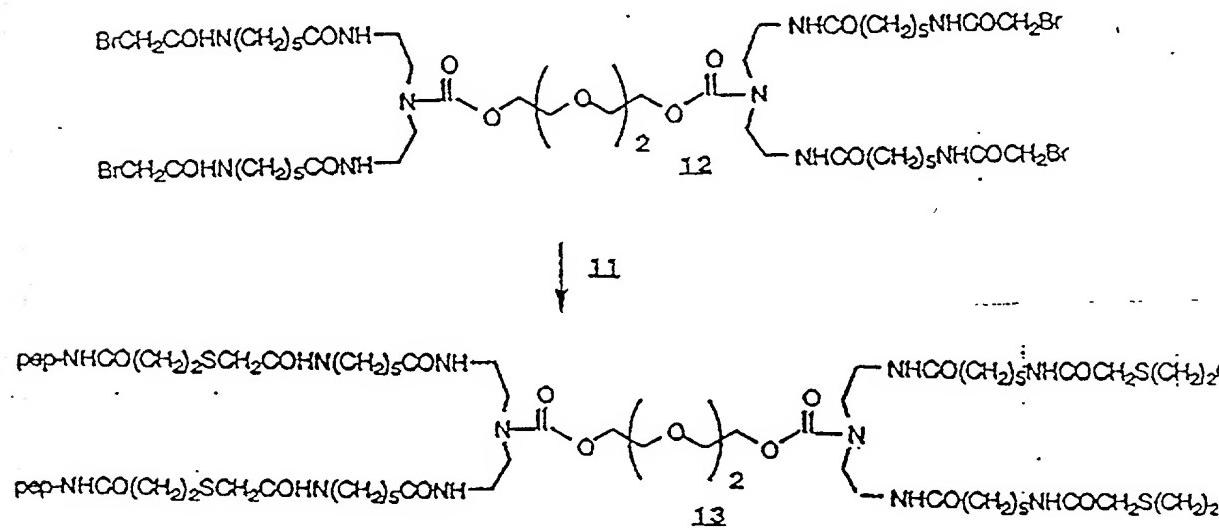
A solution of 0.05 mmol of protected peptide with free amine, (H₂N-peptide-CONH₂), 0.1 mmol of diisopropylethyl amine, and 0.01 mmol of 5 in 1 mL of DMF is prepared. BOP reagent (benzotriazol-1-yloxy-tris(dimethylamino)phosphonium hexafluorophosphate) (0.1 mmol) is added, and the mixture is stirred until the reaction is complete as evidenced by analytical HPLC. The peptide protecting groups are removed by treatment with HF in the presence of anisole at 0° to give conjugate with protecting groups removed, 7. Compound 7 is purified by preparative reverse phase HPLC.

The following scheme shows how to attach a sulphydryl linker to the amino terminus of a peptide and, in turn, attach the peptide/linker to a tetrabromoacetylated platform to give compound 13.

Haloacetyl on Platform and Sulfhydryl on Peptide



pep = cyclic non-disulfide peptide



Compound 9

Concentrated sulfuric acid (100 uL) was added to a 60°C solution of 4.48 g (17.2 mmol) of triphenyl methanol and 1.62 g (15.3 mmol, 1.3 mL) of 3-mercaptopropionic acid in 35 mL of EtOAc. The mixture was stirred at 60°C for 10 minutes, allowed to cool to room temperature (RT), and placed on ice for 1 hour. The resulting white solid was collected by filtration to give 4.52 g (75%) of 9.

Compound 10

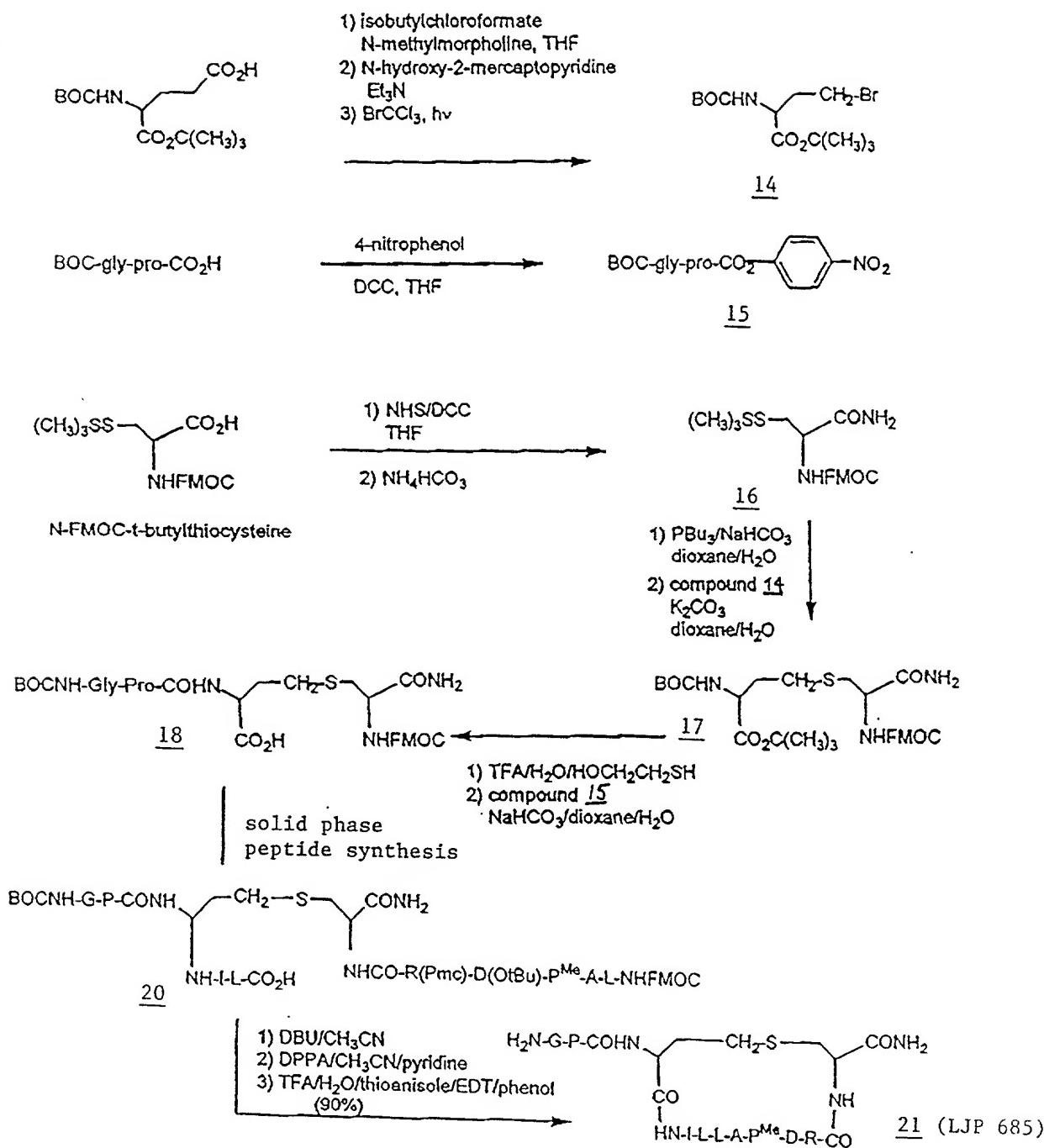
Dicyclohexyl carbodiimide (DCC) (2.41 g, 11.7 mmol) was added to a 0°C solution of 2.72 g (7.8 mmol) of 9 and 1.08 g (7.8 mmol) of p-nitrophenol in 41 mL of CH₂Cl₂. The mixture was stirred for 16 hours allowing it to come to RT. The mixture was filtered to remove *N,N*-dicyclohexylurea (DCU), and the filtrate was concentrated. The residue was crystallized from hexane/CH₂Cl₂ to give 3.17 g (86%) of 10 as pale yellow crystals.

Compound 11 - cyclic thioether peptide with mercaptopropionyl linker attached

A solution of a cyclic thioether peptide (an analogue of a disulfide cyclized peptide in which one sulfur was replaced with a CH₂) and sodium bicarbonate in water and dioxane was treated with p-nitrophenyl ester 10. If the peptide contains lysine, it must be appropriately blocked. The resulting modified peptide was treated with trifluoroacetic acid to provide thiol linker modified peptide 11.

Compound 13 - conjugate of cyclic thioether peptide and bromoacetylated platform

To a He sparged solution of 0.10 mmol of thiol modified peptide 11 in 100 mM sodium borate pH 9, was added 0.025 of bromoacetylated platform 12 as a 40 mg/mL solution in 9/1 MeOH/H₂O). The solution was allowed to stir under N₂ atmosphere until conjugation was complete as evidenced by HPLC. The conjugate was purified by reverse phase HPLC.

Example 12: Synthesis of LJP 685

γ -bromo-N-BOC- α -aminobutyric acid t-butyl ester, compound 14: A solution of 4.03 g (13.3 mmol) of N-BOC-glutamic acid- α -t-butyl ester and 1.61 mL (1.48 g, 14.6 mmol) of N-methylmorpholine in 40 mL of dry THF under N_2 atmosphere was cooled to 15°. Isobutylchloroformate (1.73 mL, 1.82 g, 13.3 mmol) was added to the mixture dropwise. The mixture was stirred for 10 minutes and a solution of 2.03 g (16.0 mmol) of N-hydroxy-2-mercaptopypyridine in 8 mL of THF was added followed by 2.23 mL (1.62 g, 16.0 mmol) of Et₃N. The mixture was covered with foil to keep out light and allowed to stir at room temperature for 1 hour. The mixture was filtered and the filtrate was concentrated on the rotary evaporator taking care to minimize exposure to light. The concentrate was dissolved in 20 mL of BrCCl₃ and the solution was cooled to -70°C. The solid was placed under vacuum, then purged with N_2 , allowed to come to room temperature, placed in a 20° water bath, and irradiated from above at close range with a 500W sunlamp for 5 minutes. The mixture was concentrated on the rotary evaporator and purified by silica gel chromatography (40 mm X 150 mm, toluene was used as eluent until UV active material finished eluting, 2% EtOAc/toluene (500 mL), 5% EtOAc/toluene (500 mL)). Impure fractions were repurified. Pure fractions by TLC (R_f 0.23, 5% EtOAc/toluene) were combined and concentrated to give 3.23 g (72%) of compound 14 as a waxy solid.

N-BOC-glycylproline-4-nitrophenyl ester, compound 15: A solution of 3.0 g (11.6 mmol) of N-BOC-glycylproline and 1.93 g (13.9 mmol) of 4-nitrophenol in 82 mL of dry THF was cooled to 0°C and 3.34 g (16.2 mmol) of DCC was added. The mixture was stirred at 0°C for 1 hour, the ice bath was removed, and the mixture was stirred for 16 hours at room temperature. HOAc (579 μ L) was added to the mixture and it was allowed to stir for 30 minutes. The mixture was kept in the freezer for 30 minutes and filtered under vacuum. The filtrate was concentrated and purified by silica gel chromatography (18 X 150 mm bed, 2.5% EtOAc/97.5% CH₂Cl₂/1%HOAc). Traces of acetic acid were removed by concentrating from dioxane several times on the rotary evaporator. The concentrate was triturated with 3/1 hexane Et₂O and the resulting white solid was collected by filtration to give 4.1 g (90%) of compound 15 as a white solid; TLC R_f 0.09, 40/60/1 EtOAc/hexane/HOAc; ¹H NMR(CDCl₃) δ 1.09-2.55 (m,

4H), 1.48 (s, 9H), 3.47-3.77 (m, 2H), 4.05 (m, 2H), 4.74 (m, 1H), 5.45 (bd s, 1H), 7.35 (d, 2H), 8.30 (d, 2H).

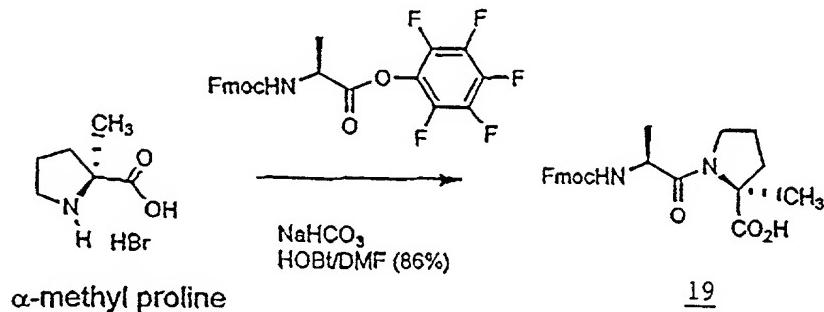
N-FMOC-S-t-butylthiocysteineamide, compound 16: A solution of 5.0 g (11.6 mmol) of FMOC-S-t-butylthiocysteine and 1.33 g (11.6 mmol) of N-hydroxysuccinimide in 115 mL of THF was cooled to 0°C. To the solution was added 3.58 g (17.37 mmol) of DCC. The mixture was stirred at 0°C for 1 hour and 42.9 mL of a solution of 1.6 g of (NH₄)HCO₃ in 50 mL of water was added. The mixture was stirred for 4.5 hours, allowing the ice bath to gradually warm to room temperature and concentrated on a rotary evaporator to remove THF and give an aqueous phase with white solid. The mixture was stirred with 200 mL of CH₂Cl₂ until most of the solid dissolved, then was shaken with 100 mL of 1N HCl solution. The CH₂Cl₂ layer was washed with 100 mL of saturated NaHCO₃ solution, dried (Na₂SO₄), and filtered. The filtrate was brought to a boil on a hot plate and crystallized from 300 mL of CH₂Cl₂/hexane to give 4.36 g (87%) of compound 16 as a white solid: mp 127°-129°C; TLC R_f 0.29, 95/5/1 CH₂Cl₂/CH₃CN/MeOH, ¹H NMR (CDCl₃) δ 1.36 (s, 9H), 3.06-3.24 (m, 2H), 4.25 (t, 1H), 4.55 (m, 3H), 5.56 (bd s, 1H), 5.70 (bd s, 1H), 6.23 (bd s, 1H); ¹³C NMR (CDCl₃) 29.8, 41.9, 47.1, 48.5, 54.2, 67.2, 120.0, 125.0, 127.1, 127.8, 141.3, 143.6, 156.1, 172.2. Analysis, Calculated: C, 61.4%; H, 6.1%; N, 6.5%. Found: C, 61.5%; H, 6.0%; N, 6.5%.

Compound 17: Water (16.9 mL) was added to 2.91 g (6.75 mmol) of compound 16 in 33.8 mL of dioxane and the solution was sparged with nitrogen for 5-10 minutes. The mixture was kept under a nitrogen atmosphere and 1.13 g (13.5 mmol) of NaHCO₃ was added followed by 1.77 mL (1.44 g, 7.09 mmol) of PBu₃. The mixture was stirred at room temperature for 1 hour and partitioned between 1 X 100 mL of 1N HCl and 2 X 100 mL of CH₂Cl₂. The CH₂Cl₂ layers were combined and concentrated and the resulting white solid was partially dissolved in 33.8 mL of dioxane. Water (9 mL) was added and the mixture was purged with nitrogen for 5-10 minutes. The mixture was kept under nitrogen atmosphere and 2.17 g (16.9 mmol) of K₂CO₃ was added followed by 2.63 g (8.1 mmol) of compound 14. The mixture was stirred for 16 hours and partitioned between 1 X 100 mL of 1N HCl and 2 X 100 mL of 10%

MeOH/CH₂Cl₂. The CH₂Cl₂ layers were combined, dried with NaSO₄, filtered and concentrated to a semisolid residue. Purification by silica gel chromatography (1/3 CH₃CN/CH₂Cl₂) gave 3.2 g (80%) of compound 17 as a white solid; TLC R_f 0.27, 1/3 CH₃CN/CH₂Cl₂. Further purification of an analytical sample was done by recrystallizing from hexane/EtOAc; mp 104-106.5°C; ¹H NMR (CDCl₃) δ 1.47 (s, 9H), 1.49 (s, 9H), 1.96 (m, 1H), 2.11 (m, 1H), 2.69 (m, 2H), 2.88 (m, 1H), 3.03 (m, 1H), 4.29 (t, 1H), 4.36 (m, 2H), 4.50 (m, 2H), 5.21 (m, 1H), 5.49 (m, 1H), 5.81 (m, 1H), 6.54 (m, 1H), 7.34 (t, 2H), 7.42 (t, 2H), 7.61 (d, 2H), 7.80 (d, 2H); ¹³C NMR (MeOH) δ 26.1, 26.7, 28.2, 28.7, 34.8, 54.8, 55.7, 68.1, 80.5, 82.7, 120.9, 126.3, 128.2, 128.8, 142.6, 145.2, 160.0, 162.7, 173.4, 175.8. Analysis, Calculated for C₃₁H₄₁N₃O₇S: C, 62.08; H, 6.89; N, 7.01. Found: C, 62.23; H, 7.12; N, 7.39.

Compound 18: A solution of 20/1/1 TFA/H₂O/mercaptoethanol (23.2 mL) was added to 1.27 g (2.11 mmol) of compound 17. The mixture was stirred at room temperature for 1 hour and concentrated to a volume of about 3 mL. 50 mL of ether was added to precipitate the product. The resulting solid was washed with two more portions of ether and dried under vacuum to give 812 mg of solid. The solid was suspended in 10.6 mL of dioxane and 10.6 mL of H₂O. NaHCO₃ (355 mg, 4.22 mmol) was added to the mixture followed by a solution of 1.33 g (3.38 mmol) of compound 15 in 10.5 mL of dioxane. The mixture was allowed to stir at room temperature for 20 hours and partitioned between 100 mL of 1N HCl and 3 X 100 mL of CH₂Cl₂. The combined CH₂Cl₂ layers were dried (Na₂SO₄), filtered and concentrated. Purification by silica gel chromatography (35 X 150 mm; step gradient, 5/95/1 MeOH/CH₂Cl₂/HOAc (1 L) to 10/95/1). Pure fractions were concentrated and the residue was triturated with ether to give 881 mg (60%) of compound 18; TLC R_f 0.59, 10/90/1 MeOH/CH₂Cl₂/HOAc; mp 116-117.5°C; ¹H NMR (CDCl₃) δ 1.41 (s, 9H), 2.00 (m, 2H), 2.18 (m, 2H), 2.56 (m, 1H), 2.69 (m, 1H), 2.85 (m, 2H), 3.49 (m, 2H), 3.62 (m, 2H), 3.85 (m, 2H), 4.08 (m, 1H), 4.12 (m, 1H), 4.22 (t, 1H), 4.38 (m, 1H), 4.49 (m, 2H), 4.61 (m, 2H), 7.78 (d, bd s, 1H), 6.12 (bd s, 1H), 6.43 (bd s, 1H), 7.35 (d, 2H), 7.40 (d, 2H), 7.61 (d, 2H), 7.78 (d, 2H); ¹³C NMR (CDCl₃) δ 25.3, 27.8, 28.6, 29.4, 32.6, 35.1, 44.1, 46.9, 47.3, 52.2, 53.9, 61.2, 67.8, 80.8, 120.8, 125.7, 128.2, 129.0,

$\text{C}_{34}\text{H}_{43}\text{N}_5\text{O}_9\text{S}$: C, 58.52; H, 6.21; N, 10.03. Found: C, 58.38, H, 6.17; N, 10.20.



N-FMOC-L-Alanyl-L-2-methylproline, compound 19: A solution of 2-methylproline (Seebach *et al.* (1983) *J. Am. Chem. Soc.* **105**:5390-5398) (1.00 g, 4.76 mmol), 4.00 g (47.6 mmol) of NaHCO₃, and 31 mg (0.23 mmol) of HOBT in 6.9 mL of DMF was cooled to 0°C and 3.18 g (6.66 mmol) of N-FMOC-L-alanine was added. The reaction was stirred for 1 hour at 0°C, then at room temperature for 18 hours. The mixture was partitioned between 50 mL of EtOAc and 3 X 50 mL of 1N HCl. The EtOAc layer was dried (MgSO₄), filtered and concentrated. Purification by silica gel chromatography (step gradient 45/55/1 EtOAc/Hexane/HOAc to 47/53/1 EtOAc/Hexane/HOAc to 50/50/1 EtOAc/Hexane/HOAc) gave 1.72 g (86%) of compound 19 as a white solid. The product was concentrated several times from dioxane to remove traces of acetic acid: mp 59-60°C; ¹H NMR (CDCl₃) δ 1.39(d, 3H), 1.93 (m, 2H), 2.06 (m, 2H), 3.78 (m, 2H), 4.22 (m, 1H), 4.40 (d, 2H), 4.56 (m, 1H), 5.09 (t, 1H), 5.69 (d, 1H), 7.32 (t, 2H), 7.42 (t, 2H), 7.62 (d, 2H), 7.78 (d, 2H); ¹³C NMR (CDCl₃) δ 17.8, 21.8, 23.8, 37.9, 47.1, 48.5, 65.9, 67.0, 120.0, 125.1, 127.1, 127.7, 141.3, 144.1, 155.6, 172.9, 175.1.

N-FMOC-L-Leucinyl-HMPB-MBHA resin: A solution of N-FMOC-L-leucine in 22.5 mL of CH_2Cl_2 and a few drops of DMF was prepared and cooled to 0°C. To the solution was added 1.71 mL (1.38 g, 10.9 mmol) of diisopropylcarbodiimide (DIC) and the mixture was stirred for 20 minutes at 0°C. The mixture was concentrated to an oil; meanwhile, enough DMF (approximately 3 mL) was added to 2.5 g (0.87 mmol/g, 2.18 mmol) HMPB-MBHA resin (Nova Biochem) to swell the resin. The concentrated oil was dissolved in a minimal amount of DMF (approximately 1 mL)

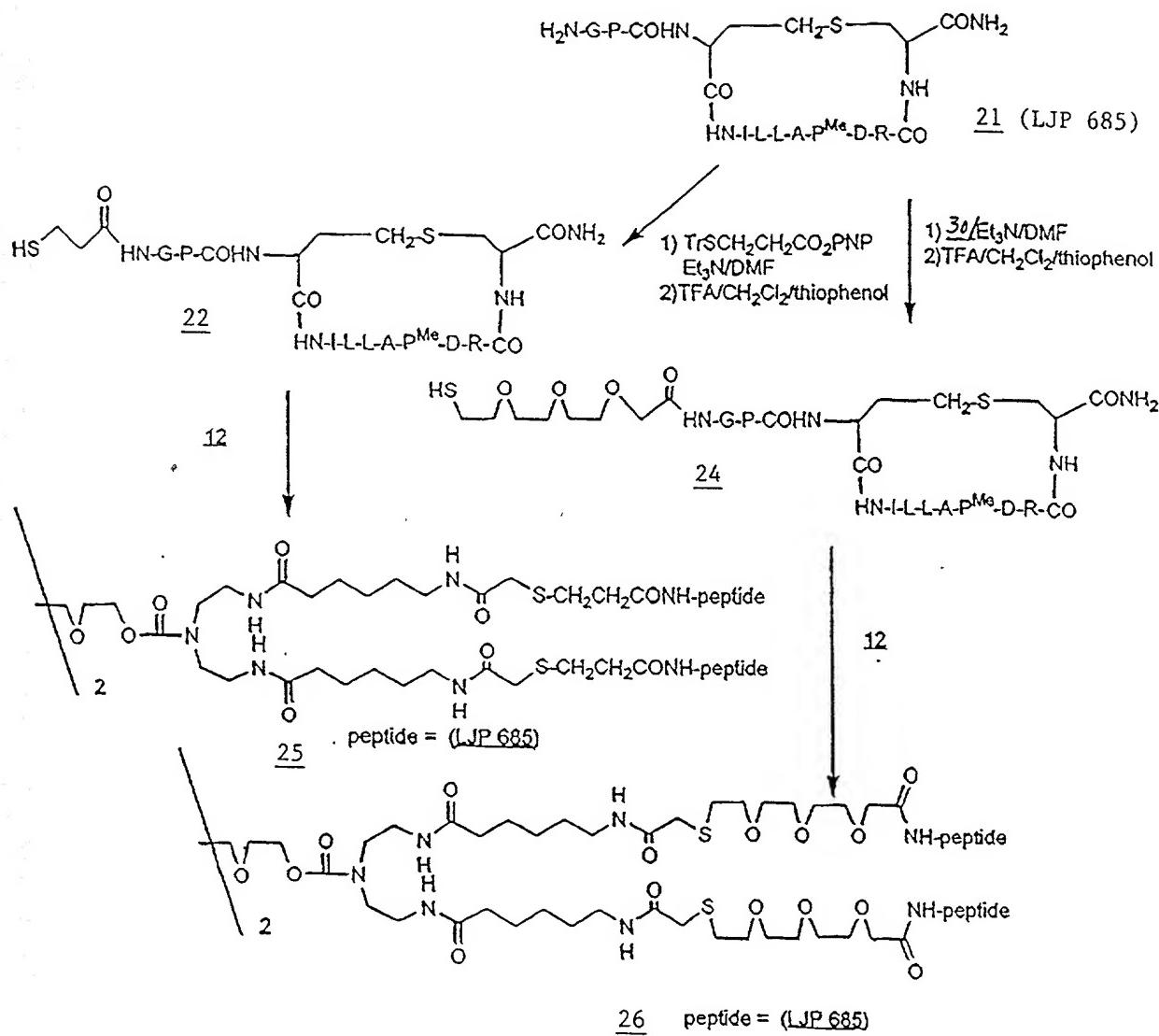
and added to the swelled resin followed by a solution of 266 mg (2.18 mmol) of DMAP dissolved in approximately 1 mL of DMF. The mixture was gently rocked for 1 hour and washed (2 X DMF, 2 X MeOH, 2 X DMF, 2 X MeOH). The resin was dried under vacuum to give 2.77 g (85%) and the substitution was determined by the Geisen test to be 0.540 mmol/g.

N-FMOC-linear peptide with t-butyl ester on aspartic acid and Pmc group on arginine and with thioether insert, compound 20: This peptide was prepared by standard FMOC synthesis on N-FMOC-L-leucinyl-HMPB-MBHA resin. Three equivalents of amino acid, HOBT and (DIC) were used for each coupling step with the exception of the coupling step of compound 18. Two equivalents of compound 18 were used with three equivalents of HOBT and diisopropylcarbodiimide. Each step was monitored by using 10 μ L of bromophenol blue indicator. Completeness of the reaction was also assessed with a ninhydrin test (beads turn blue for incomplete reaction with 1 mg is heated at 100°C for 2 minutes with one drop of pyridine and one drop of 5% ninhydrin in EtOH and one drop of 80% phenol in EtOH). Thus, 1.13 mg (0.613 mmol) of resin was used to prepare the peptide. After the final coupling step, cleavage from the resin was accomplished by treatment with 15 mL of a solution of 1% trifluoroacetic acid in CH_2Cl_2 for 2 minutes. After 2 minutes, the solution was filtered under pressure into 30 mL of 10% pyridine in MeOH. This was repeated ten times and the filtrates which contained peptide as evidenced by HPLC (C18, gradient, 60/40/0.1 $\text{CH}_3\text{CH}/\text{H}_2\text{O}/\text{TFA}$ to 90/10/0.1 $\text{CH}_3\text{CN}/\text{H}_2\text{O}/\text{TFA}$, 210 mn, 1 mL/min, 4.6 mm X 250 mm column) were combined and concentrated. The concentrate was dissolved in 40 mL of 10% HOAc solution and purified by HPLC to give 0.528 g (49%) of peptide 20.

Conversion of peptide 20 to cyclic peptide 21 (removal of FMOC group, cyclization and removal of protecting groups: A solution (5 mL) of 99 μ L of DBU in 10 mL of CH_3CN was added to 96 mg (0.052 mmol) of peptide 20. The solution was stirred for 1 hour and concentrated to a residue. The residue was triturated with 2 X 10 mL of Et_2O to give a white solid. The solid was dissolved in 100 mL of CH_3CN and 312 μ L (0.312 mmol) of 0.1 M solution of diphenylphosphorylazide (DPPA) in CH_3CN . The

mixture was stirred for 20 hours and concentrated on a rotary evaporator. The residue was triturated with 2 X 50 mL of Et₂O and the white opaque residue was treated with 5 mL of 92/3/2/3 TFA/anisole/EDT/Me₂S for 1 hour. The product was precipitated by adding the mixture to 40 mL of Et₂O in a 50 mL polypropylene centrifuge tube. The precipitate was cooled to 0°C and centrifuged for 5 minutes at 2000 rpm. The supernatant was decanted and the pellet was washed with Et₂O and recentrifuged. The pellet was dried and dissolved in 4 mL of 50/50 CH₃CN/H₂O. The mixture was diluted with 36 mL of H₂O/0.1% TFA, filtered and purified by HPLC (1" C₁₈ column, gradient, 10/90/0/1 CH₃CN/H₂O/TFA to 35/65/0.1 CH₃CN/H₂O/TFA, 230 nm). The pure fractions, as evidenced by HPLC, were lyophilized to give 52 mg (90%) of compound 21.

Example 13: Synthesis of conjugates of LJP 685



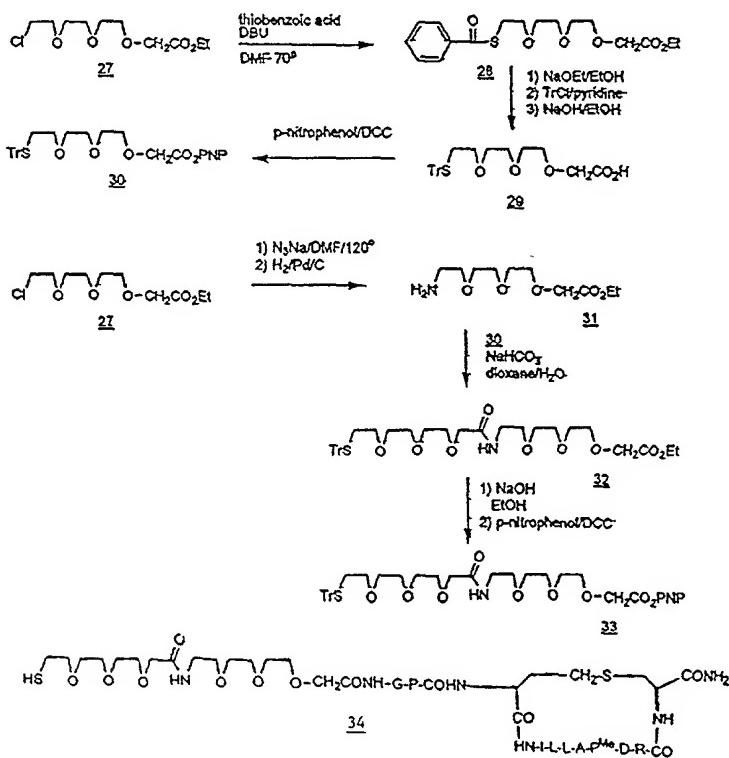
LJP 685, also referred to as compound 21, was treated with the PNP ester of 3-tritylmercaptopropionic acid and the resulting product was detritylated to give compound 22, the peptide with the free thiol linker. Reaction of an excess of compound 22 with valency platform molecule 12 produced tetravalent conjugate 25. Treatment of compound 21 with the longer linker, compound 33 (see reaction scheme below), followed by detritylation, gave compound 24. Compound 24 reacted with valency platform molecule 12 to give the tetraconjugate 26. Both conjugation reactions appeared very clean by HPLC.

Attachment of MTU linker to LJP 685, synthesis of compound 24: To 15 mg (0.013 mmol) of compound 21 was added 160 μ L of a solution of 29.5 mg of compound 23 and 17.5 μ L of diisopropylethylamine in 0.5 mL of DMF. the mixture was stirred for 2 hours and precipitated from Et₂O. The precipitate was dried and dissolved in 650 μ L of a solution of 1/1/0.056/0.040 TFA/CH₂Cl₂/thiophenol/Me₂S and the solution was allowed to stand for 1 hour. Precipitation from Et₂O gave crude compound 24 which was purified by HPLC (C₁₈, 15-45% CH₃CN/H₂O, 0.1% TFA). Fractions containing pure product were lyophilized to give 8.6 mg of compound 24 as a white solid.

LJP 685-MTU-AHAB-TEG, compound 26: To a solution of 8.6 mg (6.3×10^{-6} mol) of compound 24 in 630 μ L of He sparged pH 8.5 200 mM borate buffer was 40 μ L of a 40 mg/mL solution of compound 12 in 9/1 MeOH/H₂O. The mixture was stirred for 24 hours and 1 mL of 10% HOAc/H₂O solution was added. The mixture was purified by HPLC (C₁₈, gradient 25-55% CH₃CN/H₂O, 0.1% TFA) to give 8.3 mg of compound 26 (LJP 685-MTU-AHAB-TEG).

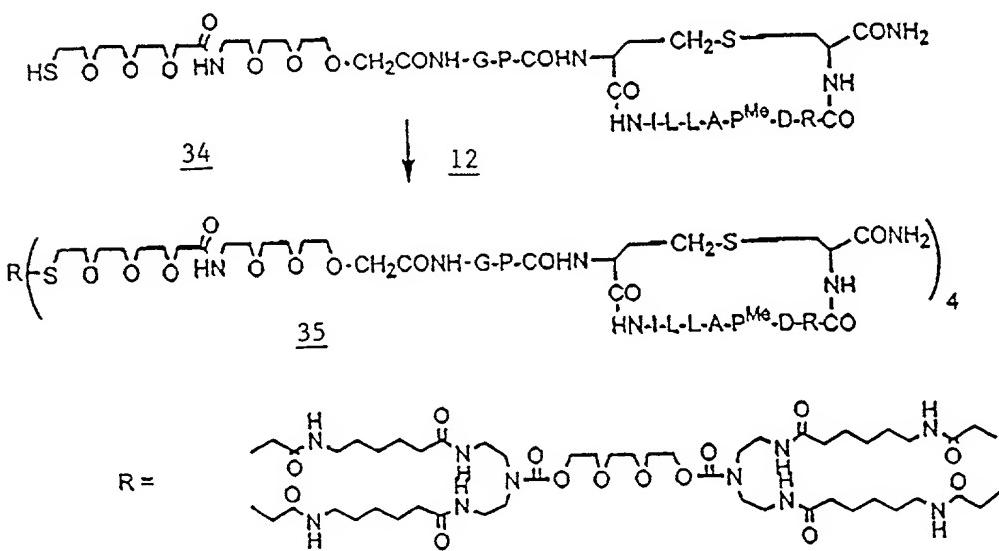
Example 14: Development of extended SH linkers

The thiobenzoate ester, compound 28, was prepared from compound 27. Compound 28 was converted to compound 29 in portions. The thiobenzoate was removed by ethanolysis and the resulting thiol was tritylated. The ethyl ester was then hydrolyzed to give compound 29. The nitrophenyl phosphate (PNP) ester, compound 30, was prepared from compound 29. Aminotrioxoudecanoicacid ethylester, compound 31, was prepared by treatment of compound 27 with sodium azide and reduction to the amine. Amine 31 was acylated with compound 30 to provide compound 32. Hydrolysis of compound 32 was achieved by treatment with sodium hydroxide to give a free carboxylic acid. The intermediate carboxylic acid was condensed with p-nitrophenol to give para-nitrophenyl (PNP) ester, compound 33. Linker 33 was attached to the peptide and the trityl group removed to give compound 34 which was used to produce an MTU-ATU-AHAB-TEG conjugate.



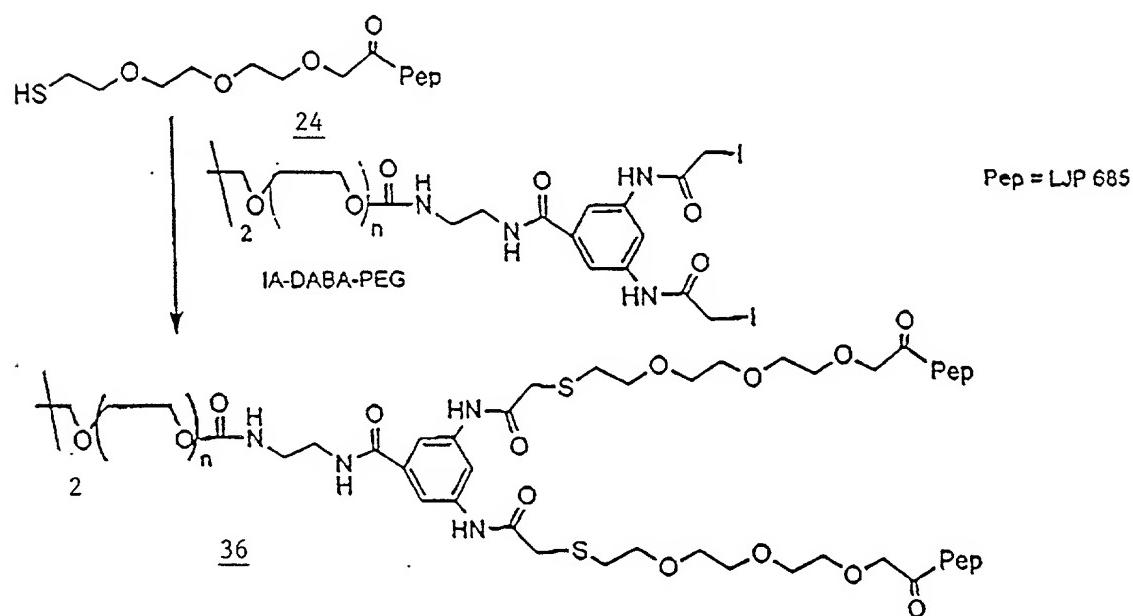
Example 15: Synthesis of a $(\text{LJP685})_4/\text{MTU-ATU-AHAB-TEG}$ conjugate, compound 35

Tetravalent conjugate 35 was prepared as shown below. The peptide with linkers attached, compound 34 was dissolved in He sparged, pH 8.5, 200 mM borate buffer. To the mixture was added 0.3 mol equivalents of platform compound 12. The mixture was stirred for 1 hour and the product was purified by HPLC.



Example 16 Synthesis of a (LJP685)₄/DABA-PEG conjugate, compound 36

Treatment of IA-DABA-PEG with compound 24 in 8.5 borate buffer gave conjugate 36.



Example 17 T cell assay for activation

Tritiated thymidine uptake by peptide-stimulated T cells was monitored in 96-well round bottom plates. A single-cell suspension of draining lymph node cells (mice) or isolated peripheral blood lymphocytes (human), 5×10^5 were mixed with between 1 and 30 μg of peptide in a final volume of 150 μL per well and incubated for 5 days at 37°C in 5% CO₂. At that point, 1 micro curie of labeled thymidine was added and incubated for an additional 15-24 hours. The harvested cells were collected on filters and counted by liquid scintillation spectrometry.

Example 18 In vitro induction of tolerance

Eight groups, each containing five C57B1/6 mice, were primed with 10 $\mu\text{g}/\text{mouse}$ of a conjugate of LJP685-KLH on alum plus *B. pertussis* vaccine as an adjuvant. After three weeks, spleen were harvested and single cells suspensions were prepared, washed three times with balanced salt solution and resuspended in complete RPMI-1640 medium at a concentration equivalent to one spleen/1.5 mL of medium. The cell suspension was divided into aliquots of 2.5 mL /petri dish and incubated for 2 hours at 37°C with (LJP685)₄-DABA-PEG, compound 36, and (LJP685)₄-TEG, compound 35, in concentrations of 100 μM , 20 μM and 4 μM . One group of cells was incubated without toleragen and acted as the positive control. The cells were then washed with large volumes of balanced salt solution and resuspended in 2.5 mL of balanced salt solution. The cells were then injected into 650R irradiated syngeneic recipient mice in such a manner that all of the cells from a given treatment group were divided evenly into five recipients. All of the recipient mice, including the positive controls, were then given a booster immunization of 10 μg of LJP 685-KLH in saline, intraperitoneally. Seven days after the booster immunization, the mice were bled and their sera tested for the presence of anti-LJP 685 antibody. Treatment with either conjugate produced a significant, dose-dependent reduction of anti-LJP 685 antibodies as shown in Figures 16 and 17, which is measured by Antigen Binding Capacity (ABC) as described in Iverson, G.M., "Assay for *in vivo* adoptive immune responses," in HANDBOOK OF EXPERIMENTAL IMMUNOLOGY, Volume 2, Cellular Immunology (Weir, D.M., ed., Blackwell Scientific Publications, Palo Alto, CA, 1986).

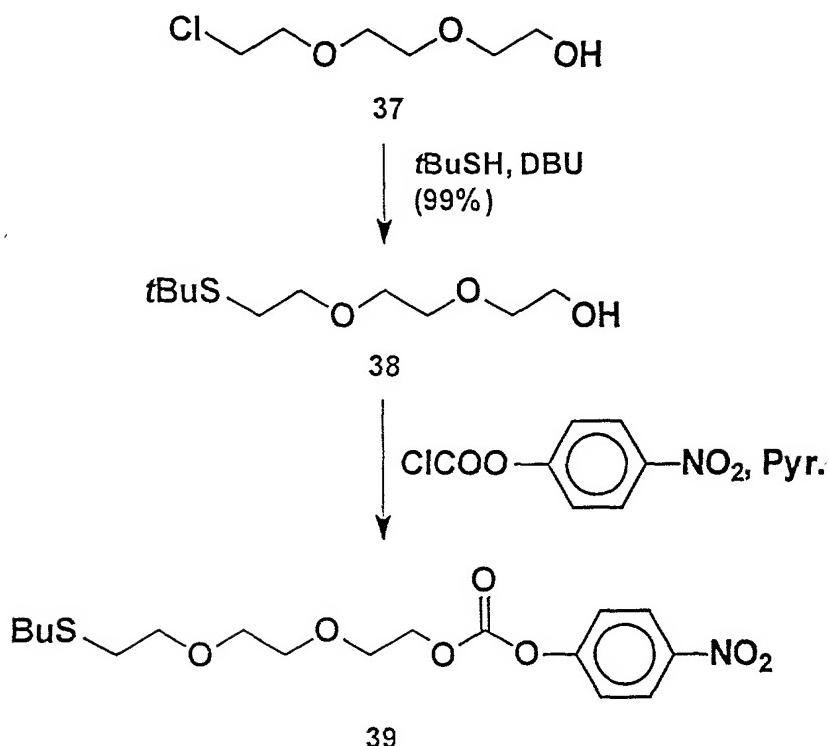
Example 19 NMR solution structure analysis of the 5A12, CB2 and 3G3 peptides

Two peptides isolated from phage library screens using the methodology described in the examples above were subjected to NMR analysis. The original peptide has a proline in the second to last position, however, this amino acid was removed since two dimensional (2D) double quantum filtered correlated spectroscopy (DQF-COSY) NMR data suggested the presence of two different structures from the *cis* and *trans* isomers at this position. Removal of the proline gave peptides with K_D 's in the 50-100 nM range for binding to ACA antibodies, and the expected number of peaks in the fingerprint region of the DQF-COSY spectrum. The resulting cyclic peptides are 5A12 (GPCLILAPDRCG) AND CB2 (GPCILLARDRCG). The major difference between the peptides was the substitution of arginine for proline at position 8 which resulted in much less dispersion in the 1D ^1H NMR spectrum of CB2 which was consistent with 5A12 being a more rigid peptide. The arginine substitution also produces a 0.55 kcal/mol stabilization of the ionized aspartyl carboxy group as reflected in pKa values. A structural analysis was carried out on the more ordered 5A12 peptide. Although deuterium exchange and temperature coefficient data show no evidence of hydrogen bonding, the Nuclear Overhauser Effect (NOE), Rotating Overhauser Effect (ROE) and coupling data are consistent with one structure. Distance geometry calculations yielded a family of 50 structures. The 15 best had a mean root mean square deviation (RMSD) for all atoms of 2.1 ± 0.2 angstroms. We determined that the peptide has an oval shape with turns at opposite ends of the molecule, at the disulfide and at Proline 8 which is *cis*. There is also a kink in the backbone LIL region. Finally, the carboxy terminal glycine is very mobile as it is the only residue with a positive intraresidue NOE. This represents the first structural information determined about a peptide that mimics the ACA epitope on β -2-GPI.

NMR data coupled with distance geometry calculations were used to determine the three dimensional structure of peptide 925 (CLGVLA KLC), a truncated version of peptide 3G3 (AGPCLGVLGKLC PG) with alanine substituted for glycine in position 6 of the 925 peptide. The structure of peptide 925 was determined in water at pH 3.8 and at 25 °C. An ensemble of nine structures were calculated all of which were consistent with the NMR data. The RMSD for all non-hydrogen atoms was 2.45

∓ 0.36 angstroms when each structure was compared to the centroid. Figure 18 displays the structure closest to the centroid of the ensemble and, therefore, is a reasonable representation of the shape of the peptide 925 molecule. Figure 19 compares the structure of peptide 925 (labeled at the bottom of the figure as 3G3) with the structure of peptide 5A12. Both peptides have turns at approximately the same positions in the peptide sequence.

The pharmacophore of the peptides has been tentatively identified as a small hydrophobic group and a positively charged group. The gem-dimethyl and amino groups of peptide 925 are tentatively identified as the pharmacophore of this peptide as shown in Figure 20. The hydrocarbon linkers that tether the pharmacophore groups to some scaffold have the lengths specified in Figure 20 and the points at which these linkers are attached to the scaffold are separated by the distance specified. Finally, the dihedral angle defining the relative orientation of the two linkers was determined to be 22° .

Example 20 Synthesis of TEG carbamate linker

2-[2-(2-*tert*-Butylthioethoxy)ethoxy]ethanol, compound 38: To a mixture of 2-[2-(2-chloroethoxy)ethoxy]ethanol (11.0 g, 65.24 mmol) and *tert*-butylthiol (7.35 mL, 65.23 mmol) cooled in an ice bath was slowly added 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 9.75 mL, 65.23 mmol). The reaction mixture was allowed to warm to room temperature and stirred overnight. This mixture was then diluted with ethyl acetate and filtered. The crude product in the filtrate was purified on a filter column eluted with ethyl acetate to give yellowish oil (14.4 g, 64.6 mmol, 99%): ¹H NMR (300 MHz, CDCl₃): δ 3.73 (br s, 2H), 3.69-3.60 (m, 8H), 2.75 (t, *J* = 7.4, 2H), 2.62 (br s, 1H), 1.32 (s, 9H); ¹³C NMR (75 MHz, CDCl₃): δ 72.47, 71.08, 70.33, 70.27, 61.71, 42.09, 30.95, 27.87; MS (ESI): m/e (M+1) Calcd. for C₁₀H₂₃O₃S: 223, obsd.: 223.

2-[2-(2-*tert*-Butylthioethoxy)ethoxy]ethyl *p*-nitrophenyl carbonate, compound 39: To a solution of compound 38 (2.0 g, 8.98 mmol) and *p*-nitrophenol chloroformate (1.81 g, 8.98 mmol) in 5 mL of dry THF cooled in an ice bath was slowly added dry pyridine (0.73 mL, 8.98 mmol) in 1 mL of dry THF. White precipitate came out immediately. After stirred at room temperature for 15 min, the reaction mixture was diluted with 10 mL

of ether and filtered. The filtrate was concentrated and used directly in the peptide synthesis without further purification.

Example 21 Synthesis of the Tetravalent Platform IA/DABA/ATEG, 46.

Bis-N-(t-butoxycarbonyl)-diaminobenzoic acid, compound 40: A solution of 7.18 g (32.9 mmol) of di-t-butylidicarbonate in 5.5 mL of MeOH was slowly added to a solution of 2.5 g (16.4 mmol) of 3,5-diaminobenzoic acid and 2.76 g (32.9 mmol) of NaHCO₃ in 44.5 mL of H₂O and 22.5 mL of MeOH, and the mixture was stirred at room temperature for 24 h. The mixture was cooled to 0°, and 6.53 g of citric acid was added, and the mixture was extracted with EtOAc. The combined EtOAc layers were dried (MgSO₄), filtered, and concentrated. The residue was dissolved in 40 mL of Et₂O and the solution was filtered through Celite. The Et₂O layer was extracted with two 40 mL portions of HCl. The Et₂O layer was dried (MgSO₄), filtered, and concentrated to give 3.81 g (66%) of 40 as a foamy pink solid.

N-hydroxysuccinimidyl ester of compound 40, compound 41: Dicyclohexylcarbodiimide (3.34 g, 16.2 mmol) was added to a solution of 3.8 g (10.8 mmol) of compound 40 and 1.24 g (10.8 mmol) of N-hydroxysuccinimide in 55 mL of EtOAc which had been cooled to 0°, and the resulting mixture was stirred for 18 h allowing to come to room temperature. To the mixture was added 0.55 mL of acetic acid. The mixture was stirred for 30 min and placed in the freezer for 2 h. The mixture was filtered to remove solids, and the filtrate was concentrated to give 5.80 g of pink foamy solid. Purification by silica gel chromatography (60/40/1 hexane/EtOAc/HOAc) gave 4.30 g (89%) of compound 41 as a slightly pink solid.

Mono-N-(t-butoxycarbonyl)-ethylenediamine, compound 42: A solution of 1.5 g (25.0 mmol) of ethylenediamine in 15 mL of CH₂Cl₂ was cooled to 0°, and a solution of 1.82 g (8.33 mmol) of di-t-butylidicarbonate was added slowly to the mixture. The mixture was stirred at room temperature for 18 h and filtered, and the filtrate was concentrated. Purification by silica gel chromatography (90/10/1 CH₂Cl₂/MeOH/HOAc) gave 0.98 g (67%) of compound 42 as an oil.

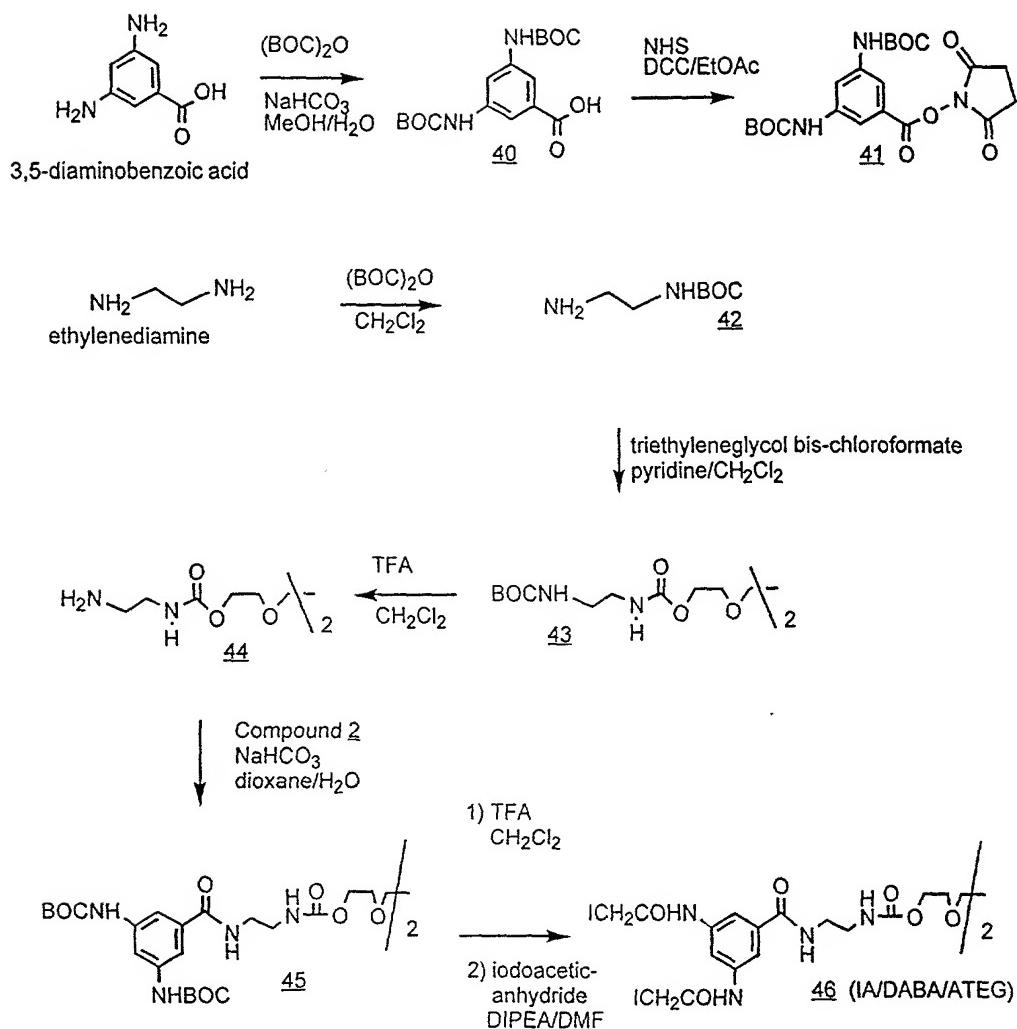
Bis- N-(t-butoxycarbonyl)-amino-TEG, compound 43: To a solution of 750 mg (4.25 mmol) of compound 42 and 345 uL (337 mg, 4.25 mmol) of pyridine in 6 mL of CH₂Cl₂ was added 445 uL (559 mg, 2.02 mmol) of triethyleneglycol bis-chloroformate. The mixture was stirred for 3.5 h,

and the mixture was partitioned between 35 mL of CH_2Cl_2 and 35 mL of 1 N HCl. The CH_2Cl_2 layer was washed with H_2O , dried (Na_2SO_4), filtered and concentrated to give 1.14 g of crude compound 43 which was used directly in the next step.

Diamino-TEG bis-trifluoroacetate salt, compound 44: 300 mg (0.57 mmol) of compound 43 was dissolved in 3.5 g of CH_2Cl_2 , and 3.5 mL of trifluoroacetic acid was added. The mixture was stirred for 3 h at room temperature, and the solution was concentrated to give 398 mg of crude compound 44 which was used directly in the next step.

Compound 45: A solution of 567 mg of compound 41 in 6 mL of dioxane was added to a solution of 398 mg of crude compound 44 (est. 316 mg, 0.57 mmol) and 193 mg (2.30 mmol) of NaHCO_3 . The mixture was stirred for 3 h and acidified with 1 N HCl and partitioned between 20 mL of 1 N HCl and 30 mL of EtOAc. The combined EtOAc layers are washed with sat NaHCO_3 solution, dried (MgSO_4), filtered and concentrated to give 490 mg (86%) of crude compound 45 as a white foamy solid. Purification by silica gel chromatography (EtOAc) gave 276 mg (48%) of compound 45 as a white foamy solid.

IA/DABA/ATEG, Compound 46: A solution of 100 mg (0.1 mmol) of compound 45 was prepared, and 1 mL of trifluoroacetic acid was added, and the mixture was stirred for 1 h at room temperature. The mixture was concentrated under vacuum. The residue was triturated with Et_2O and dried under vacuum to give a white crystalline solid. The solid was dissolved in 1 mL of DMF, 104 uL (77 mg, 0.6 mmol) of diisopropylethylamine was added, the mixture was cooled to 0°, and 212 mg (0.6 mmol) of iodoacetic anhydride was added. The ice bath was removed, and the mixture was stirred at room temperature for 2 h. The mixture was cooled to 0°, acidified with 1 N H_2SO_4 , and partitioned between 10 mL of 1 N H_2SO_4 and 6 X20 mL portions of 8/2 $\text{CH}_2\text{Cl}_2/\text{MeOH}$. The combined organic layers were dried (MgSO_4), filtered, and concentrated to give 330 mg of orange oil. Purification by preparative HPLC (25 cm X 22.4 mm C_{18} , gradient: 30%B to 40%B 0-40 min, A = $\text{H}_2\text{O}/0.1\%$ TFA, B = $\text{CH}_3\text{CN}/0.1\%$ TFA, 12 mL/min) gave 19 mg of compound 46 as a white solid.



Example 22 Synthesis of the Tetraivalent Platform BA/PABA/DT/TEG, 51.

N-(t-butoxycarbonyl)PABA, compound 47: A solution was prepared of 3.0 g (21.9 mmol) of p-aminobenzoic acid in 60 mL of H₂O. Na₂CO₃ (2.16 g, 25.7 mmol) was added slowly followed by 30 mL of MeOH. When all solids were dissolved, a solution of 4.77 g (21.9 mmol) of di-t-butyl dicarbonate in 10 mL of MeOH was added and the mixture was stirred at room temperature for 18 h. To the mixture was added 4.92 g (25.6 mmol) of citric acid and the resulting cloudy mixture was partitioned between 200 mL of H₂O and 200 mL of EtOAc. The EtOAc layer was washed successively with 200 mL of 0.1 N HCl and 200 mL of H₂O, dried (Na₂SO₄), filtered, and concentrated to yield 3.0 g (58%) of compound 47 as a white solid.

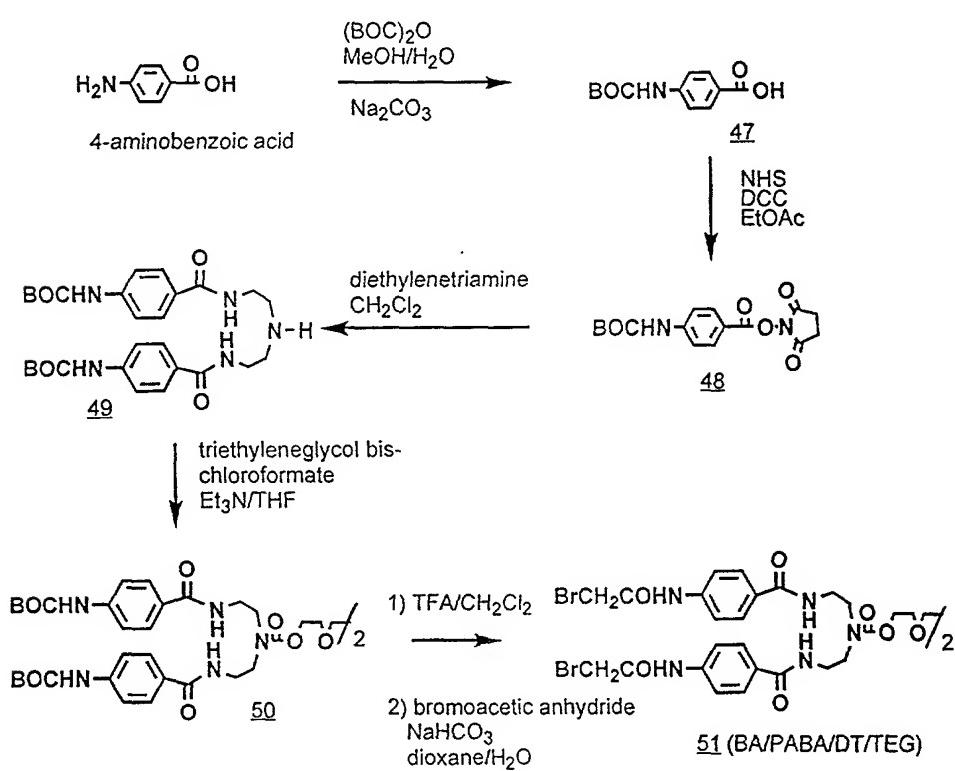
N-(t-butoxycarbonyl)PABA N-hydroxysuccinimidyl ester, compound 48: DCC (2.61 g, 12.6 mmol) was added to a 0° solution of 2.0 g (8.43 mmol) of compound 47 and 0.97 g (8.43 mmol) of N-hydroxysuccinimide in 50 mL of EtOAc. The ice bath was removed, the mixture was stirred for 16 h at room temperature, and 0.5 mL of acetic acid was added. The mixture was stirred for an additional 30 min, placed in the freezer for 1.5 h, filtered, and concentrated to give 3.75 g of crude 48. Purification by silica gel chromatography (50/50 hexane/EtOAc) gave 2.53 g (90%) of compound 48 as a white solid.

Compound 49: A solution of 3.00 g (8.97 mmol) of compound 48 in 50 mL of CH₂Cl₂ was added dropwise over 30 min to a solution of 485 uL (4.49 mmol) of diethylenetriamine in 30 uL of CH₂Cl₂ which had been cooled in an ice bath. The mixture was stirred at 5-7° for 30 min then at room temperature for 16 h. The milky mixture was placed in a separatory funnel with 200 mL of H₂O, the pH of the H₂O layer was adjusted to 10 with 3 M NaOH solution, and the mixture was extracted with 200 mL of 4/1 CH₃Cl/MeOH. The organic phase was dried (Na₂SO₄) and concentrated to give 0.971 g (40%) of compound 49 which was pure enough to be used in the following step. The aqueous layer was extracted with six 100 mL portions of 4/1 CH₃Cl/MeOH. The organic layers were combined, dried (Na₂SO₄), and concentrated to give another 1.08 g (44%) of compound 49 which required further purification. Further purification can be accomplished by silica gel chromatography (gradient, 80/20 to 70/30 CH₃Cl/MeOH).

Compound 50: A solution of 135 uL (0.66 mmol) of triethyleneglycol bis-chloroformate in 0.3 mL of THF was added to a 0° solution of 855 mg (1.58 mmol) of compound 49 and 275 uL (1.58 mmol) of diisopropylethylamine in 13 mL of THF. The cloudy mixture cleared when the ice bath was removed. An additional 70 uL of diisopropylethylamine was added to maintain a basic pH. The mixture was stirred at room temperature for a total of 3 h and partitioned between 25 mL of H₂O and 25 mL of EtOAc. The aqueous layer was extracted with a second 25 mL portion of EtOAc, and the combined EtOAc layers were dried (Na₂CO₃), filtered, and concentrated to give 0.986 g of crude 50. Purification by silica gel chromatography (80/4/16 CH₃Cl/dioxane/isopropanol) gave 516 mg (61%) of compound 50 as a white solid.

Preparation of BA/PABA/DT/TEG, compound 51: Compound 50 (487 mg, 8.79 mmol) was dissolved in 9 mL of CH₂Cl₂ and 5 mL of trifluoroacetic acid. The mixture was stirred for 1.5 h

and concentrated. The residue was triturated with 7 mL of Et₂O, and dissolved in 5 mL of MeOH and 1 mL of 48% HBr solution. The mixture was concentrated and placed under vacuum until a dry. The resulting HBr salt was dissolved in 10 mL of H₂O, and 191 mg (2.27 mmol) of NaHCO₃ was added. A solution of 591 mg (2.27 mmol) of bromoacetic anhydride in 10 mL of dioxane was added to the mixture. An additional 2 mL of dioxane was used to rinse. More NaHCO₃ was added as needed to maintain a basic pH. The mixture was stirred for 2 h at room temperature, and acidified with 1 N H₂SO₄. The mixture was extracted with 3 X 25 mL of EtOAc. The combined EtOAc layers were dried (Na₂SO₄), filtered, and concentrated to give 773 mg of an oil. Purification was accomplished by silica gel chromatography (gradient, 90/10 to 85/15 CH₃Cl/MeOH) to give 401 mg (77%) of 51 as a white solid.



Example 23 Synthesis of the Tetraivalent Platform BMP/TEG, 55.

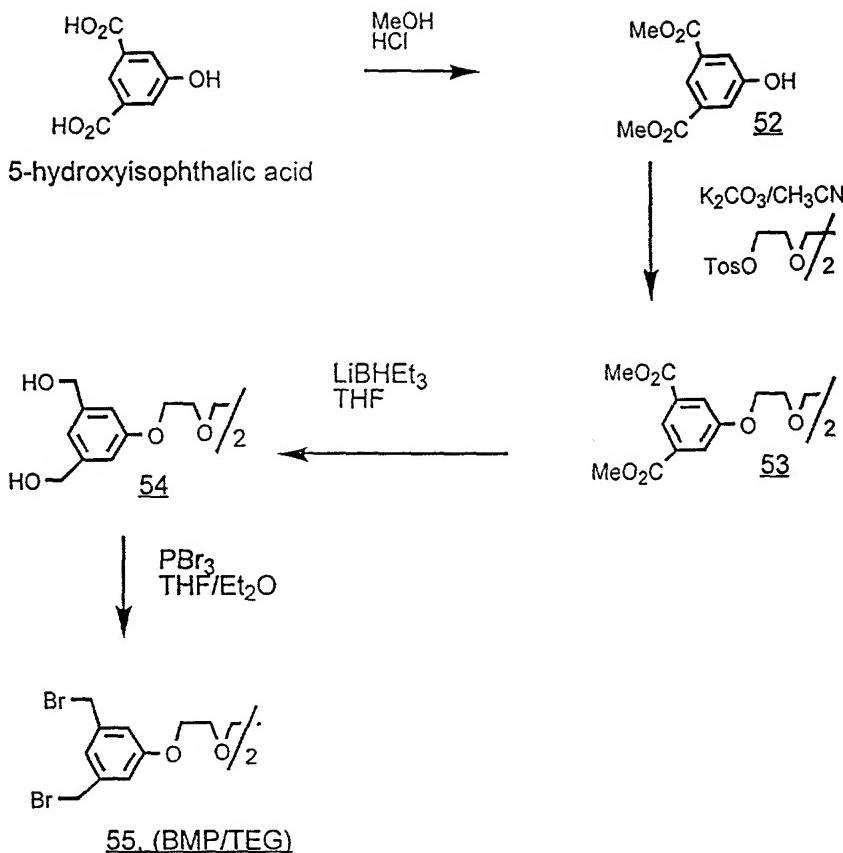
Dimethyl-5-hydroxyisophthalate, compound 52: A solution of 2.00 g (11 mmol) of 5-hydroxyisophthalic acid and 0.5 mL of con HCl in 30 mL of MeOH was refluxed for 5 h. The mixture was concentrated and the resulting residue was dissolved in 100 mL of EtOAc. The

EtOAc solution was washed successively with two 50 mL portions of 5% NaHCO₃ solution and two 50 mL portions of sat NaCl solution, dried (Na₂CO₃), filtered, and concentrated to give 2.09 g (90%) of 52 as a white crystalline solid.

Compound 53: Triethyleneglycol ditosylate 546 mg (1.19 mmol) was added to a suspension of 500 mg (2.38 mmol) of compound 52 and 395 mg (2.86 mmol) of K₂CO₃ in 11 mL of CH₃CN and the mixture was refluxed under N₂ for 16 h. The mixture was concentrated, and the residue was dissolved in 25 mL of CHCl₃ and washed with 25 mL of H₂O and 25 mL of sat NaCl solution. The CHCl₃ layer was dried (Na₂SO₄), filtered, and concentrated. Purification by silica gel chromatography (gradient, 50/50 hexane/EtOAc to 100% EtOAc) gave 93 mg (41%) of compound 53.

Compound 54: A suspension of 93 mg (0.18 mmol) of compound 53 in 4 mL of THF was stirred under a N₂ atmosphere while 1.82 mL (1.82 mmol) of a 1 M solution of LiBHEt₃ was added to the mixture dropwise. A clear solution was obtained which was stirred for 18 h at which time a 10% solution of HOAc in water was added until the pH was acidic. The mixture was concentrated under vacuum and the residue was dissolved in 10 mL of water. The mixture was extracted with 3 X 10 mL portions of EtOAc, and the combined EtOAc layers were dried (Na₂SO₄), filtered and concentrated. Purification was accomplished by silica gel chromatography (gradient, 90/10 to 85/15 CH₃Cl/MeOH) to give 20 mg (27%) of 54 as a white solid.

Compound 55: To a suspension of 20 mg (0.048 mmol) of 54 in 5 mL of Et₂O and 1 mL of THF is added 9.6 uL (0.1 mmol) of PBr₃. The mixture is stirred for 3 h and partitioned between water and EtOAc. The EtOAc layer is dried (Na₂SO₄), filtered and concentrated, and the residue is purified by silica gel chromatography (CH₃Cl/MeOH) to provide compound 55.



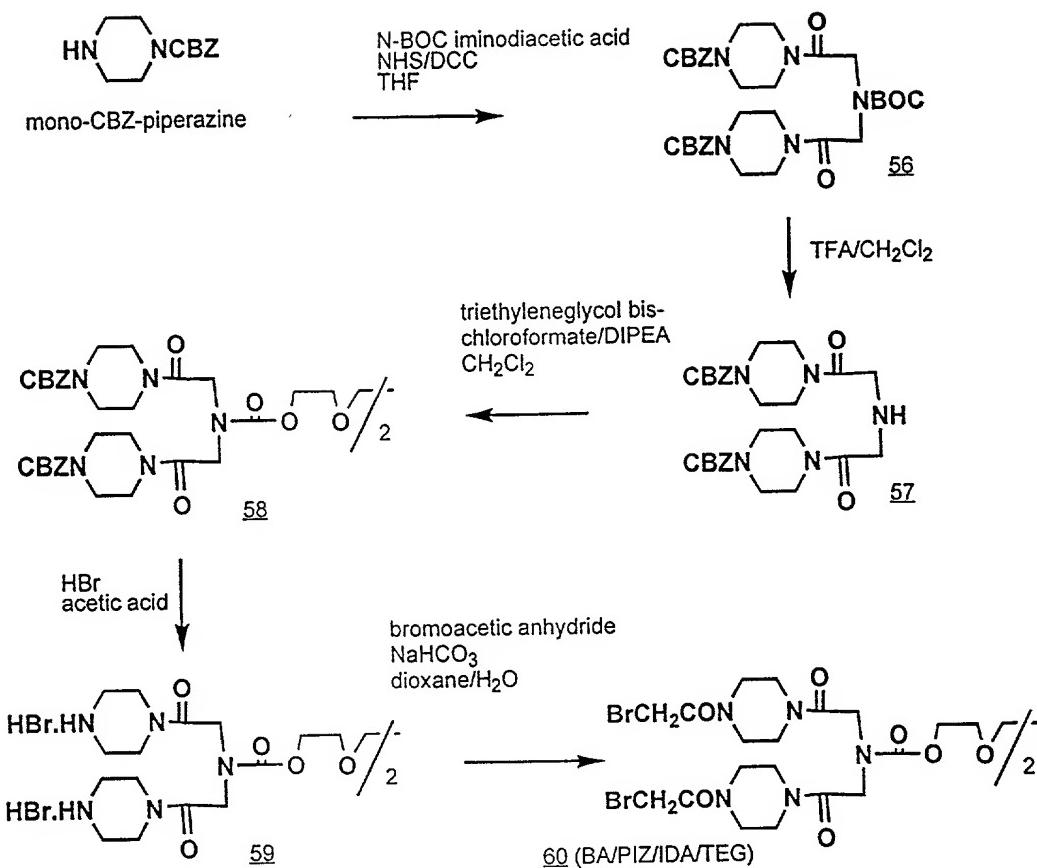
Example 24 Synthesis of the Tetravalent Platform BA/PIZ/IDA/TEG, 60.

Compound 56: To a solution of 1.02 g (4.37 mmol) of N-(t-butoxycarbonyl)-iminodiacetic acid (compound **5** in U.S. 5,552,391, Chemically-Defined Non-Polymeric Valency Platform Molecules and Conjugates Thereof) and 1.01 g (8.75 mmol) of N-hydroxysuccinimide in 50 mL of dry THF, cooled to 0°. was added 2.26 g (10.94 mmol) of dicyclohexylcarbodiimide. The mixture was stirred for 16 h allowing to slowly warm to room temperature, and a solution of 2.22 g (10.1 mmol) of mono-CBZ-piperazine in 25 mL of THF was added to the mixture followed by 1.22 mL (887 mg, 8.75 mmol) of Et₃N. The mixture was stirred for 7 h at room temperature, and filtered. The filtrate was concentrated and the residue was dissolved in 125 mL of EtOAc and shaken with 2 X 125 mL portions of 1 N HCl, 125 mL of sat NaHCO₃ solution, dried (MgSO₄), filtered, and concentrated to give 2.39 g of a sticky solid. Purification by silica gel chromatography (95/5 CH₂Cl₂/MeOH) gave 1.85 g (66%) of **56**.

Compound 57: To a solution of 1.74 g (2.74 mmol) of compound 56 in 10 mL of CH₂Cl₂ was added 10 mL of trifluoroacetic acid, and the mixture was stirred for 3 h at room temperature. The mixture was concentrated, and the residue was dissolved in 5 mL of CH₂Cl₂. The mixture was cooled to 0° and 100 mL of sat NaHCO₃ was added. The mixture was then extracted with four 100 mL portions of CH₂Cl₂. The CH₂Cl₂ layers were combined, dried (MgSO₄), filtered, and concentrated to give 1.46 g (99%) of 57 as a sticky hygroscopic solid which was used directly in the next step.

Compound 58: To a solution of 0.7 g (1.3 mmol) of compound 57 and 226 uL (168 mg, 1.30 mmol) of diisopropylethylamine at 0° was added a solution of 127 uL of triethyleneglycol bis-chloroformate in 4 mL of CH₂Cl₂, and the mixture was stirred for 3 h at room temperature. The mixture was partitioned between 80 mL of CH₂Cl₂ and 80 mL of 1 N HCl. The CH₂Cl₂ layer was washed with two 80 mL portions of water, dried (MgSO₄), filtered, and concentrated to give 736 mg (93%) of compound 58 as a crystalline solid.

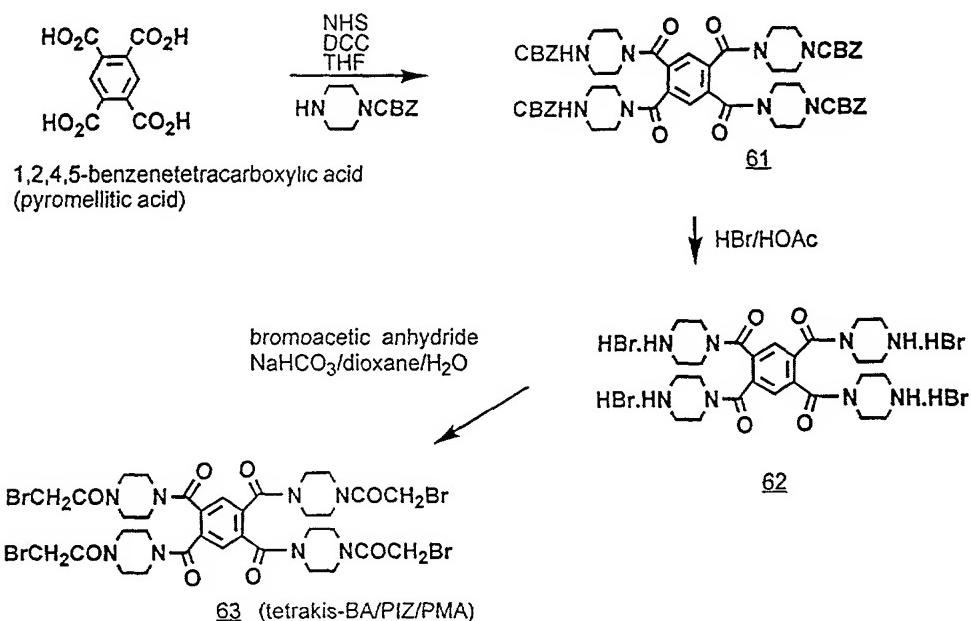
Compound 60: Compound 58 (61 mg, 0.48 mmol) was dissolved in 3 mL of 30% HBr/HOAc and the resulting mixture was stirred at room temperature for 1h at which time 5 mL of Et₂O was added. The mixture was placed in the freezer for 1 h and centrifuged. The resulting pellet was washed with Et₂O and dried to give the tetrahydrobromide salt 59 which was dissolved in 1 mL of H₂O. To the mixture is added 49 mg (0.58 mmol) of NaHCO₃ and 3 mL of dioxane. More NaHCO₃ is added, if needed, to make the mixture basic. The mixture is cooled to 0°, and 748 mg (2.89 mmol) of bromoacetic anhydride is added. The mixture is stirred for 2 h and partitioned between 20 mL of 1 N H₂SO₄ and 20 mL of 80/20 CH₂Cl₂/MeOH. The organic layer is dried (Na₂SO₄), filtered and concentrated to give crude 60 which is purified by silica gel chromatography (CH₂Cl₂/MeOH) to give 60.



Example 25 Synthesis of the Tetraivalent Platform tetrakis-BA/PIZ/PMA, 63.

Compound 61: To a 0° solution of 640 mg (2.52 mmol) of pyromellitic acid and 1.16 g (10.1 mmol) of N-hydroxysuccinimide in 50 mL of THF was added 2.6 g (12.6 mmol) of dicyclohexylcarbodiimide, and the mixture was allowed to come to room temperature while stirring for 16 h. A solution of 2.5 g (11.3 mmol) of mono-CBZ-piperazine in 25 mL of THF was added to the mixture followed by 1.4 mL (1.02g, 10.1 mmol) of Et₃N. The mixture was filtered, and the filtrate was concentrated. The residue was partitioned between 100 mL of EtOAc and 2 X 100 mL of 1 N HCl, and the EtOAc layer was washed with 100 mL of sat NaHCO₃, 100 mL of H₂O, 100 mL of sat NaHCO₃, dried (MgSO₄), filtered, and concentrated. Purification was accomplished by silica gel chromatography (97.5/2.5 CH₂Cl₂/MeOH) to give 1.78 g (66%) of 63 as a white crystalline solid.

Compound 63: Compound 61 is converted to compound 63 in essentially the same manner as described for the conversion of 58 to 60 in Example 23. Purification is accomplished using silica gel chromatography.



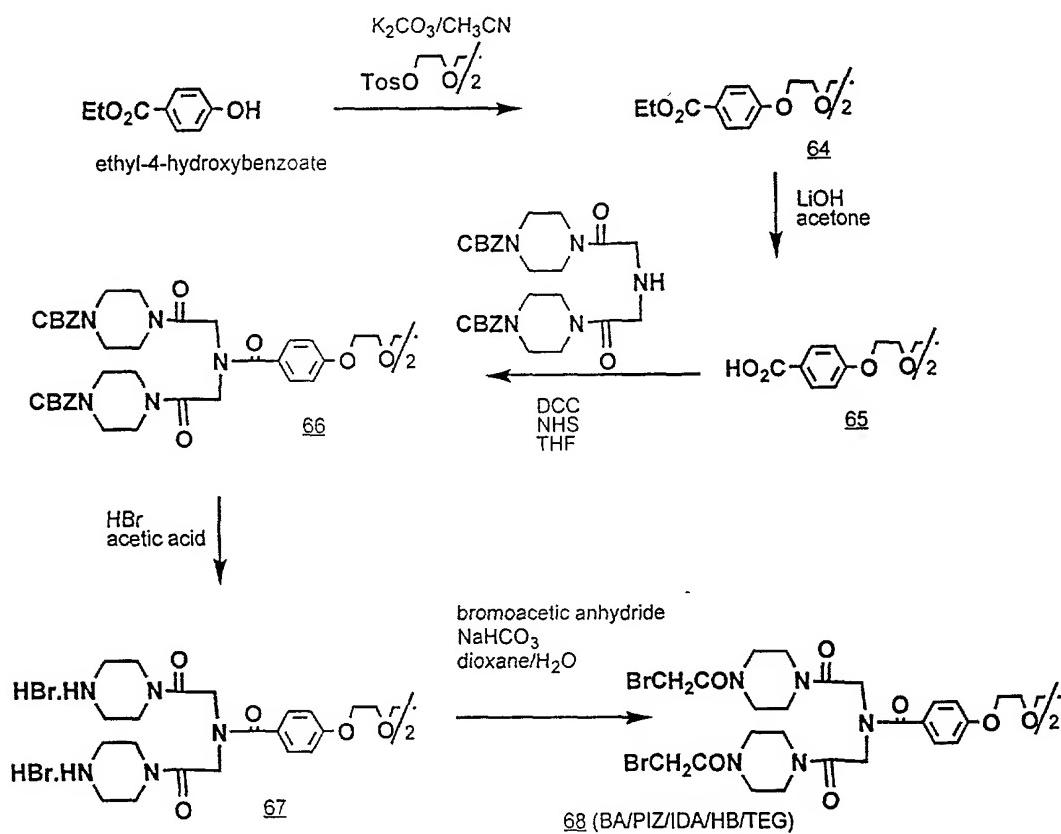
Example 26 Synthesis of the Tetravalent Platform BA/PIZ/IDA/HB/TEG, 68.

Compound 64: Triethyleneglycol ditosylate (1.0 g, 2.18 mmol) was added to a solution of 725 mg (4.36 mmol) of ethyl 4-hydroxybenzoate and 723 mg (5.23 mmol) of K_2CO_3 , and the mixture was refluxed for 16 h. The mixture was concentrated, and the residue was partitioned between 20 mL of water and 3 X 20 mL of Et_2O . The combined organic layers were washed with 2 X 40 mL of sat $NaHCO_3$ solution, 40 mL of sat $NaCl$ solution. The aqueous layers were washed with Et_2O , and the combined Et_2O layers were dried ($MgSO_4$), filtered, and concentrated. Purification by silica gel chromatography (70/30 hexanes/ $EtOAc$) have 902 mg (93%) of **64** as a crystalline solid.

Compound 65: Compound **64** is dissolved in acetone containing 2.2 equivalents of LiOH and the mixture is stirred for 3 h (until complete as evidenced by TLC). The mixture is acidified with acetic acid and concentrated, and the residue is purified by silica gel chromatography to give **65**.

Compound 66: Compound 66 is prepared similarly to the method of preparing compound 56 in example 23. Compound 65 is used instead of N-BOC-iminodiacetic acid, and compound 57 is used instead of mono-CBZ-piperazine. Purification is accomplished using silica gel chromatography.

Compound 68: Compound 66 is converted to compound 68 in essentially the same manner as described for the conversion of 58 to 60 in Example 23. Purification is accomplished using silica gel chromatography.

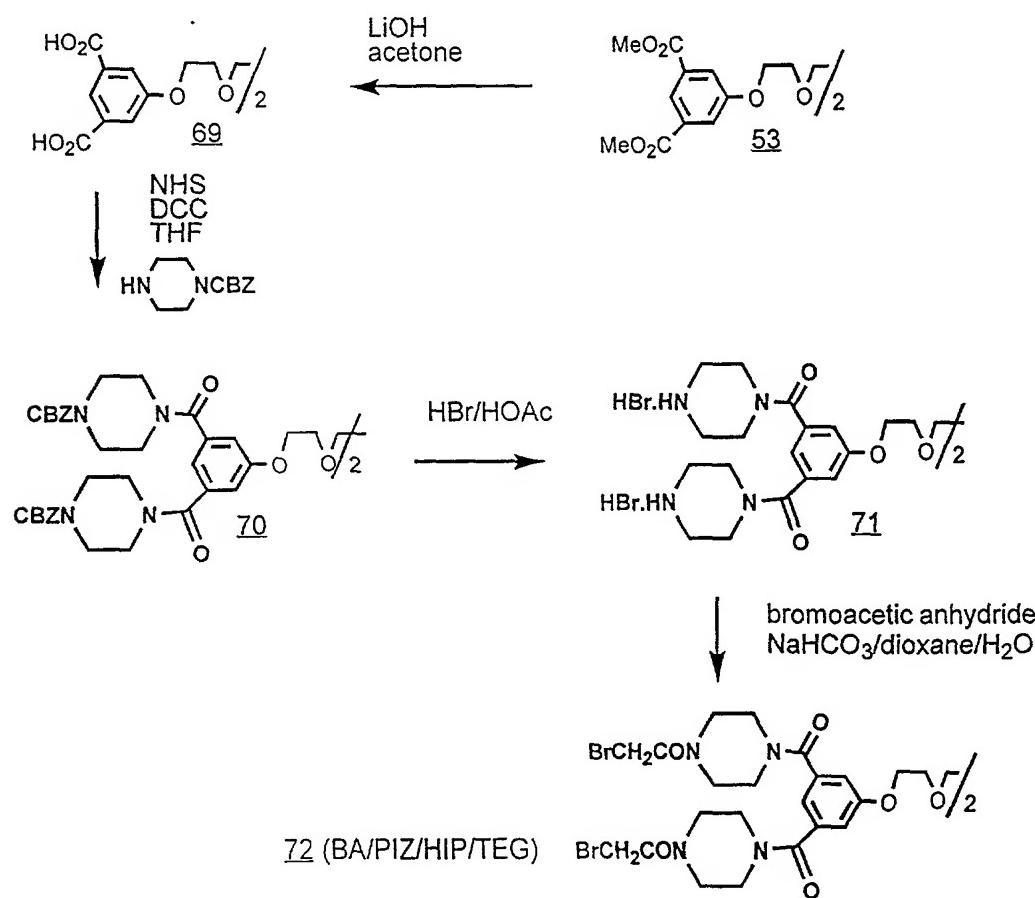


Example 27 Synthesis of the Tetravalent Platform BA/PIZ/HIP/TEG, 72.

Compound 69: Compound 53 is hydrolyzed with LiOH in essentially the same manner as described for the hydrolysis of 64 in example 25 with the exception that 4.4 equivalents of LiOH is used.

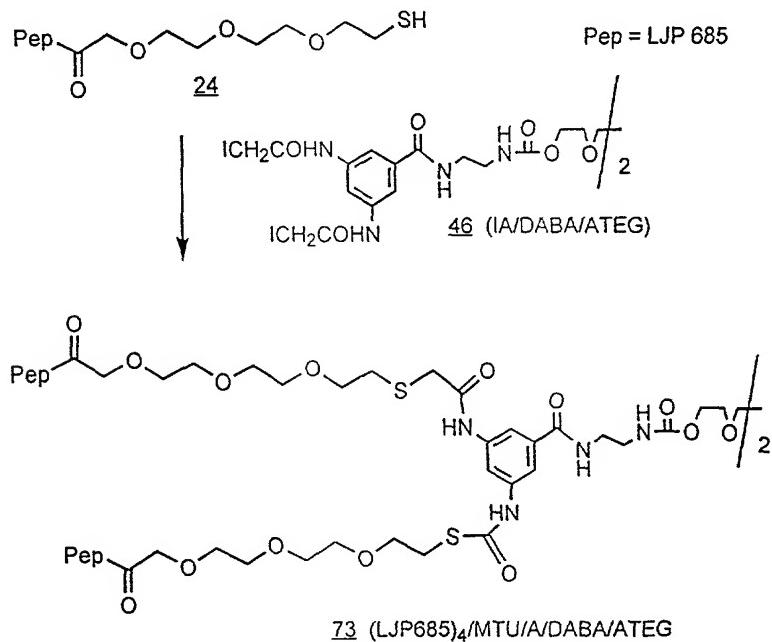
Compound 70: The tetra-acid, compound 69, is converted to compound 70 in essentially the same manner as described for the conversion of pyromellitic acid to 61 in example 24 with the exception that 69 is used instead of pyromellitic acid.

Compound 72: Compound 70 is converted to compound 72 in essentially the same manner as described for the conversion of 58 to 60 in Example 23. Purification is accomplished using silica gel chromatography.

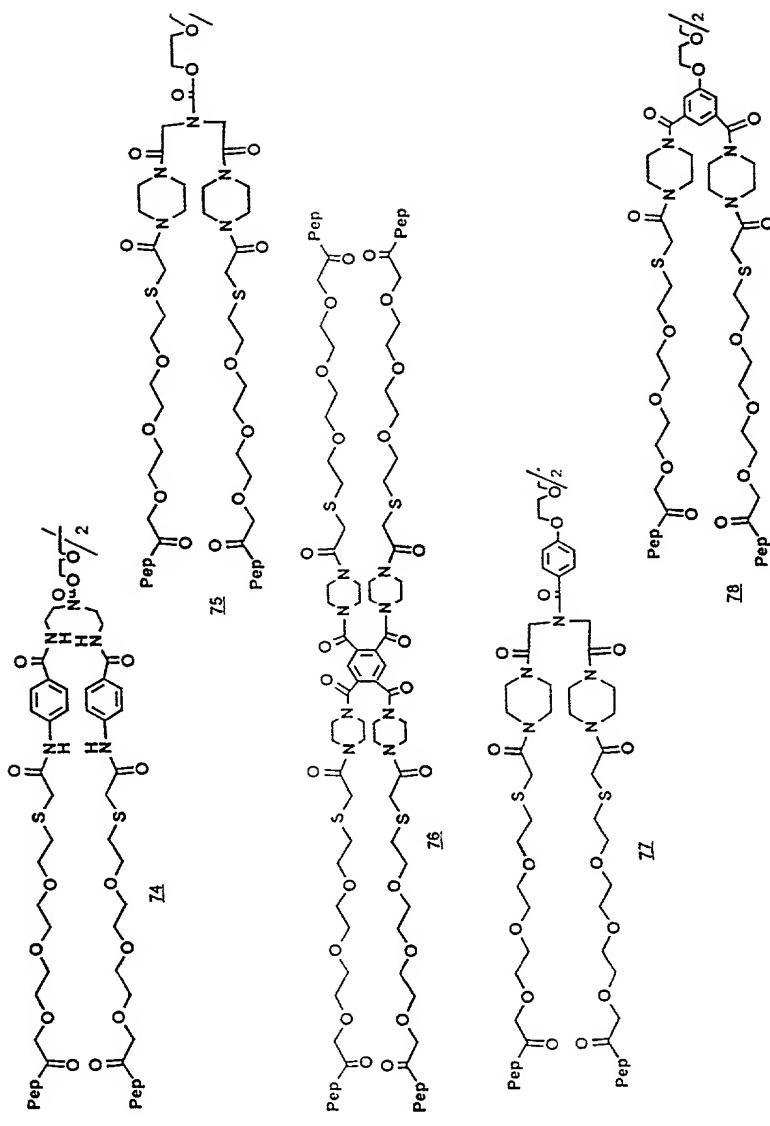


Example 28 Synthesis of Conjugates of Haloacetylated Tetravalent Compounds

Synthesis of $(\text{LJP685})_4/\text{MTU/A/DABA/ATEG}$ conjugate compound 73: A solution was prepared of 21 mg (15.5 μmol) of compound 24 in Helium sparged pH 8.5 200 mM borate buffer. To the solution was added a second solution consisting of 3.25 mg (2.6 μmol) of IA/DABA/ATEG, compound 46, dissolved in 396 μL of MeOH. A precipitate formed, and 1 mL of MeOH was added to dissolve all the solids. The mixture was stirred for 18 h at room temperature, and 6 mL of 9/1 $\text{H}_2\text{O}/\text{HOAc}$ was added. The mixture was diluted with 50 mL of 9/1 $\text{H}_2\text{O}/\text{CH}_3\text{CN}$ and loaded onto an HPLC preparative column. Purification was accomplished by preparative HPLC (25 cm X 22.4 mm C₁₈, gradient: 35% B to 45% B 0-40 min, A = $\text{H}_2\text{O}/0.1\%$ TFA, B = $\text{CH}_3\text{CN}/0.1\%$ TFA, 12 mL/min) to provide 10.8 mg (67%) of compound 73 as a white solid after lyophilization.

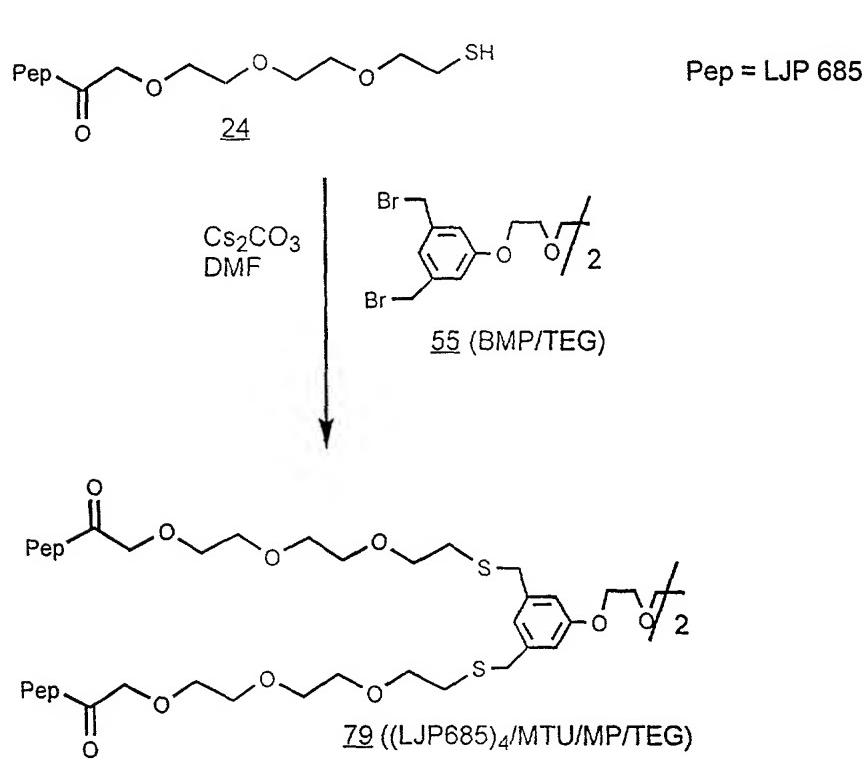


Synthesis of $(\text{LJP685})_4/\text{MTU}$ conjugates, compounds 74, 75, 76, 77, and 78: These conjugates are prepared from platform compounds 51, 60, 63, 68, and 72, respectively, by the same reaction conditions as described above for the synthesis of conjugate 73 by substituting the appropriate platform compound for platform compound 46. Pep can be LJP685 or other relevant peptide.



Example 29 Synthesis of Conjugates Tetravalent Platform 55, Conjugate Compound 79, (LJP685)₄/MP/TEG:

5 A solution is prepared of four equivalents of compound 24 and five equivalents of Cs₂CO₃ in DMF. One equivalent of BMP/TEG, platform compound 55, is added to the mixture which is then stirred for 1 h. The mixture is diluted with 80/20/10 H₂O/CH₃CN/HOAc and loaded onto a preparative HPLC column. Purification is accomplished by preparative HPLC (C₁₈, A = H₂O/0.1% TFA, B = CH₃CN/0.1% TFA) to give compound 79 after lyophilization. Pep can be LJP685 or other relevant peptide.

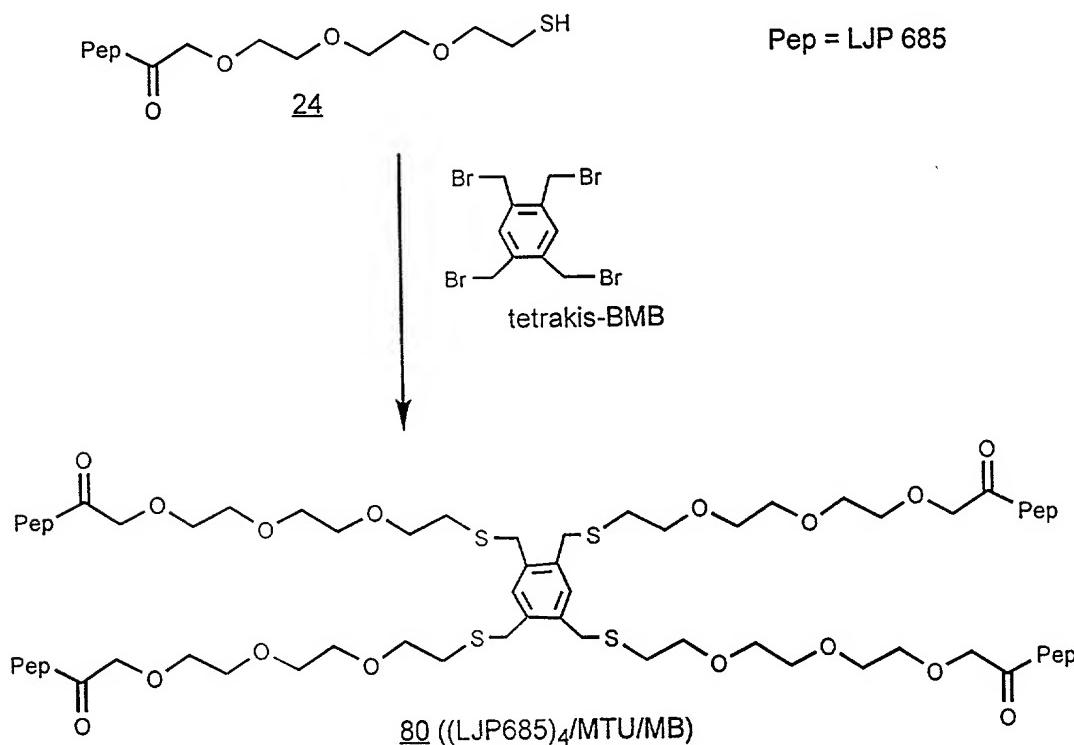


Example 30 Synthesis of Conjugates Tetraivalent Platform tetrakis-BMB, Conjugate Compound 80, $(\text{LJP685})_4/\text{MTU/tetrakis-MB}$:

A solution is prepared of four equivalents of compound **24** and five equivalents of Cs_2CO_3 in DMF. One equivalent of tetrakis-bromomethylbenzene is added to the mixture which is then

stirred for 1 h. The mixture is diluted with 80/20/10 H₂O/CH₃CN/HOAc and loaded onto a preparative HPLC column. Purification is accomplished by preparative HPLC (C₁₈, A = H₂O/0.1% TFA, B = CH₃CN/0.1% TFA) to give compound 80 after lyophilization. Pep can be LJP685 or other relevant peptide.

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Example 30 Fluorescence Polarization Peptide Binding Assays

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Synthesis of FITC-GPCILLARDRCG (CB2*)

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A solution of the cyclic disulfide peptide GPCILLARDRCG (20.0 mg, 14.4 μ mol) and fluorescein isothiocyanate (FITC) (5.6 mg, 14.4 μ mol) in 20 mL of ACN/water (1:1), containing 20 mg sodium carbonate (Na₂CO₃, pH ~ 10.5), was stirred at room temperature. The reaction was monitored by analytical HPLC. After consuming the fluorescent labeling reagent, the crude material was purified on a preparative HPLC eluted at 10 mL/min with a linear gradient from 30 to 55% B over 40 minutes where A was 0.1% (v/v) TFA in H₂O and B was 0.085% (v/v)

TFA in CAN. The FITC peptide was obtained as a bright yellow powder after lyophilization (3.7 mg, 15% yield): MS (ESI): m/e (M+1) Calcd. for C₇₃H₁₀₂N₁₉O₂₀S₃: 1661, obsd.: 1661.

5

Direct Binding Fluorescence Polarization Binding Assay (dbFP)

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The methodology is described in PanVera Application Guide (1994), PanVera Corporation. Briefly, a trace amount of fluorescein isothiocyanate (FITC) labeled peptide (CB2*-F) is titrated with antibody (ACA 6501 or 6701) and the polarization for the sample is plotted versus the antibody concentration. Polarization was measured with the PanVera Beacon instrument. Data were fitted to Equation 1.

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$$\text{Equation 1} \quad Y = \frac{[P_L * (K_D/R) + P_H]}{K_D/R + 1.0}$$

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where Y is the Y-axis value (milli-polarization units, mP), R is the total concentration of antibody receptor, P_L is the polarization for free FITC-labeled peptide (F), and P_H is the polarization for F complexed with R (FR). K_D is the dissociation constant (reciprocal binding constant) for F from the FR complex. For these equations to be valid, it must be true that F << R. This titration is shown in Figures 22 and 23 for CB2*-F binding to ACA-6701 and ACA-6501 antibodies, respectively. A complete titration was not obtained with ACA-6501 as shown in Figure 23, but a previous titration shown in Figure 24 gave a K_D of 241 nM. By adding CB2* (GPCILLARDRCG) in slight excess over antibody 6701, to displace CB2*-F from 6701 (see Figure 25), a dissociation rate constant of K_{off} = 0.0184 sec⁻¹, which corresponds to t_{1/2} = 38 seconds, was determined for CB2*-F. Given the K_D of 256 nM, this corresponds to an association rate constant of K_{on} = 3.6 X 10⁴ M⁻¹sec⁻¹ (after correcting for antibody bivalence). Thus, CB2*-F binding to ACA-6701 is limited only by diffusion of these two molecules together.

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Competitive Fluorescence Polarization Assay (cFP)

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The above described dbFP assay provides binding constants for FITC-labeled peptides and requires on the order of 0.5 mg of purified antibody. The cFP assay provides binding constants for peptides that lack the FITC group and it consumes less antibody, on the order of 10 μ g. The cFP assay is modified from that reported in PanVera Applications Guide (1994) PanVera Corporation such that it consumes 50-fold less antibody. Briefly, antibody (ACA 6701) is mixed with trace FITC labeled peptide (CB2*-F) and enough time is allowed for equilibrium to be reached. This was 1 hour for ACA 6701 and CB2*-F.

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Increasing concentrations of the unlabeled peptide being tested (CB2* or 3B10) are then added to the tube. After each addition, sufficient time, approximately 15 minutes, is allowed for equilibrium to be reached and the mP value was read. Although it is necessary to choose concentrations of 6701 ® and CB2*-F (F) such that $F \ll R$, the concentration of R need only be high enough such that the measured polarization (P_H) is significantly higher than P_L (see Figure 22). This $\Delta(mP)$ value should be 20 or more mP units to insure reliable results.

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$$\text{Equation 2} \quad \Delta(mP) = P_{H'} - P_L$$

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As unlabeled (no FITC) peptide (I) is added to inhibit F from binding to R, Y decreases from its maximum value of $P_{H'}$ to a plateau of P_L which should agree with that in Equation 1. These titrations are shown in Figures 26 and 27 for displacement of CB2*-F from ACA-6701 by CB2* and 3B10, respectively. The equation describing this titration was derived and is:

30

$$\text{Equation 3}$$

$$Y = \frac{P_{H'} + P_L * (I/K_I')}{I/K_I' + 1.0}$$

where P_L is the same as in Equation 1. I is the concentration of unlabeled peptide competitor, and K_I' is the *apparent* dissociation constant for that peptide. Values for these parameters were obtained by fitting cFP titration data to the above equation.

5 The true dissociation constant for I is obtained from Equation 4.

$$\text{Equation 4} \quad K_I = K_I' / (1.0 + R/K_D)$$

10 where R and K_D are defined as in Equation 1. The R/K_D ratio is obtained from the values of P_H' (from Equation 3) and P_H and P_L (from Equation 1) and using Equation 5.

$$\text{Equation 5} \quad R/K_D = (P_H - P_L) / (P_H - P_H')$$

15 In general, Equation 5 can be used to determine aPL antibody concentrations once the titration defined by Equation 1 is performed as a "standard curve." Thus, in addition to providing a means for determining K_I , this method provides a means of standardizing all aPL antibody stock solution concentrations and of analyzing their binding activity/stability over time using only 5-10 µg of antibody per cFP assay.

DISSOCIATION CONSTANTS DETERMINED BY dbFP AND cFP

Peptide	Antibody	Sequence	K_D^a or K_I
CB2*-F	6501	FITC-GPCILLARDRCG	482 nM ^a
CB2*-F	6701	FITC-GPCILLARDRCG	512 nM ^a
25 CB2*	6701	GPCILLARDRCG	35 nM
3B10	6701	AGPCLLAPDRCPG	313 nM

^a The K_D values have been multiplied by 2 to correct for K_D values determined from Equation 1 that do not factor in the bivalence of the antibody.

The results demonstrate that CB2*-F cross reacts with two very different aPL antibodies, ACA-6501 and ACA-6701, binding to both with equal high affinity. Removal of the FITC group improved binding of CB2* to ACA-6701 by 14-fold. Binding of a related peptide, 3B10, was 9-fold less than binding of CB2* to ACA-
5 6701. While this result may be due to additional framework residues on 3B10, it may also be due to the substitution of a proline for arginine at position 8 in CB2*. Previous NMR structure studies of 5A12, a peptide similar to 3B10, showed that this proline that is in a turn position gives the structure rigidity. CB2 is a much more flexible peptide and it has an arginine in this position. A more flexible peptide like
10 CB2 may be more cross-reactive because it may more readily adjust its shape to fit a given antibody binding site. The implications of drug rigidification on binding affinity are discussed in Koehler *et al.*, p. 251, GUIDEBOOK ON MOLECULAR MODELING IN DRUG DESIGN (Academic Press, N. Cohen, ed., 1996).

15 **Example 31 Tolerance Activity of Peptide Conjugates**

Two different conjugates containing the same peptide were tested for their ability to induce antigen specific tolerance *in vivo*. Briefly, mice were immunized with the peptide conjugated to the immunogenic carrier Keyhole Limpet Hemocyanin (KLH) to generate peptide-specific memory B cells. Three weeks later, groups of 5 mice per group were treated with various doses of the test conjugates, one group of mice was not treated and acted as the control. Five days later, all of the mice, including the control group, were boosted with the peptide conjugated to KLH and seven days later all of the mice were bled and their sera assayed for anti-peptide antibodies using a modified Farr assay. The Antigen Binding Capacity (ABC) was calculated for each individual serum sample according to the method described in G. M. Iverson, "Assay for *in vivo* adoptive immune response," Volume II, Chapter 67, HANDBOOK OF EXPERIMENTAL IMMUNOLOGY (Blackwell Scientific Publications, Weir *et al.*, eds., 4th ed., Oxford, 1986). These values were then used to determine a mean and standard deviation for all of the individuals of a group.
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While one of the conjugates induced tolerance, the other one did not inhibit anti-peptide antibodies over the dose range tested. The most likely explanation for this difference is that the latter conjugate has a short *in vivo* half-life. To address this problem, a system was employed that induced tolerance *in vitro*, thereby negating half-life consideration, and then the cells were transferred to irradiated recipients.

5 Briefly, spleen cells from mice primed with the peptide conjugated to KLH were harvested and incubated in complete RPMI-1640 medium for 2 hours at 37°C with various doses of the test conjugates. One group of cells was incubated without toleragen and acted as the positive control. The cells were washed, transferred to

10 irradiated syngeneic recipients and boosted with the peptide conjugated to KLH. Seven days later all of the mice were bled and their sera assayed for anti-peptide antibodies. The conjugate that did not induce any detectable tolerance when tested in the *in vivo* model did induce tolerance when tested in this *in vitro* model. This result supports the assumption that the difference between conjugates is due to a short half-life of the conjugate. To directly test this hypothesis, the conjugate was administered continuously by an implanted osmotic pump over a prolonged period of time. The results clearly show that this conjugate induced tolerance when administered by sustained release but not when administered as a bolus as shown in Figure 32.

15

20 Testing of (LJP685)₄/MTU-AHAB-TEG for Tolerance Activity in the *in vivo* Model

Mice were primed with LJP-685-KLH on alum plus *pertussis* as an adjuvant. Three weeks later, the mice were treated with a range of doses of the (LJP685)₄/MTU-AHAB-TEG conjugate. One group was not treated and acted as a control group. Five days later, all of the mice, including the control group, were boosted with 10 µg LJP685-KLH and seven days later the mice were bled. Their sera were analyzed for anti-LJP685 antibodies by a modified Farr assay as described above. The results as shown in Figure 28 demonstrate that the treatment with the (LJP685)₄/MTU-AHAB-TEG conjugate, over a dose range of 1 to 50 nmoles, had no detectable effect on the anti-LJP685 response.

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Testing the (LJP685)₄/MTU-DABA-TEG Conjugate for Tolerance Induction in the *in vivo* Model

Mice were primed with LJP685-KLH on alum plus *pertussis*. Three weeks later, the mice were treated with 5, 10 or 50 nmoles of the (LJP685)₄/MTU-DABA-TEG conjugate. One group was not treated and acted as a control group. Five days later, all of the mice, including the control group, were boosted with 10 µg of LJP685-KLH and seven days later the mice were bled. Their sera were analyzed for anti-LJP685 antibodies by a modified Farr assay. The results as shown in Figure 29 demonstrate that (LJP685)₄/MTU-DABA-TEG conjugate induces tolerance in the *in vivo* model with an ED₅₀ of 5 nmoles.

Testing the (LJP685)₄/MTU-AHAB-TEG Conjugate for Tolerance Induction in the *in vitro* Model

Spleen cells from mice primed 3 weeks earlier with LJP685-KLH were harvested and incubated in complete RPMI-1640 medium for 2 hours at 37°C with 4, 20 or 100 µM of (LJP685)₄/MTU-AHAB-TEG conjugate. One group of cells was incubated without toleragen and acted as a positive control group. The cells were washed, transferred to irradiated recipients and boosted with 10 µg of LJP685-KLH. Seven days later, the mice were bled and their sera were analyzed for anti-LJP685 antibodies by a modified Farr assay. The results as shown in Figure 30 clearly illustrate that the (LJP685)₄/MTU-AHAB-TEG conjugate can induce tolerance when tested in the *in vitro* model achieving an IC₅₀ of < 4 µM.

Testing the (LJP685)₄/MTU-DABA-TEG conjugate for Tolerance Induction in the *in vitro* Model

Spleen cells from mice primed 3 weeks earlier with LJP685-KLH were harvested and incubated in complete RPMI-1640 medium for 2 hours at 37°C with 0.4, 1.3 and 4 µM of (LJP685)₄/MTU-DABA-TEG conjugate. One group of cells was incubated without toleragen and acted as a positive control group. The cells were washed, transferred to irradiated recipients and boosted with 10 µg of LJP685-KLH.

Seven days later, the mice were bled and their sera were analyzed for anti-LJP685 antibodies by a modified Farr assay. The results as shown in Figure 31 demonstrate that (LJP685)₄/MTU-DABA-TEG conjugate can induce tolerance when tested in the *in vitro* model, achieving an IC₅₀ of < 4 μM.

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Testing the (LJP685)₄/MTU-AHAB-TEG Conjugate for Tolerance Induction *in vivo* Using a Continuous Delivery Pump

Mice were primed with LJP685-KLH on alum plus *pertussis*. Three weeks later, the mice were divided into 5 groups of five mice per group. On day 1, one group was treated with a bolus of saline and another group was treated with a bolus containing 50 nMoles of the (LJP685)₄/MTU-AHAB-TEG conjugate. The three remaining groups were implanted with osmotic pumps. In one group, the pumps were filled with saline and delivered at 1 μL/hour for 3 days. The two remaining groups received pumps filled with the (LJP685)₄/MTU-AHAB-TEG conjugate (50 nMoles). One group received pumps that deliver at 1 μL/hour for three days and the other received pumps that deliver at 0.5 μL/hour for seven days. On day 5, the pumps that deliver for three days were surgically removed. On day 7, all of the mice, including the control group, were boosted with 10 μg of LJP685-KLH. On day 10, the pumps that deliver for seven days were surgically removed. On day 14, all of the mice were bled. Their sera were analyzed for anti-LJP685 antibodies by a modified Farr assay. The results are shown in Figure 32.

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Testing the (LJP-Peptide)₄/MTU-BMP-TEG Conjugate for Tolerance Induction in the *in vivo* Model

Mice are primed with peptide-KLH on alum plus *pertussis*. Three weeks later, the mice are treated with the (LJP-peptide)₄/MTU-BMP-TEG conjugate, one group is not treated and acts as the control group. Five days later, all of the mice, including the control group, are boosted with 10 μg of peptide-KLH and seven days later the mice are bled. Their sera are analyzed for anti-peptide antibodies by a modified Farr assay.

The results show that the (LJP-peptide)₄/MTU-BMP-TEG conjugate induces

tolerance in the *in vivo* model at a potency equal to or greater than that of the (LJP685)₄/MTU-AHAB-TEG conjugate.